

# **Aptamers for Detection and Diagnostics (ADD): Can mobile systems linked to biosensors support molecular diagnostics of SARS-CoV-2? Should molecular medicine explore multiple alternatives as adjuvants to or replacement for traditional and non-traditional vaccines?**

Shoumen Palit Austin Datta<sup>1,2,3</sup>

<sup>1</sup> MIT Auto-ID Labs, Department of Mechanical Engineering, Massachusetts Institute of Technology, Room 35-206, 77 Massachusetts Avenue, Cambridge, MA 02139, USA ([shoumen@mit.edu](mailto:shoumen@mit.edu))

<sup>2</sup> Medical Device (MDPnP) Interoperability and Cybersecurity Labs, Biomedical Engineering Program, Department of Anesthesiology, Massachusetts General Hospital, Harvard Medical School, 65 Landsdowne Street, Suite 232, Cambridge, MA 02139, USA ([sdatta8@mgh.harvard.edu](mailto:sdatta8@mgh.harvard.edu))

<sup>3</sup> NSF Center for Robots and Sensors for Human Well-Being (RoSeHuB), Collaborative Robotics Lab, School of Engineering Technology, Purdue University, 193 Knoy Hall, West Lafayette, IN 47907, USA

*This article is dedicated to the memory of*

# *Bernard Lown*

*June 7, 1921 – February 16, 2021*

Friend, mentor, inventor, physician, scientist, cardiologist, statesman, humanist and grandfather to Ariel Lown Lewiton.



Dr Bernard Lown with Dr H el ene Langevin Joliot-Curie, granddaughter to Marie Curie and Pierre Curie (middle) and Mrs Louise Lown (1946-2019).

Photograph by Shoumen Datta. March 01, 1997. Bedford, Massachusetts.

<https://www.nytimes.com/2021/02/16/health/bernard-lown-dead.html>

<https://www.aa.com.tr/en/life/nobel-peace-prize-winner-bernard-lown-dies-at-99/2147924>

## DISCUSSION

ABSTRACT	4
1. ADD for SARS-CoV-2	4
2. Aptamers for Detection and Diagnostics (ADD)	5
3. ADD Digital Data Design	6
4. Beyond ADD	9
5. Prevention follows Detection and Diagnostics	11
6. Alternative Arrays	13
7. Array of Targets	15
8. Discussion	22
9. Complexity of the biomedical scenario, socio-economic catastrophe and public health crisis	24
10. Post-Pandemic Health & Healthcare: Broad Spectrum use of Sensors with Smartphones	29
11. Comments	34
APPENDIX I	37
Components for SARS-CoV-2 ADD Decision System and Data Templates	
APPENDIX II	44
APTAMER-AS-A-DRUG (AAAD) FOR THE LESS AFFLUENT UNDERSERVED WORLD?	
APPENDIX III	47
THE MOLECULAR BASIS OF DISEASE	
APPENDIX IV	54
APTAMERS as ADJUVANTS and/or ALTERNATIVE to ANTIBODIES	
APPENDIX V	58
ENGINEERING RNASE RESISTANT RNA APTAMERS	
APPENDIX VI	58
SCIENCE, SOCIETY VS SENSITIVITY, FREQUENCY OF TESTING AND COST	
APPENDIX VII	62
ROOM FOR <i>IN SILICO</i> CREATIVITY IN PREDICTING APTAMER DESIGN?	
APPENDIX VIII	67
FIELD TESTING THE APTAMER SENSOR, ALONE, MAY NOT BE PRUDENT	
APPENDIX IX	71
IS INNATE IMMUNITY TOO FAR FETCHED EVEN TO BE A RATIONAL IDEA?	
APPENDIX X	73
REFOCUS ON DETECTION: LOW-COST SENSORS FOR THE UNDERSERVED	
INCONCLUSIVE CONCLUSION	77
WHERE CAN WE GO FROM HERE?	83
ACKNOWLEDGEMENTS	89
REFERENCES	90
Cartoon for Reference #372	154

**Aptamers for Detection and Diagnostics (ADD): Can mobile systems linked to biosensors support molecular diagnostics of SARS-CoV-2? Should molecular medicine explore multiple alternatives as adjuvants to or replacement for traditional and non-traditional vaccines?**

“CITCOM” precedes this proposal. PDF is in the MIT Library <https://dspace.mit.edu/handle/1721.1/128017>

**ABSTRACT**

Engineering a biomedical device as a low-cost, non-invasive, detection, and diagnostic platform for surveillance of infections in humans, and animals. The system embraces the IoT “*digital by design*” metaphor by incorporating elements of connectivity, data sharing and (secure) information arbitrage. Using an array of aptamers to bind viral targets may help in detection, diagnostics, and potentially prevention in case of SARS-CoV-2. ADD may spawn a broader approach from ADD diagnostics to AAAD (aptamer-as-a-drug) therapeutics. CoVID-19 as an airborne endemic infectious disease (EID) may be analogous to a social IED (improvised explosive device) which may profoundly transform geo-political economy. Desperate demand for alternatives to traditional vaccines may be partially met by lower cost molecules for diagnostics and other types of vaccine alternatives for therapeutics, globally.

**1. ADD for SARS-CoV-2**

The scale of mortality and morbidity due to SARS-CoV-2 evokes us to explore unconventional approaches to mitigate the risks presented by pandemics. Scientists may be less aware of the discovery of aptamers thirty years ago but the “fit” of aptamers with respect to the molecular biology of the current problem makes it worthwhile to propose new tools. Innovation may arise from the combination of chemistry and molecular biology with sensor engineering and opportunity for data dissemination<sup>1</sup> to benefit public health<sup>2</sup> by integrating the principle<sup>3</sup> of internet of things<sup>4</sup> (IoT<sup>5</sup> as a design metaphor).

	Emergence	Cases	Fatality Rate	Transmissibility
SARS	2003	8,098	11%	+
MERS	2011	2,519	34%	+
SARS-CoV-2	2019	~ 250 million	0.5-1% Est	+++
SARS-CoV-3?	??	??	??	??
SARS-CoV-4?	??	??	??	??

Table I: Is the eight<sup>6</sup> year interval between SARS, MERS and COVID-19 just an unrelated coincidence?

4 • ADD is in the MIT Library <https://dspace.mit.edu/handle/1721.1/128017> and <https://dspace.mit.edu/handle/1721.1/145774>



## 2. Aptamers for Detection and Diagnostics (ADD)

Single stranded (ss) DNA aptamers (ssRNA<sup>7</sup> are equally useful<sup>8</sup> but susceptible to degradation by ribonuclease) bind with **specificity** to SARS-CoV-2<sup>9</sup> proteins (Nucleocapsid, Spike, Nsp1). Aptamers are conjugated with carbon or cadmium quantum (QT) nano-dots. If there are viruses (1, 10, 100, 1000) in a sample (sputum, saliva) at a detectable level, then it triggers QT.DNA (QTD) conjugated complex to transmit optical property change (EIS or electrochemical impedance spectroscopy is another option for signal transduction). An optical signal transduction mechanism may offer low cost data acquisition, enabling billions of people to use ADD (detection tool) *at home or anywhere* (**AHA**). The end-user must have access to the "QTD" conjugate (distributed by health departments in hamlets, towns and cities). QTD (product) may be a slurry in a tube labeled as "CoV-2-DETECTION & DIAGNOSIS" (C2DD). It remains to be investigated if inclusion of endo-b-N-acetylglucosaminidases (ENGases<sup>10</sup>) in the slurry may be necessary to expose the binding sites by partially removing the N-glycan coat if the viral Spike protein is the target (Figure 20). Imagine **C2DD** as a tube of lip balm or similar form factor. For supply chain and logistics, it will reduce operational cost of distribution if C2DD may be shipped as a tamper-proof sterile vial without the need for cold supply chain or special storage to extend shelf-life.

**First**, end-user uses her **smartphone holo-lens "QTD" app** (not limited to Microsoft HoloLens, the concept can be re-developed anywhere to reduce cost) to take an image of the C2DD vial/tube *without* sample (no virus). Priming (tuning) step is **critical** to establish a baseline for signal transduction and app-embedded data analytics engine to set the system to "without virus" ground state to obtain an optical "ground zero" (baseline will be different for EIS). Open question for instrumentation is the need for UV activation (for traditional nanodots) to record the shift (valence electron transfer). Can the app be configured to *perform the activation* and record the photoluminescence change? Using visible light to activate and coupling activation/quenching with the app needs innovative chemical/device engineering.

**Second**, the end-user spits (or adds a small volume saliva or sputum using a swab/spoon) in the test tube (vial). *There is room for controversy in this step but it is the easiest non-invasive procedure.*

**Third**, end-user uses her smartphone holo-lens "QTD" **app** to record optical change (as soon as possible after adding saliva/sputum). Perhaps similar to bar code or EPC or QR code scanning.

**Fourth**, end-user uses her smartphone holo-lens "QTD" **app** to record optical changes every 5 minutes for 30 min (from the time of adding the sample). There will be questions about ENGase activity, binding kinetics of the aptamer, signal to noise ratio ([filtering algorithms (Kalman<sup>11</sup> filter), error correction], activation/quenching issues, damping of signals due to interference from host proteins, salinity and **pH** of mucus-mucin/saliva/sputum sample (any or all could jeopardize binding and signal).

### 3. ADD Digital Data Design

Baseline versus change over time will appear as a plot in the app (analytics, Figure 1, uses basic machine learning (ML) tools, for example, SVM or support vector machine). Fool-proof visualization by generating a "traffic signal" visual [green oval (NO virus detected); red oval (virus detected); yellow oval (inconclusive/ambiguous)]. Data gathered by the smartphone app (if enabled by user) to be transmitted to national centers of epidemiology (eg CDC in USA, ECDC in EU) and local hospitals (the choice will be user-dependent). Allowing collection of anonymized data may be one alternative (without recording IPv6/IPv4 addresses) but pros/cons to be considered for the greater good, public safety and privacy<sup>12</sup>.

This app is a "frontline" detection tool which may be used **everyday** or each week, At Home or Anywhere (AHA), by individual users. The "C2DD" vial has no therapeutic value. Positive results (red oval - virus detected) may have to be re-confirmed using lab tests (PCR, mAbs) in a clinic or hospital. **C2DD PRODUCT and associated SERVICE "QTD" app if combined**, are *data-informed tools*. It does not offer or guarantee further testing or treatment. Distribution and pricing of the hypothetical C2DD product and proposed pay-per-use (PAPPU<sup>13</sup>) service for QTD will be debated by corporations. Free distribution of C2DD and a **micro-payment** model (pay-per-use) for the "QTD" app is *advocated*.

Users may hide or selectively control data/information sharing as well as access to surveillance data (data from daily screening for infection by the infectious agent in question). Secure sharing of surveillance data by users (**citizen science**) is recommended to generate a robust and representative status of the community or infected demographics in the region in terms of **molecular epidemiology**.

In general, data from molecular epidemiology is critical for resource-constrained healthcare supply chains to optimize planning (humanitarian logistics), allocate human resources (medical professionals) and organize transportation of materials to areas where assistance is needed. Citizen science<sup>14</sup> efforts are germane for the efficacy of healthcare systems in case of widespread infections (epidemics/pandemics). The tools which makes citizen science possible and effective may be viewed as global public goods. Similar systems for **animal surveillance** (farms, cattle, poultry, meat) are necessary to reduce infection in domestic animals (pets) and from crippling the food supply chain.

Components of the ADD system (QTD, C2DD) including mobile data collection, information arbitrage and public health applications are not limited to SARS-CoV-2 but is a **platform approach** which includes digital design elements illustrated<sup>15</sup> in Figure 1. Citizen science supported public health may immensely benefit from detection of viruses, bacteria, fungi, prions or *any infections agent* as long as an aptamer (oligonucleotide based on the idea<sup>16</sup> of an "anti-sense" approach<sup>17</sup>) may bind a small molecule or a macromolecule (peptides or proteins) with sufficient specificity, sensitivity and selectivity to generate credible data which may be *distributed* in real-time to inform and initiate subsequent steps. Scoring data from test sample, negative and positive control (for same person) will improve accuracy.

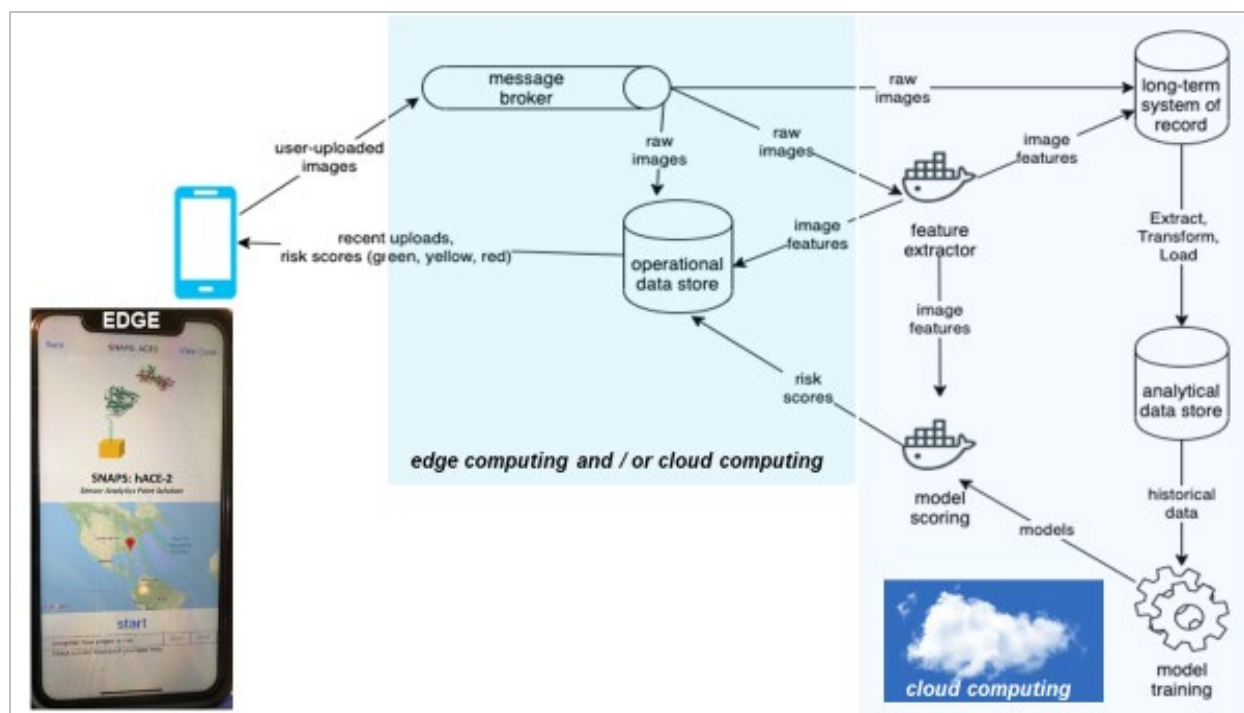


Figure 1: ADD system also includes data acquisition, analytics and data distribution which includes elements of the concept referred to as internet of things (IoT) which is a “digital<sup>18</sup> by design” metaphor. Cartoon shows the potential path of raw data from the hypothetical binding between a sensor and a target<sup>19</sup> molecule. Raw data from signal transduction due to binding activity is transmitted and acquired at the “edge” by the smartphone. The raw data is “processed” using tools either at the edge (embedded operations in the smartphone) or data may be uploaded to the cloud. Post-cloud computing analytics is returned to the edge device for display within an ADD application portal on the smartphone. The choice between edge versus cloud computing is a function of infrastructure (availability of wireless bandwidth, at the edge). The user may observe a difference in the time that it takes to process the data and display information (delayed visualization due to latency, function of bandwidth and speed).

Data scoring and processing is recommended due to variability of systems. ADD proposes the use of aptamers but other alternative arrays (see section 6) which may use the general approach (above) may “weigh” the information based on probability of false positive / false negative outcomes from tests<sup>20</sup> (separate from false positives / false negatives in machine learning<sup>21</sup> models). Assigning weighted risk to data and running other analytics can be performed on the mobile device (smartphone) or in the cloud, depending on access to and quality of telecommunications infrastructure. Cloud computing could add latency<sup>22</sup> between upload and display of information or prediction, depending on availability, reliability and connectivity to the internet. Several regions of the world still lack sufficient access to the internet<sup>23</sup>.

Scoring, processing and assigning risk within the analytical engine may benefit from machine learning (ML) tools to create a set of models or rules, to be described by and agreed on by experts. The system may scan and screen the image or data from the holo lens app (optical signal) to compare with these models or rules. Assigning an “image risk score” (IRS) may influence the presentation of the raw data where the “traffic signal” “red” may be provided with a sub-text containing a confidence score or include a qualitative comment (*likely* presence of virus) associated with a Likert-type<sup>24</sup> indicator/scale.

In any procedure, enabling the IRS to influence the raw data must be stringently controlled. Models or rules must be agreed by global experts whose credibility may be above question. Hence, these models and rules must stay outside the realm of testing services or labs or groups that are involved with creating systems, for example, ADD. It is preferable if model scoring (assigning risk score to an upload) runs on a platform which is not influenced by the local operator or the mobile user. The smartphone uploading the data may use a “tool” that applies the IRS engine residing in a secure infrastructure in a public cloud (FDA, NIH, NSF, CDC, ECDC) using appreciable level of cybersecurity (eg. Microsoft Azure, Amazon AWS). The smartphone must have the permission and physical availability to wireless internet or mobile data network to remotely access analytical tools in the cloud, such as, the IRS engine.

One alternative is to install (and update) the data scoring models/rules (IRS engine) in the ODS (operational data store, see Figure 1). The ‘message broker’ receives uploads and sends them to the ODS, which serves “hot” data to the app. ODS database is tuned for rapid reads, and serves requests made by the mobile app (only recent uploads and metadata about those uploads, including “risk scores”).

Risk scores are generated from models which are trained from historical data relevant to the test in question (using aptamer or antibody or other molecules, for example, hACE2). There must be access to sufficient statistical data from each type of test to create a credible risk score. If the model is based on bad data (garbage in), the risk score and IRS engine will spew bad information (garbage out). The model’s responsibility to assign “risk score” impacts the “traffic signal” and could alter the outcome. Model training<sup>25</sup> requires vast quantities of historical data, curated and pooled across multiple users who used the test and *verified* their outcome. If the binding was positive it must be corroborated by PCR<sup>26</sup> or another test with even higher specificity to confirm the result from the binding test using ADD tool.

Model-building is an iterative exercise that requires lab data from testing to be evaluated by credible scientists before data scientists can use it (curate?) to train ML models, which are error prone<sup>27</sup>. In model scoring, a model (in the IRS engine) is called to act on the uploaded (input) data. This analysis generates a prediction, displayed on the smartphone as information or recommendation for the user. The outcome the user views depends on the *design choices* made in ML model<sup>28</sup> training. It is absolutely central that model scoring requires “*features*” (characteristics germane to model/analysis). Creating features<sup>29</sup> is the task of a *team* of specialists (scientists collaborating with data experts). Harvesting

feature vectors and data relevant to the feature is the task of feature extractors. It may be provided by humans or we may use automated<sup>30</sup> feature<sup>31</sup> selection/extraction<sup>32</sup> to generate features from raw data.

#### 4. Beyond ADD

Scientific and engineering challenges to design ADD must embrace trans-disciplinary activities. But, no new physics is necessary. ADD may be available to **billions**, as a low-cost mobile AHA (at-home) *product* linked to IoT-type *service* app. The user experience is related to the service, not the product. The convergence of hardware and software with science and engineering as well as analytics and machine learning to *meaningfully* ascend the DIKW pyramid (data, information, knowledge, wisdom) is key to creating any detection **platform** where other tools and devices may upload data using open data APIs and standards-compliant data interoperability (DDS<sup>33</sup>) tools to aggregate or explore cumulative analytics, integrated with other systems, for example, geographic information systems<sup>34</sup> or GIS.

In the broader spectrum, ADD is an embryonic element of a potential *global health surveillance platform* (GSP) which may be pivotal as an early warning signal for humans and animal farms. Lessons from tsunami detection are sorely missing from public health policy discussions. Implementation of GSPs are neither a part of any local public health strategy nor on the agenda of precision population health management organizations (CDC, ECDC, WHO).

An important element of the *global health surveillance platform* (GSP) may include data from non-invasive profiling, referred to as “*pay-per-pee*” healthcare, which may be instrumental in molecular profiling for longitudinal studies on health and wellness<sup>35</sup>. GSPs may try not to dwell on genomics<sup>36</sup> (DNA) and expression<sup>37</sup> (RNA<sup>38</sup>) in imprecision<sup>39</sup> medicine but include proteomics because gene expression is insufficient unless the *functions* are *implemented* by proteins. Aptamers<sup>40</sup> in proteomic profiling (GWAS<sup>41</sup>, metabolomics) and other applications<sup>42</sup> including ADD may benefit from synergistic integration to help predict status of health (collected papers<sup>43</sup> provide select applications of aptamer).

Genomics is a “snapshot” (static structure of the infrastructure) and transcriptomics (RNA, GTEX) is an indicator of expression, which is data, but data may not (always) contain information. Proteins bind<sup>44</sup> in a myriad of ways<sup>45</sup> and translates *data to usable information* to maintain standard dynamic operating procedures (physiology, homeostasis, metabolomics).

Proteomics is a “time series” but its analysis over time may be interrupted due to feasibility and logistics of implementing programs like *pay-per-pee* healthcare, not to mention the complexity involved in extracting sense, often cryptic, from thousands of protein profiles, *over time*. Static protein profiles using NMR and mass spec<sup>46</sup> tools only capture *snapshots*. Can proteomics make sense<sup>47</sup> of a cytokine storm as markers of counter-anti-inflammatory response<sup>48</sup> even before the infectious agent is detected? Perhaps it is utopian to expect proteomic profiling as a daily practice in healthcare and home-health.



## Pay-Per-Pee Home Health IoT Wireless Toilet Bowl Connected to Health IT



A woman spits into a tube so that her saliva can be tested for the presence of the novel coronavirus.

COVID-19

### Spit shines for easier coronavirus testing

Tests using saliva are cheaper and faster than those with nasal swabs—and can be just as accurate

Figure 2: (Top) *Pay-per-pee* healthcare may provide time series data for precision medicine. (Left) “Collection of saliva samples by patients themselves negates the need for direct interaction between health care workers and patients. This interaction is a source of major testing bottlenecks and presents a risk of nosocomial infection. Collection of saliva samples by patients themselves also alleviates demands for supplies of swabs and personal protective equipment. Given the growing need for testing, our findings<sup>49</sup> provide support for the potential of saliva specimens in the diagnosis of SARS-CoV-2 infection.”

## 5. Prevention follows Detection and Diagnostics

If viewed<sup>50</sup> as non-classical antibodies<sup>51</sup> then the role of aptamers vastly exceeds that of detection. It spills over into prevention, perhaps as an alt-vaccine, albeit non-immunogenic. Identifying aptamers that can detect viral proteins in saliva implies that the aptamers may also bind the same protein (albeit with altered kinetics<sup>52</sup>) if administered topically (nasal spray, throat spray, soft-mist inhaler). Protecting the naso-pharyngeal area by saturating it with aptamers which binds (irreversibly?) to proteins from respiratory viruses (SARS) may be a preventative measure. Asymptomatic<sup>53</sup>, pauci-symptomatic and COVID-19 patients clearly expressing symptoms associated with SARS-CoV-2 may continue application of the aptamer cocktail to reduce the spread of infection by disabling (?) nascent virions. Aptamers preventing the spike protein (S1 RBD) of SARS virion from *attaching* to the ACE-2<sup>54</sup> viral receptor protein of uninfected cells may slow down the infection and development of COVID-19.

It follows that aptamers can also bind to any or all viral proteins not only in the extracellular space but also *inside* the cell. Delivering a portfolio of functional aptamers inside the cytosol must face the challenges posed by bio-availability and toxicity due to the potential for perturbing functions of essential<sup>55</sup> cellular proteins. Creating aptamers as *alt-vaccines* for *any* infecting organism (virus, bacteria, fungi, prion) which uses a protein in its lifecycle may be an (~30 year) old idea. Will the use of aptamers gain greater prominence in global public health practices, as a low-cost *global public goods tool* to contain the current and future epidemics and/or pandemics, worldwide, in humans and animals?

Single stranded RNA or ssDNA aptamers are not linear “tapes” but 3-dimensional *shapes* as illustrated by the discovery of tRNA<sup>56</sup> by Paul Zamecnik, Mary Louise Stephenson and colleagues at MGH, HMS. Publication of the discovery of tRNA by Zamecnik in 1958 catalyzed an array of milestones including the discovery of mRNA by Brenner<sup>57</sup> and Gros<sup>58</sup> as well as the *lac operon* model of feedback inhibition by Jacob and Monod<sup>59</sup>, all three published in 1961. The role of proteins in regulation<sup>60</sup> emerged as central to physiology and metabolism. In transcription, translation and replication<sup>61</sup> the binding between proteins and nucleic acids acted as a “switch” (mechanism of action). The notion<sup>62</sup> of aptamers<sup>63</sup> germinated<sup>64</sup> in 1990 but it drew on knowledge from binding between oligonucleotides and proteins. Aptamers may be 20-60<sup>65</sup> oligonucleotides or more. Binding specificity<sup>66</sup> of an enriched pool may be orders of magnitude different ( $K_d$ ) between a nearest neighbor or an analog. Sequential steps<sup>67</sup> are necessary from a starting sample (for example,  $9 \times 10^{14}$  ssDNA oligonucleotides) to arrive at an enriched pool of aptamers (19 ssDNA aptamers). The process has evolved<sup>68</sup> in complexity<sup>69</sup> and unique structures may be involved<sup>70</sup> in conferring specificity. In many applications<sup>71</sup> of aptamers<sup>72</sup> the debate also involves issues pertaining to trust and doubts<sup>73</sup> due to the constant demand for increasing accuracy and precision with respect to sensitivity, selectivity and specificity, in detection and diagnostics.



Current and future<sup>74</sup> application<sup>75</sup> of aptamers include chemistry<sup>76</sup>, chemotherapy<sup>77</sup>, food<sup>78</sup> safety, diagnostics<sup>79</sup>, antibodies<sup>80</sup>, alt-vaccines<sup>81</sup>, imaging<sup>82</sup> and different<sup>83</sup> types<sup>84</sup> of biosensors<sup>85</sup>. ADD as a detection tool for SARS-CoV-2 proposes aptamer-based sensors (aptasensors) to detect SARS-CoV-2 proteins. When an aptamer binds with the target, the signal (data) will be transduced and captured by a mobile device. Analytical tools will process data and display information on smartphones (Fig 1). Data dissemination will follow according to user preferences, to inform public health authorities or hospitals.

Optimism for aptamers as detection tools<sup>86</sup> extend to SARS-CoV-2 due to the detection of SARS-CoV (etiologic agent of 2008 SARS epidemic) C-terminal of N (nucleocapsid) protein at a concentration as low as 2 picograms/mL using a RNA<sup>87</sup> aptamer in a nanoarray. Tests using saliva<sup>88</sup> may be unsuitable for RNA<sup>89</sup> aptamers due to presence of ribonuclease<sup>90</sup> (RNase). DNA aptamers previously shown to bind to the N protein of SARS-CoV ( $K_d$  4.93±0.3nM<sup>91</sup>) also<sup>92</sup> binds to the N protein of SARS-CoV-2. The N protein<sup>93</sup> of SARS-CoV-2 shares 91% sequence homology with the N protein<sup>94</sup> of SARS-CoV but is less similar (16% - 38%) with N protein from the other 5 known human coronaviruses. Thus, detection<sup>95</sup> of N protein in saliva using an aptamer-based ADD aptasensor is *possible*. Aptamer-based technologies<sup>96</sup> directed toward SARS-CoV-2 Spike protein are gaining<sup>97</sup> momentum<sup>98</sup>. Blocking<sup>99</sup> the S protein from attaching to hACE-2 may perturb viral entry and prevent<sup>100</sup> the spread of infection. Aptamers created against the S1 RBD<sup>101</sup> may block binding to hACE-2 (internally) or serve as a detection tool (external ADD aptasensor) to test saliva/sputum for SARS-CoV-2. Other<sup>102</sup> SARS-CoV-2 targets<sup>103</sup> including Nsp1<sup>104</sup> may be less accessible in saliva because they are synthesized after viral entry. But, during the burst cycle, when new virions are released, viral proteins inside the host cell may be exposed. The targets are not limited to external viral proteins (spike, nucleocapsid, envelope proteins; Figure 4).

*Signal transduction* and *data acquisition* follows detection. In addition to EIS (electrochemical impedance spectroscopy<sup>105</sup>) signals, optical signals are preferred because data acquisition using cameras and apps in smartphones are feasible in locations where resources may be limiting. Protein<sup>106</sup> detection<sup>107</sup> by conjugating aptamers with quantum dots<sup>108</sup> is a tried<sup>109</sup> and true<sup>110</sup> process<sup>111</sup> which may be the optical signal (data) for this *system*. Changes in optical characteristics due to binding may be captured by cameras on mobile phones or HoloLens<sup>112</sup> app in smartphones may scan the saliva sample (think barcode or QR<sup>113</sup> code scan). Cameras (sensors) associated with the holo-lens (Kinect<sup>114</sup>) can scan the “field” and collects data to create a digital geometry<sup>115</sup> (digital model, 3D image). For ADD, HoloLens tools required for holographic functions<sup>116</sup> may be unnecessary, for example, accelerometer (speed of movement), gyroscope (tilt, orientation) and magnetometer (compass). Optical data captured from saliva containing testing vials will be analyzed (machine learning tools; see Figure 1) followed by visualization of information on the mobile device and (secure) information arbitrage, if authorized.

## 6. Alternative Arrays

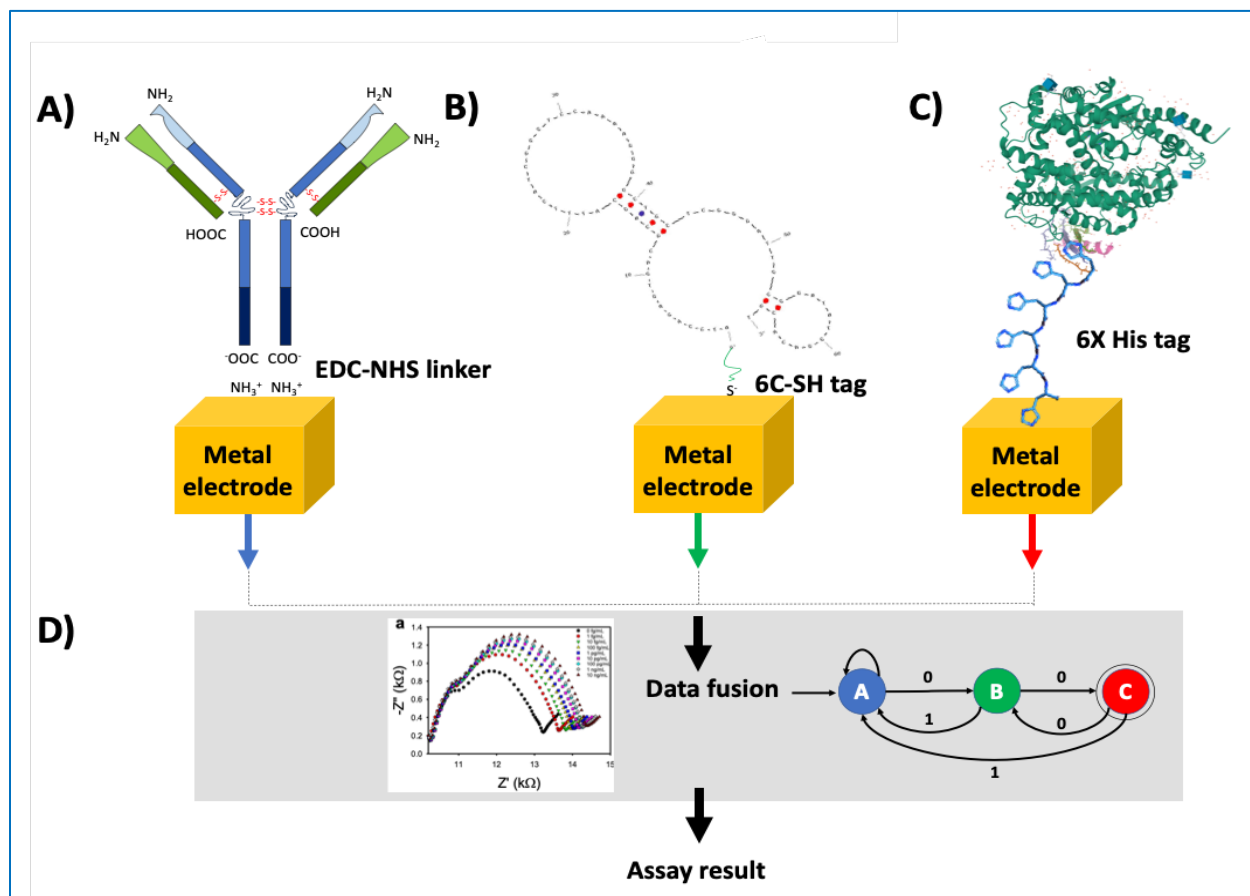


Figure 3: **Multiplexed<sup>117</sup> Detection Tool for SARS-CoV-2.** Upper panel presents potential recognition and detection chemistries. The data (fusion, middle panel) will be analyzed and assay results displayed (bottom). The data and information will be displayed on a mobile device (see cartoon in Figure 1). Three distinct binding targets for SARS-CoV-2 spike protein RBD are presented in sections A, B (ADD) and C. In (A) RBD-antibody (SARS-CoV-2 strain specificity) is functionalized with EDC-NHS chemistry to metal (gold, Au) nanoparticles (or may be attached/adsorbed on laser inscribed graphene, LIG). In (B) single-stranded DNA aptamers with thiol linker is adsorbed to metallized LIG (ADD aptasensor). In (C) histidine-tagged human ACE2 is adsorbed to metallized LIG. (D) Binding elicits signal (EIS, impedance spectroscopy) which is transduced to a mobile device. Data acquisition is followed by “hot” data upload to embedded tiny database (tinyDB<sup>118</sup>) in the device (ODS in Figure 1). Analytics may be executed on the device (embedded logic, machine learning tools) or uploaded to cloud server. The data fusion (model scoring) step may be necessary to make sense of the data, *in combination*, to provide not only raw data (results from A, B and C) but information, extracted from data and processed according to a simple SNAPS<sup>119</sup> paradigm to convey the *meaning of the outcome*, to inform the non-expert end-user.

Interpretation of data may be necessary due to the caveats of target binding and recognition. The specificity of the antibody used in the tool may not bind or bind with lower affinity ( $K_d$ ) with viral target protein (Spike protein) due to mutations in the epitope which generated the immunoglobulin (IgG). Lack of binding or lower affinity of binding can interfere with signal generation and failure to log signal over noise. Thus, individuals carrying SARS-CoV-2 may fail to test positive (false negative) if the viral variant possesses mutations preventing the antibody (A) to bind with the mutated Spike protein. Other factors (temperature, pH, salinity) may also interfere with signal (see “model scoring” in Appendix).

In (B), binding with the aptamer is highly specific but it depends on precisely which oligonucleotide (sequence of the ssDNA from an enriched pool) binds to which part of the Spike protein. For ADD, one aptamer may bind to the RBD (receptor binding domain) of the SARS-CoV-2 Spike protein. The length of the RBD (primary sequence) used in screening and enriching for the aptamer(s) may influence the shape (structure) of the RBD during selection phase. The complementarity of the shape of the RBD and the secondary/tertiary structure of the ssDNA *complex* is key to the binding specificity and affinity. If the test sample contains the whole Spike protein (includes RBD) as well as fragments (peptides with different lengths of amino acid sequence) which may or may not contain the RBD then the binding to the aptamer may fluctuate (widely) because the primary sequence of the protein may influence the secondary and tertiary structural outcome. The latter may change the configuration of the RBD in a given fragment and prevent binding to the aptamer, generating a false negative. If a sample contains other proteins and peptides, it is possible that the 3D configuration of an arbitrary protein or protein fragment could mimic or compete, albeit partially, with the RBD, and elicit a signal by binding with the aptamer, even if the binding is ephemeral due to reduced affinity (false positive result).

Binding of the Spike protein RBD to the immobilized hACE2 protein target (C) is probably the weakest link in this tripartite approach. Presence of mutations, dynamic or modified configuration and the effect of the environment (temperature, pH, salinity) may perturb binding and corrupt the signal.

Error correction and data curation may be necessary to prevent data corruption (false negative, false positive, limit of detection) to improve the information and recommendation for end-users. If the confidence in the raw data from each element is high, then the data may be responsibly combined (*after data scoring, image risk score*) to display the information with an assigned degree of confidence which may be more than the sum of the parts (positive, negative, false positive, false negative). The strategy from data acquisition and display vs information and recommendation must reduce risk, optimize level of precision and accuracy to maximize the value of the information for the user and/or the community. Of greater concern is the *accumulation* of errors, which when aggregated (time series data from ADD used as a surveillance tool), may generate spurious results with respect to the status of the population.

## 7. Array of Targets

The ADD approach for detection of infectious agents is based on targets identified from the biology and/or lifecycle of the organism and its interaction with the host (humans, animals). The RBD (receptor binding domain) of the Spike protein from SARS-CoV-2 and the human ACE2 cellular receptor (in bats, rats, pangolins and related animals in the phylogenetic tree; reviewed in reference 9) are under intense scrutiny. But, exploring the biology of SARS-CoV-2 reveals other equally potent targets. Developing drugs, antibodies and aptamers may benefit from a brief review of the viral biology. For SARS-CoV-2 detection alone, there are at least two other external proteins which may serve as targets for binding to aptamers, the M protein and the E protein in addition to S protein (Figure 4).

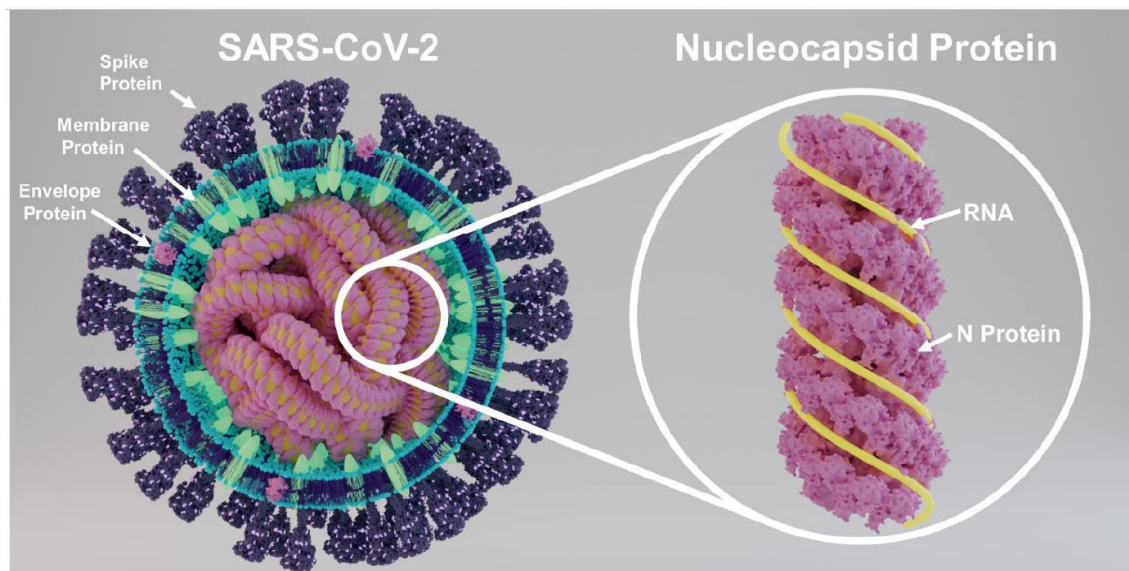


Figure 4: ssRNA genome of SARS-CoV-2 is longer compared to other RNA viruses (HIV, Influenza, Zika, Ebola; see Figure 16). It is encased in a nucleocapsid core (N protein<sup>120</sup>) and resides inside the virus. External surface of the virus is studded with S (spike), M (membrane) and E (envelope) proteins.

The receptor binding domain (RBD) of the Spike protein appears to make the first contact with the human cellular receptor ACE2 (angiotensin converting enzyme 2). Disrupting this event is the Holy Grail for preventing the virus from entering the cell. The mechanism by which Spike protein facilitates viral entry is not merely due to the recognition (between RBD and ACE2) but a cascade of events that begins after successful binding. The events that follow result in *fusion* of the viral envelope with the cell membrane, thereby allowing the viral genetic material (+ssRNA) to be delivered inside the cell in order to create progeny viruses. *Fusion* is mediated by the *fusion machinery* and *fusion peptide* sub-segments of Spike S2 protein which includes a step resembling a “jack-in-the-box” toy<sup>121</sup>. These segments of the Spike protein are *better conserved* and occupy a distinctly different part of the Spike protein (Figure 5).

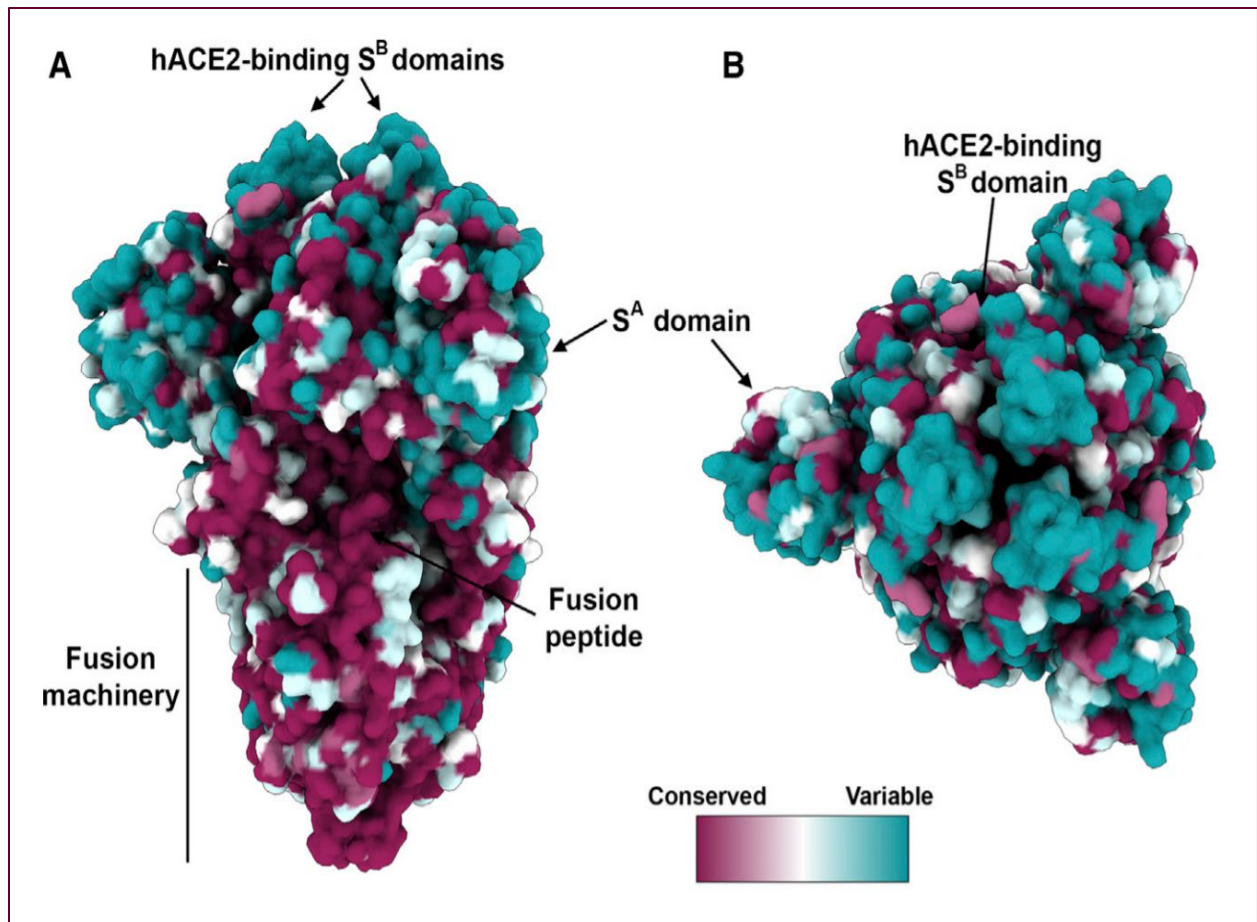


Figure 5: Sequence conservation<sup>122</sup> of sarbecovirus S glycoproteins plotted on the SARS-CoV-2 Spike protein structure [viewed from the side (A) and top (B)]. The receptor binding domain (RBD S1) is separate from the region of the Spike S2 protein necessary to initiate viral entry. The latter is better conserved (fusion machinery, fusion peptide) and perhaps better targets for ADD aptasensors.

The better conserved segment of the Spike protein may offer valuable epitopes<sup>123</sup> and potential binding sites for aptamers (unless glycan moieties interfere). In addition to the RBD (which appears to be more variable), the conserved portions of the S2 subunit responsible for fusion (fusion machinery, fusion peptide) are likely targets for aptamer binding. It remains to be seen if reagents (monoclonal antibodies, aptamers) aimed at the fusion specific domain of the S protein can disrupt viral entry and serve as tools for detection *as well as* prevention.

Interfering<sup>124</sup> with the human cellular proteins ACE2 and TMPRSS2 (which are viral targets) to prevent viral binding may not be prudent. Reagents directed against proteases, usually non-specific, may perturb physiological functions essential for homeostasis. The events which follow after the viral Spike protein docks with the human ACE2 protein are illustrated (Fig 6 copied from Scientific American<sup>125</sup>).



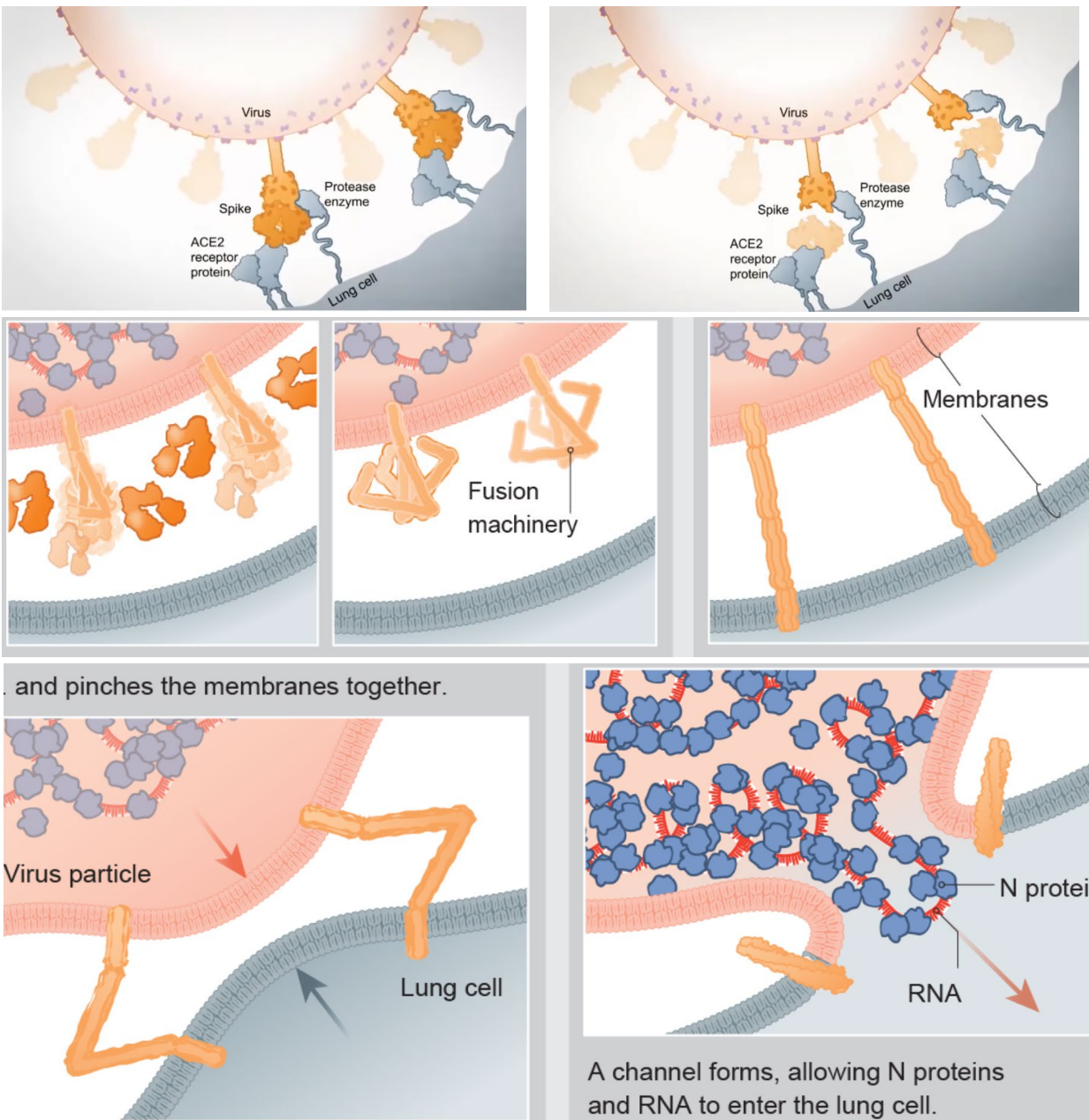


Figure 6: Cascade of events<sup>126</sup> leading to viral entry into host lung cell identifies the “*jack-in-the-box*” mechanism as a pivotal tool used by the fusion machinery of the Spike protein to deliver the viral RNA inside the host cell. Selectively disabling the fusion machinery of the Spike protein is an attractive target for aptamers and other reagents. If available, the latter may not only detect and diagnose but prevent infection, even if virus particles may have already reached the human apical surface<sup>127</sup> area. Superior region of the lungs are more vulnerable to infection due to higher number of hACE2 receptors. The number of hACE2 decreases from superior to inferior. Lower part of the lungs have less ACE2 and TMPRSS2 proteins, corroborated by the observation that these genes are expressed at a higher level in upper nasal epithelial tissue compared with bronchial and small airway epithelial brushings<sup>128</sup>.

The +ssRNA of SARS-CoV-2 (positive strand serves as mRNA) generates at least 27 or more viral proteins by creatively manipulating the host translational machinery. Theoretically, any or all viral proteins could serve as targets for anti-viral<sup>129</sup> strategies. Virus-encoded proteases<sup>130</sup> are distinct<sup>131</sup> from cellular proteases and may serve as good<sup>132</sup> targets. The viral protease<sup>133</sup> 3-chymotrypsin-like protease<sup>134</sup> or 3CLpro<sup>135</sup> aka M<sup>pro</sup> is encoded by Nsp5 and appears<sup>136</sup> to cleave (see “scissors” in Figure 7) essential viral proteins from “polyproteins” generated from translation of open reading frame (ORF) 1a and 1b (Fig 7). Papain-like protease<sup>137</sup> PLpro (ORF 1a, Nsp3<sup>138</sup>), cleaves<sup>139</sup> proteinaceous post-translational (ref 131) modifications on host proteins to evade host anti-viral immune responses. Nsp1<sup>140</sup> suppresses host translation by cleaving cell mRNAs<sup>141</sup> and competes<sup>142</sup> with mRNAs for binding to human 40S ribosomal mRNA channel<sup>143</sup> (as well as 43S, 80S subunits). Type 1<sup>144</sup> interferon<sup>145</sup> (IFN-1) response<sup>146</sup> is modulated by Nsp1, Nsp 6 and Nsp13, which interferes indirectly with IFN-1 by suppressing the phosphorylation and/or nuclear translocation of other cellular molecules<sup>147</sup> involved in catalyzing the IFN-1 response.

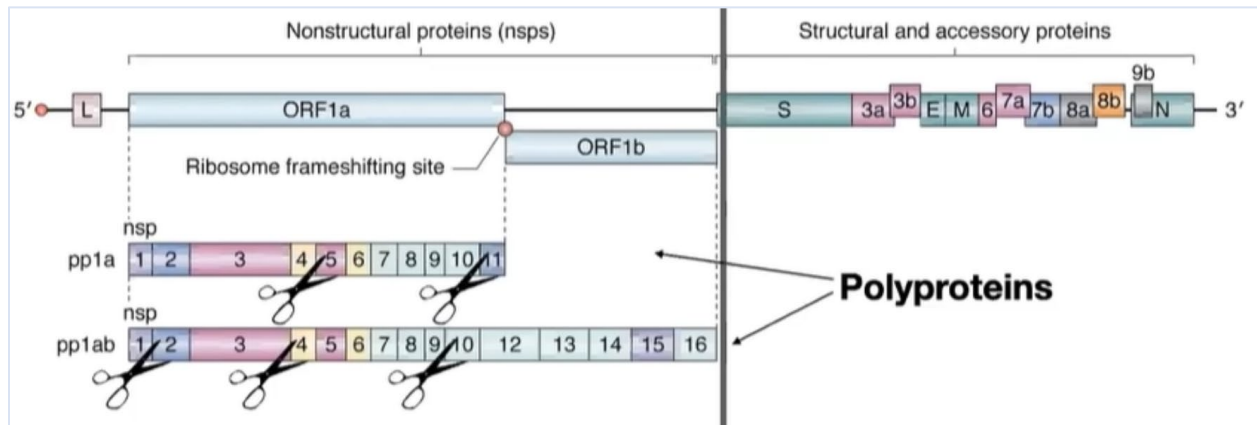
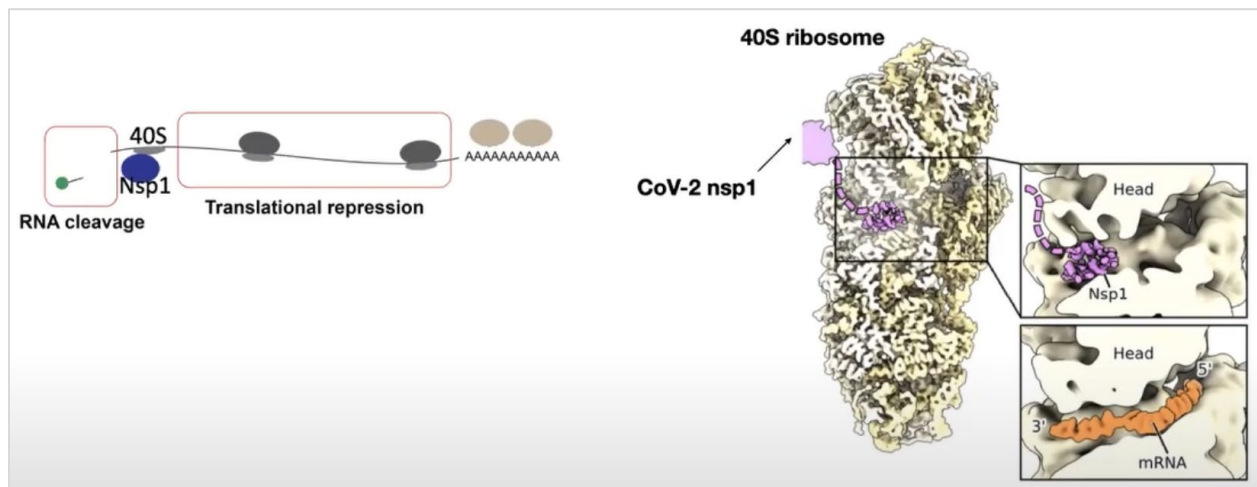


Figure 7: (Top) SARS-CoV-2 genome<sup>148</sup> encodes nonstructural proteins (nsp), structural and accessory proteins. Nsps are encoded by ORF1a & ORF1b generating pp1a (nsps 1-11) or pp1ab (nsps 12-16). The structural and accessory proteins are synthesized by translation of their respective sub-genomic mRNAs. (Bottom) Translational repression (Kamitani *et al*) and binding to 40S ribosome (Thoms *et al*) by Nsp1.





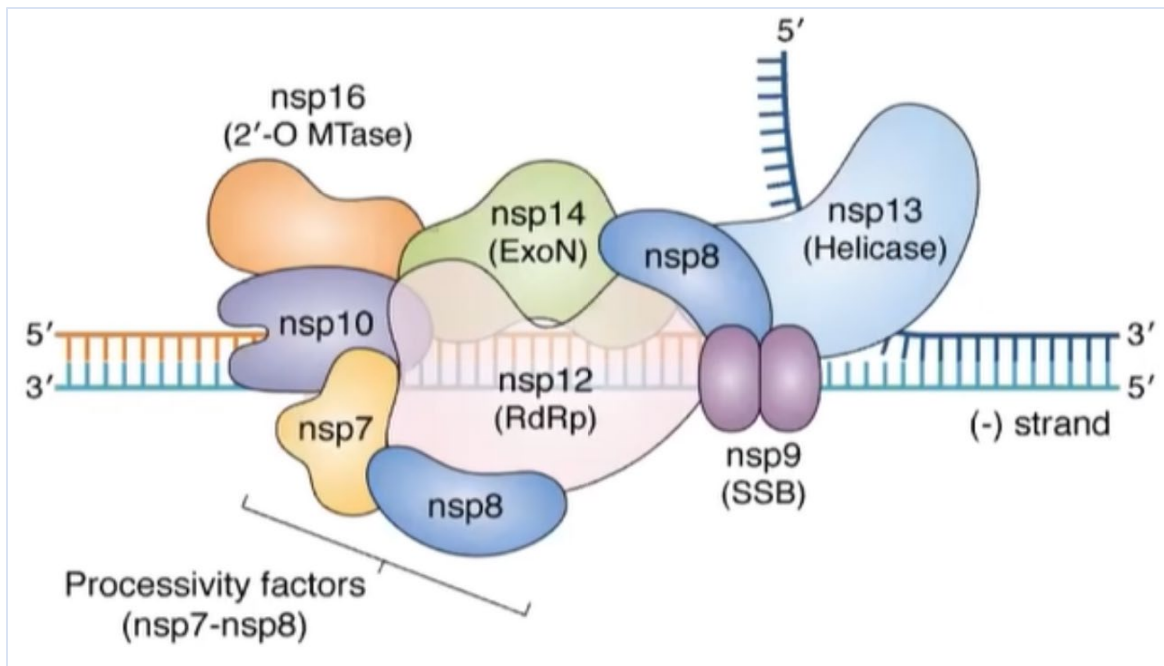


Figure 8: The positive sense (+ss) RNA genome is translated by the host translation machinery to make polyproteins that are co-translationally cleaved by proteases (PLpro/Nsp3 and 3CLpro/Nsp5) encoded in the polyprotein to generate components of RdRp or RNA dependent RNA polymerase (Hartenian and Nandakumar *et al*). The RdRp complex uses the genome as a template to generate negative sense subgenome and genome length RNAs, which are in turn used as templates for synthesis of positive sense full length progeny genomes and subgenomic mRNAs. Each and/or any protein factor in this complex may be a target for anti-viral reagents, for example, aptamers, antibodies, small molecules and inhibitors.

The conundrum and complexity presented by an abundance of anti-viral targets, a variety of strategies and potentially many cell types susceptible to infection, adds to the pharmaceutical dilemma where the problems of bio-availability, cross-reactivity and toxicity may force a solution to extinction. Viral proteins are distinct but structural homologies and overlapping functional issues are non-trivial.

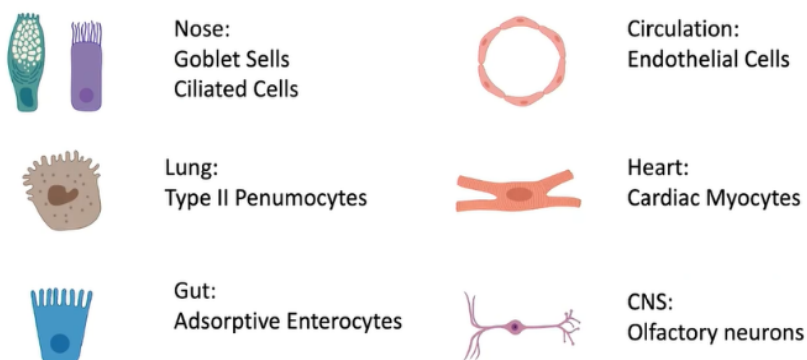


Figure 9: Identification of ACE2 receptors on many other cells (humans). The SARS-CoV-2 virus is not only a respiratory virus or results only in pneumonia. It is causing systemic diseases presenting a vast array of symptoms and acuity.

The medical chaos due to our lack of understanding of the biological minutiae of SARS-CoV-2 is not completely without a silver lining, albeit bleak. The ray of “hope” emanates from ExoN (Figure 8) the protein produced from Nsp14 segment of ORF1b (see Figure 7). It appears that SARS-CoV with inactivated ExoN is growth impaired and mutates at a much higher level (>20-fold<sup>149</sup> higher, see right panel in Figure 10). SARS-CoV with one of the longest genomes (see Figure 16) among common RNA viruses (HIV, Influenza, Rhino, Ebola) abhors errors<sup>150</sup> in replication (not corrected in other common RNA viruses with low fidelity RNA replication). High fidelity replication has enabled SARS-CoV to maximize its genome size (see Figure 16) using RNA-dependent proof reading system, repair and error correction implemented by Nsp14-ExoN (there are Nsp14 homologs in other viruses). Lack of error correction in humans<sup>151</sup> may result in disease, dysfunction and death, even due to point mutations.

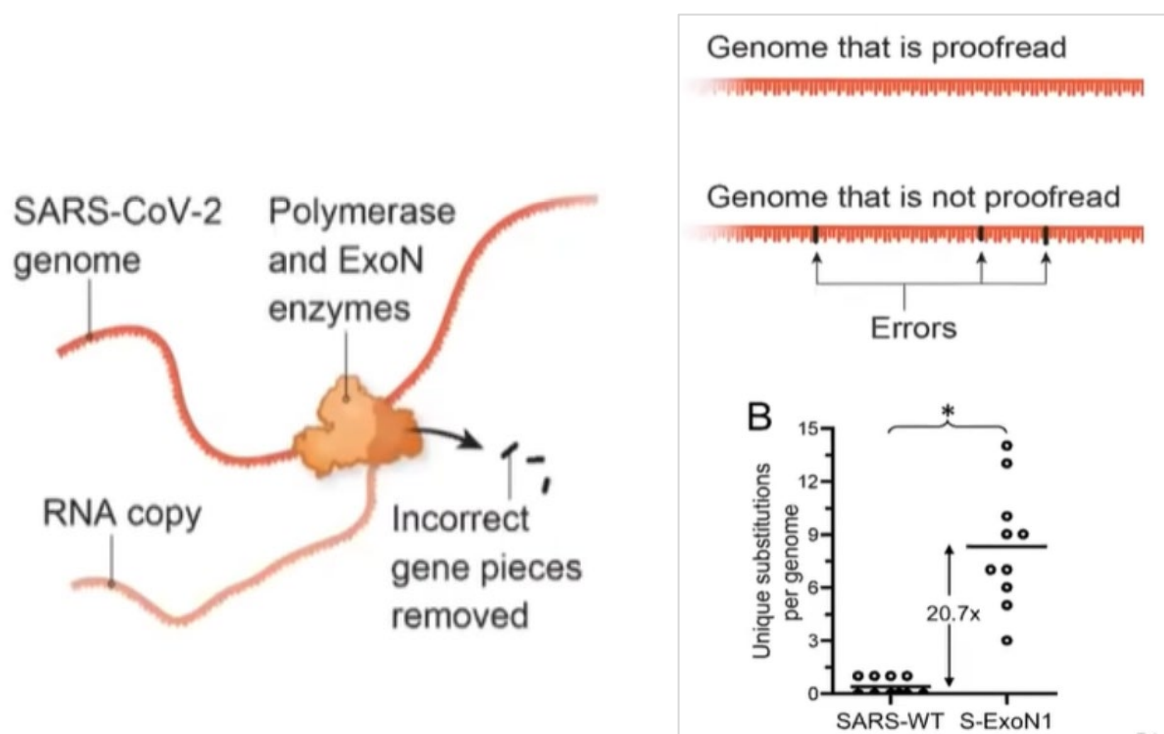


Figure 10: Mutated or inactivated Nsp14-ExoN results in >20-fold increase (Eckerle *et al* 2010) in genomic errors (B, right panel). ExoN in RdRp of SARS-CoV-2 enables error correction (left panel).

Error correction in SARS-CoV-2 may have implications for optimizing target selection for anti-viral strategies. The choice of the receptor binding domain in subunit 1 (S1 RBD) of the SARS-CoV-2 Spike protein, therefore, may be incomplete as a target (Figure 3). It appears that the fusion machinery and the fusion peptide (FP) region of the Spike protein (subunit 2) is better conserved and will *continue* to *remain* better conserved due to the error correction mechanism (see Figure 10). Hence, sub-segments within subunit S2 of S protein may be better targets. The obvious caveat in this discussion is whether the chosen sub-segments in S2 may be sufficiently exposed or available to bind with the anti-viral molecules.

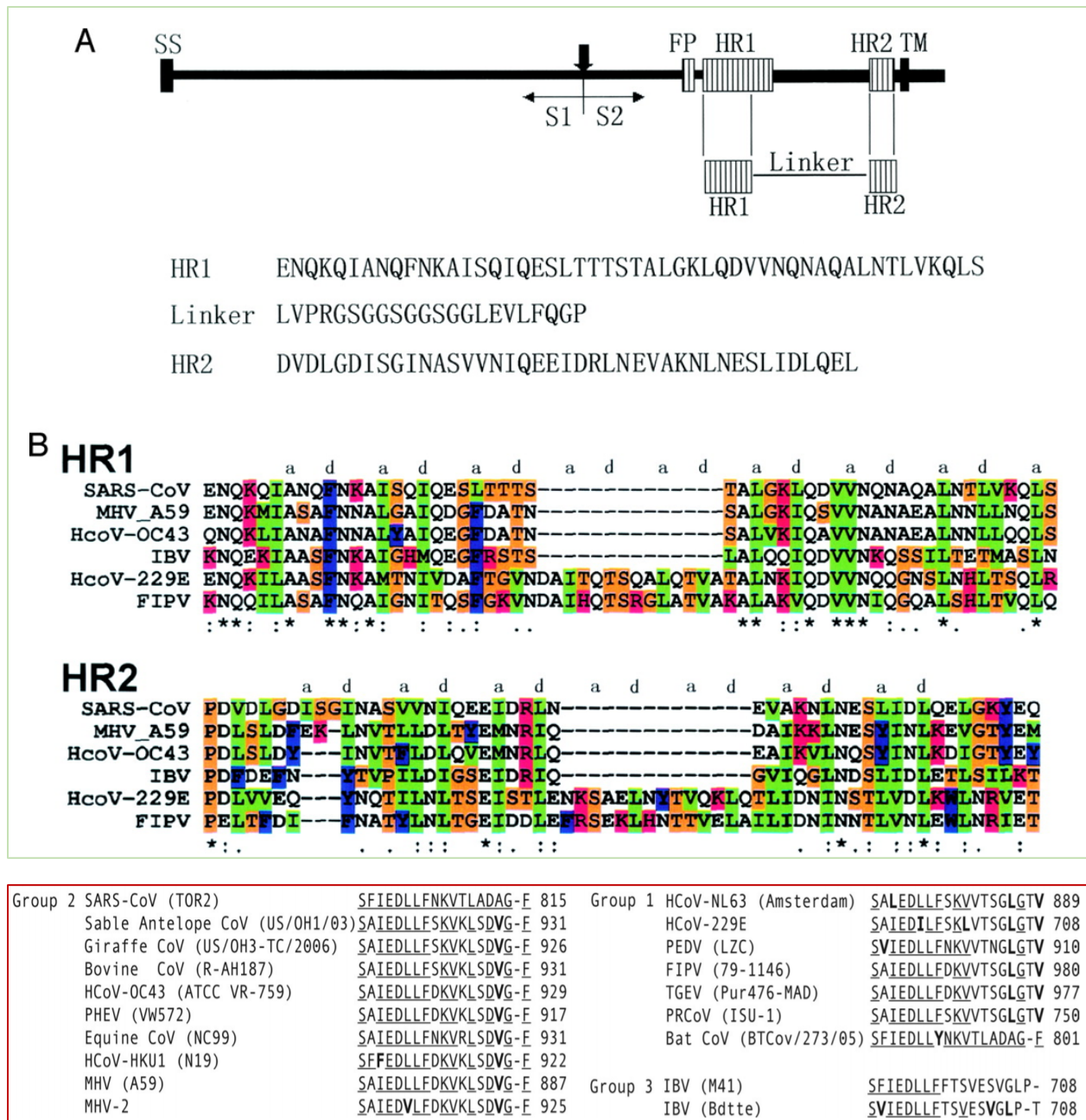


Figure 11: S protein (trimer) consists of 2 structurally noncovalently linked domains, S1, contains RBD (receptor binding domain) and S2 contains the fusion machinery and the fusion peptide<sup>152</sup> (FP). Site of proteolytic cleavage → vertical arrow. S2 contains 2 HR (heptad repeat)<sup>153</sup> regions HR1 (898 –1005) and HR2 (1145–1184) connected by 22-amino acid linker (LVPRGSGGSGGSGGLEVLVQGP). Hydrophobic residues (a and d positions in heptad repeat regions) are conserved. SS (N-terminal signal sequence), TM (transmembrane domain, C terminus), FP (fusion peptide, bottom<sup>154</sup>), IBV (infectious bronchitis virus), FIPV (feline infectious peritonitis virus), MHV (murine hepatitis virus - murine coronavirus).

## 8. Discussion

Could we detect SARS-CoV-2 in saliva, prevent<sup>155</sup> membrane fusion and block viral entry<sup>156</sup> with the same aptamer or another type of molecule based<sup>157</sup> on the better conserved S2 of the SARS-CoV-2 Spike protein? Could we detect SARS-CoV-2 in saliva of asymptomatic individuals without COVID-19? ADD may use better targets for its aptasensors beyond RBD S1 and hACE2. ADD *can* be accomplished, as suggested by the evidence from creation of DeMEA<sup>158</sup> (but it uses high cost microfluidics<sup>159</sup>).

Even if ADD is successfully engineered to be a low-cost biomedical device for non-invasive detection, *dissemination* of ADD and other systemic surveillance tools will still depend on community-specific economics of technology<sup>160</sup> to facilitate diffusion and adoption. Bringing data and information together to make sense and extract foresight (uncertain of the value of hindsight<sup>161</sup>) will be a challenge which new initiatives<sup>162</sup> must address. Diffusion of the tool to vulnerable communities will be restricted unless the end-to-end system is cost-effective at a level where it is sustainable for repeated use, preferably daily, as a surveillance tool for humans, pets and farm animals.

Data when transformed into *usable* information may deliver value for the greater good, for the greatest number. ADD is one small surveillance tool but it isn't enough. Healthcare cannot be a knee-jerk reaction to epidemics and pandemics. Continuous monitoring (even for high risk individuals) may remain a mirage in view of the disproportionate socio-economic imbalance. While we must ADD up to address the crisis<sup>163</sup> at hand, we must also utilize this disaster as an opportunity to deploy profiling as a healthcare staple. Other tools, for example, wastewater<sup>164</sup> analysis<sup>165</sup> may offer transparency<sup>166</sup> and guide public health strategies regarding elements the community must address, in advance, to prevent melt-down of health services. When an emergency presents itself we must not disintegrate into quagmire.

Precision medicine and precision public health may benefit if we probe the broader question of physiological status as expressed by proteins but further complicated by our microbiomes<sup>167</sup>. Isolated snapshots of data may be rate-limiting for communities under economic constraints. But, convergence of data from ADD along with multiple levels of profiling<sup>168</sup> (DNA, RNA, protein, RDW<sup>169</sup>) as well as environmental<sup>170</sup> and wastewater<sup>171</sup> data<sup>172</sup>, if included<sup>173</sup>, may augment the value of information, which could be catalytic for medicine<sup>174</sup>, in general, if aggregated and shared between open<sup>175</sup> platforms.

Analytical skills necessary to deconstruct the data and reconstruct its meaning, relevant to the individual and/or the community, may pose a rather insurmountable barrier in terms of tools and/or human resources. The ill-informed inclination is to hastily pursue a “quick and dirty” version (perhaps shoddy, yet masquerading as good enough) without a long term view or a vision that embraces a sense of service, science for the good of society and access to global public goods for all. It goes without saying that one shoe does not fit all. It is obvious that ADD is not enough to better prepare for the future<sup>176</sup>.



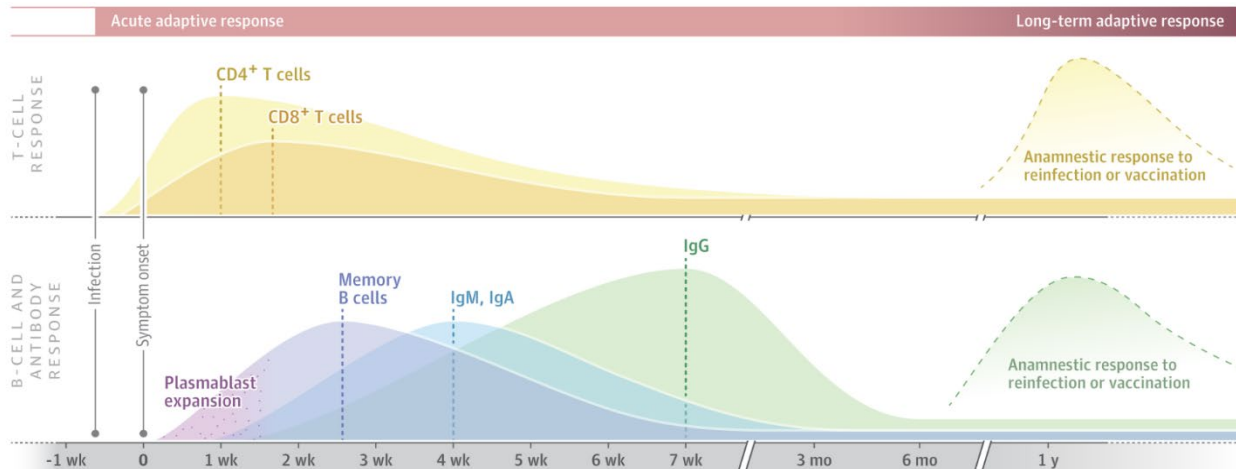
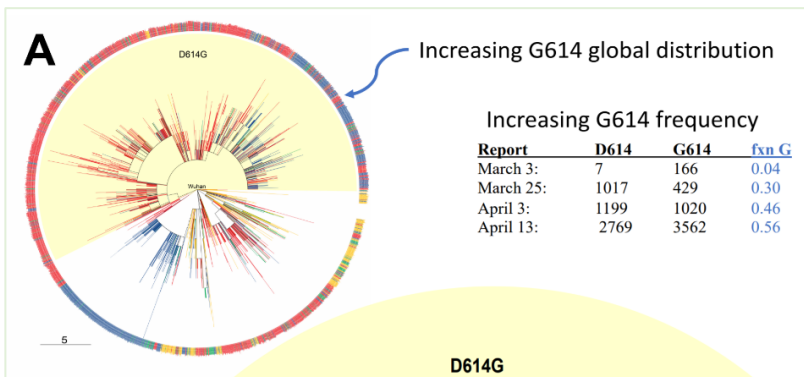
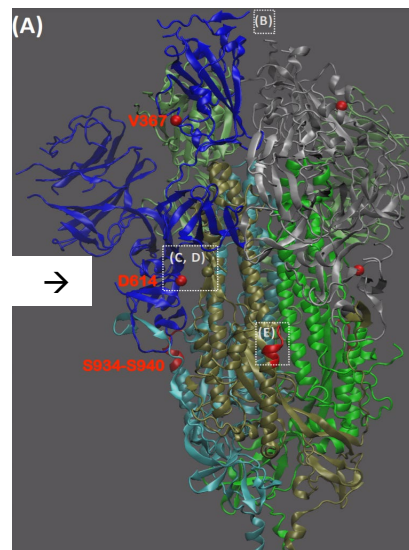
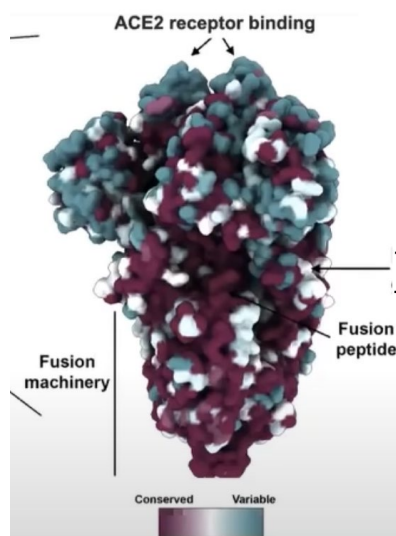


Figure 12: Serum from a significant percentage of patients (one third) recovering from COVID-19 have low viral neutralizing activity. Depending on the acuity of the infection, patients may or may not follow *standard* immune profile (top<sup>177</sup>). Low variation (Fig 10) in SARS-CoV-2 Spike protein is good news but mutations, D614G (middle panel and bottom) may still complicate<sup>178</sup> the immune response and expected anamnestic response to reinfection or use of classical<sup>179</sup> approaches<sup>180</sup> to vaccination.



If immunity from traditional vaccines are uncertain<sup>181</sup>, can we supplement with *alt-vaccines* (which are non-immunogenic, for example, aptamers), to better prepare for low-cost and rapid<sup>182</sup> response to public health during future epidemics / pandemics?

### 9. Complexity of the biomedical scenario, socio-economic catastrophe and the public health crisis

Since 1980's the HIV epidemic has infected ~76 million people<sup>183</sup> (~1% of the global population) and almost half are dead (~33 million AIDS related deaths, disease caused by HIV) and currently the other half is still living or struggling with the disease. Yet, the thrust for HIV vaccine pales compared to the warp speed vaccine development collaboration<sup>184</sup> against SARS-CoV-2, which erupted in 2020 as the COVID-19 pandemic. Is it because SARS-CoV-2 is irreverent and indiscriminate in infecting humans?

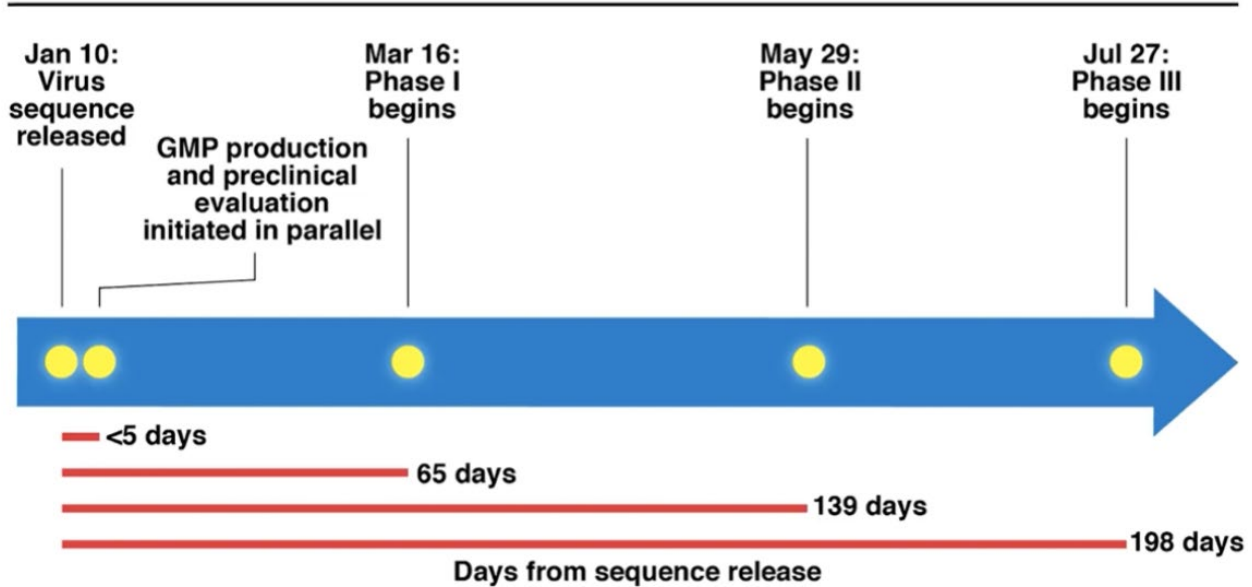


Figure 13: The timeline of SARS-CoV-2 Vaccine Development<sup>184</sup> (mRNA-1273 vaccine<sup>185</sup>) to control COVID-19 (codeveloped by NIAID, NIH and Moderna, Cambridge, MA). The mRNA encodes the SARS-CoV-2 full-length spike glycoprotein trimer, S-2P (stabilized<sup>186</sup> with two Proline<sup>187</sup> substitutions at the top of the central helix in S2 subunit). mRNA is encapsulated in lipid nanoparticles (0.5 mg per mL) and diluted with normal saline to achieve the final target vaccine concentrations<sup>188</sup>.

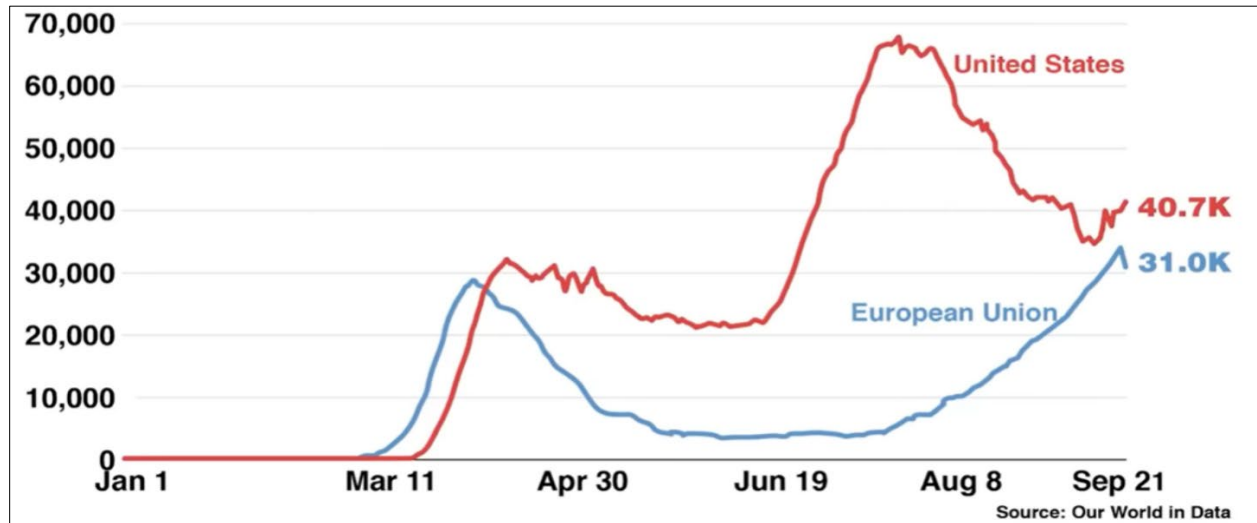


Figure 14: 7-day rolling average of new COVID-19 cases<sup>189</sup> from January through September 21, 2020.

Coronaviruses have long co-existed with humans and animals. Error correction (Figure 10) has made the genome of the coronavirus one of the largest among viruses (Figure 16). What does it mean? Compared to diseases<sup>190</sup> due to relatively unknown viruses<sup>191</sup>, and despite the flu pandemic ~100 years ago, the coronavirus, in less than six months, has changed, perhaps permanently, global thinking, trends and technology. Tobacco Mosaic Virus (TMV) was discovered around 1890-1892<sup>192</sup> but after more than 100 years<sup>193</sup> of virus discovery, we have *just now* acknowledged the threat to global health from viruses. Understanding the molecular basis of virulence is the single most important questions in basic biology which must be investigated by the best and brightest, if we ever expect to mitigate the risk from viruses.

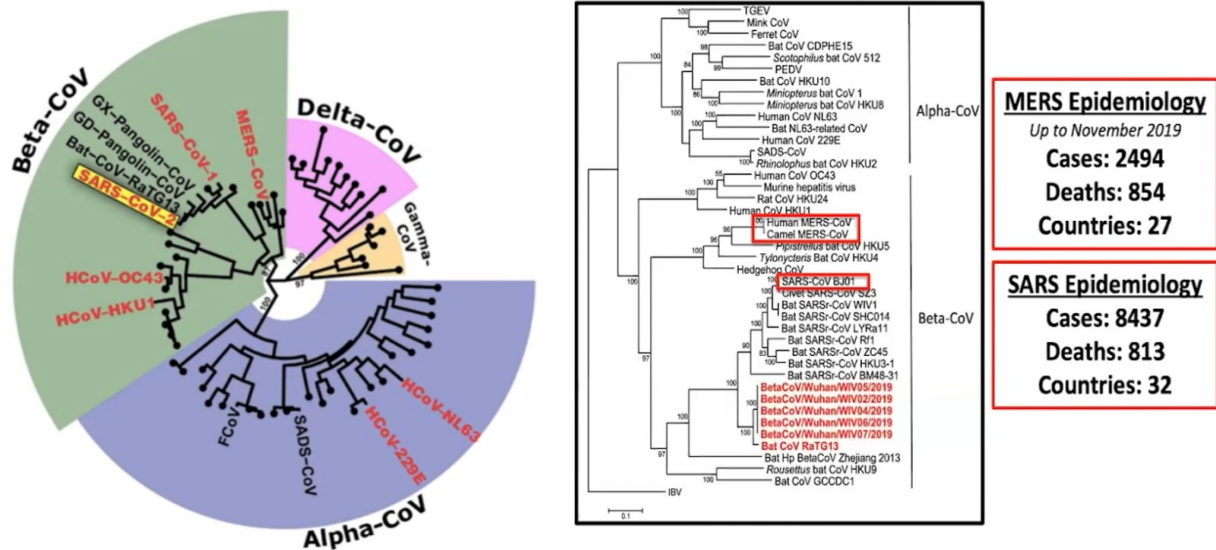


Figure 15: A family of (corona) viruses with pandemic potential (courtesy of S. M. Gygli, NIAID)<sup>194</sup>

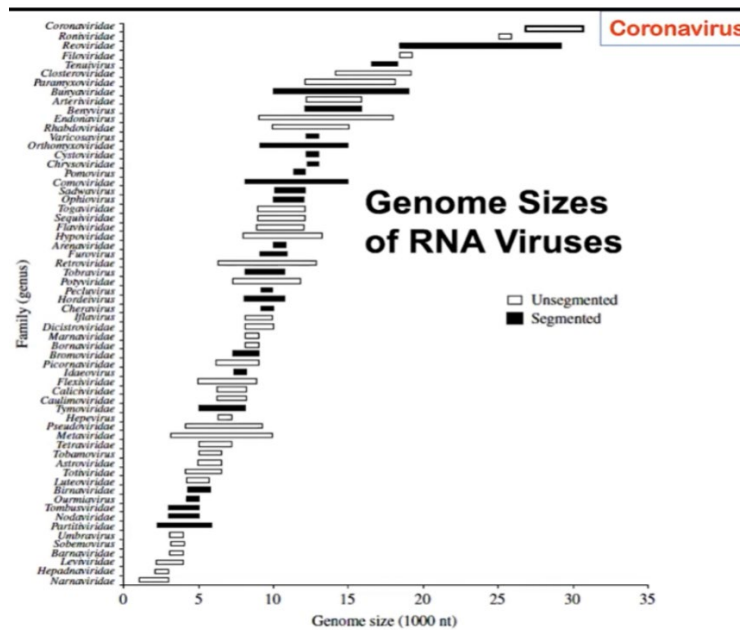


Figure 16: Coronavirus pandemic wasn't really expected<sup>195</sup> according to at least one global expert<sup>196</sup>. The coronavirus has the largest RNA genome. Is it just a coincidence or is there any bio-medical correlation?



While we still remain clueless about what constitutes virulence, the genome size does not offer any solace or solution. New evidence about Neuropilin-1 as a host factor<sup>197</sup> which facilitates cell entry<sup>198</sup> further thickens the plot. The deep quagmire<sup>199</sup> about  $R_0$  and  $k^{200}$  for COVID-19 defies epidemiological models<sup>201</sup> but prefers apocryphal Pareto<sup>202</sup> principles which suggests that 80% of new infections may be caused by only 20% (or less - 10% - see Figure 17 and reference 216) of the infected individuals. Are these individuals a high risk group due to inborn errors of immunity caused by mutations of genes involved in regulation of type I and type III interferons<sup>203</sup> (IFN)? How many more genetic factors may underlie the differentiation between super-spreaders<sup>204</sup> vs sub-spreaders for SARS-CoV-2? If the latter is true then how valuable is *generalizing* infection dynamics<sup>205</sup> from communities as a prediction tool for *overall* public health, advance planning and use as early warning<sup>206</sup> for cautionary preparation?

In future, genomic analysis may enlighten us if there are polymorphisms<sup>207</sup> which may partially account for this differentiation. It may be worth digressing to note that some individuals may be more susceptible to leprosy, caused by *Mycobacterium leprae*. Genes<sup>208</sup> associated with leprosy include HLA (human leukocyte antigen) proteins. Analysis of eleven HLA genes in 1155 Vietnamese individuals revealed 4 leprosy-associated independent amino acid variants [HLA-DR $\beta$ 1 positions 57 (D) and 13 (F), HLA-B position 63 (E) and HLA-A position 19 (K)] which comprised 2 pairs of linked genes, with one set conferring susceptibility [HLA-DR $\beta$ 1 and HLA-A] and one being protective<sup>209</sup>.

The demographics of infection by SARS-CoV-2 may be due to genetic<sup>210</sup> determinants<sup>211</sup> and individual outcomes<sup>212</sup> may be determined by our genes<sup>213</sup> as well as epigenetic factors which may be mapped to biomarkers<sup>214</sup>. At this point it is unclear whether the etiologic agent of this 2019 coronavirus pandemic should be referred to as SARS-CoV-2 where SARS imply severe acute respiratory syndrome.

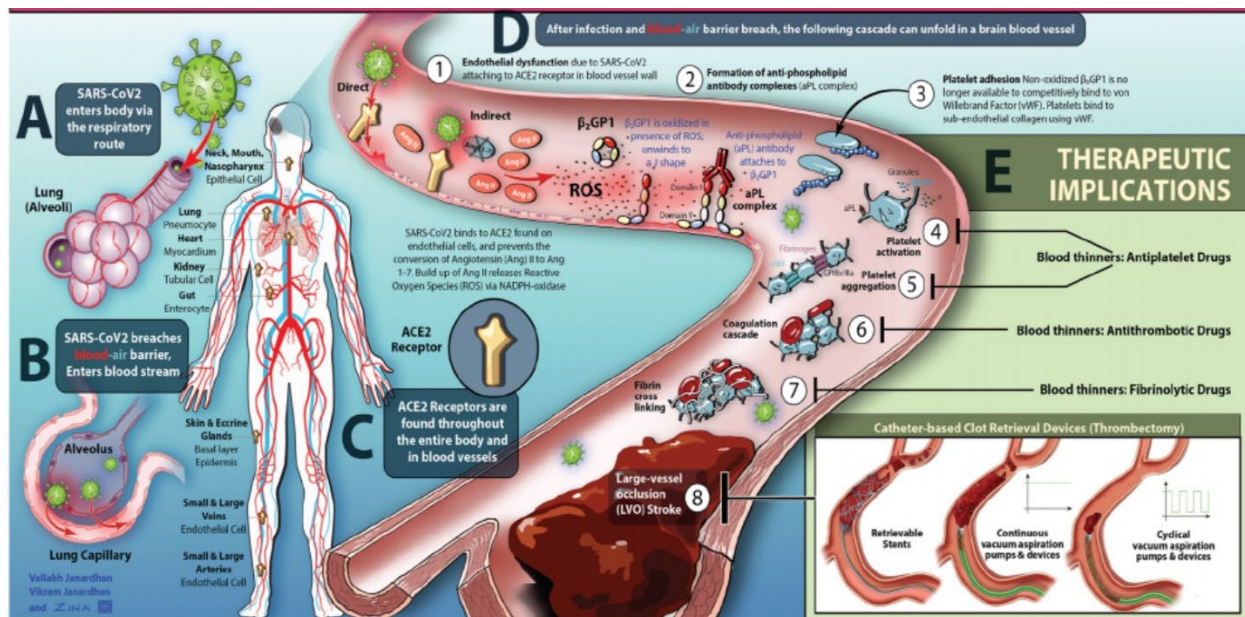


Figure 17: What is COVID-19? Respiratory illness? Blood clotting disorder<sup>215</sup>? Cardiovascular disease? Autoimmune disease? Opportunistic “killer” for (~10%) patients with severe COVID-19 pneumonia and high titers of autoantibodies<sup>216</sup> against different types (type I IFN- $\alpha$ 2 and IFN- $\omega$ ) of interferons?

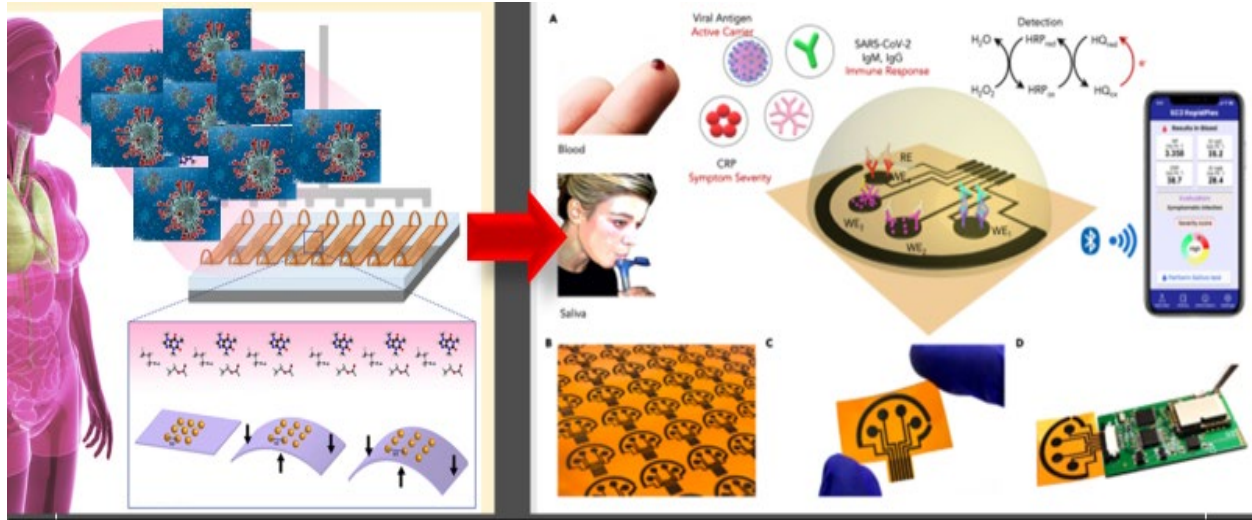


Figure 18: Hypothetical<sup>217</sup> 5<sup>218</sup> cent<sup>219</sup> *déjà vu* graphene sensor (RIGHT) detects SARS-CoV-2 antigens. Can smartphone detection<sup>220</sup> adapt<sup>221</sup> to other<sup>222</sup> sensors<sup>223</sup> (LEFT) to detect<sup>224</sup> SARS-CoV-2 in exhaled<sup>225</sup> breath<sup>226</sup> by mouth? Smartphone<sup>227</sup> *breathalyzer* for malaria<sup>228</sup> and marijuana (tetrahydrocannabinol)<sup>229</sup> is close at hand. Can it serve<sup>230</sup> as a global surveillance tool to bridge the chasm of inequity?

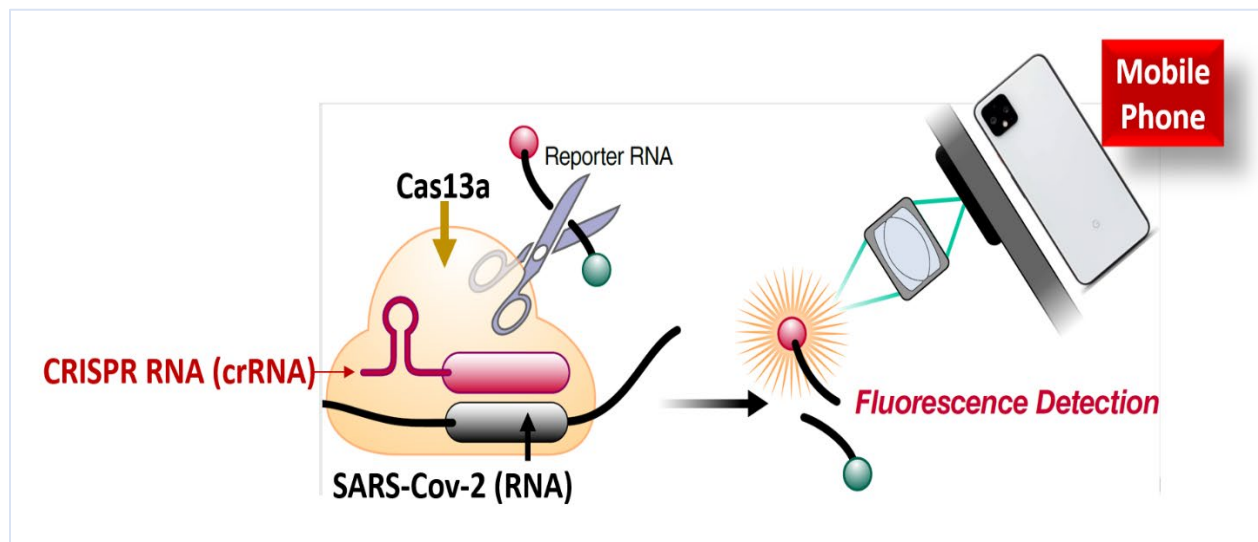


Figure 19: Detection of one copy RNA per  $\mu\text{L}$  (microL) from SARS-CoV-2<sup>231</sup> with mobile phone camera. Cas13a (C2c2) is complexed with a CRISPR RNA (crRNA) containing a programmable spacer sequence (red tube) to form a nuclease-inactive ribonucleoprotein complex (RNP). When the RNP binds to a complementary *target* RNA, it activates HEPN (higher eukaryotes and prokaryotes nucleotide-binding domain) motifs of Cas13a that then indiscriminately cleaves surrounding ssRNAs. Target RNA binding and subsequent Cas13 cleavage activity can therefore be detected with a fluorophore-quencher pair linked by an ssRNA, which will fluoresce after cleavage by active Cas13. Ott *et al* used the SARS-CoV-2 nucleocapsid (N) gene as the template (detection *target*) to create an array of crRNA spacer (red tube).

The socio-economic fall-out from the stochastic spread of infection and non-deterministic trends affecting certain countries, select groups (race, ethnicity) and underserved clusters, may be an example of “*writing on the wall*” we are slow to acknowledge. The cost of testing 100,000 individuals in the US approximate \$6 million. If 30 million tests are performed weekly it would require an additional \$75 billion and adding the cost of contact tracing might bring the total to approach \$100 billion<sup>232</sup>.

The “*writing*” says that the successful NIAID-Moderna mRNA-1273 vaccine or any other safe and effective vaccine against SARS-CoV-2, when it may become available in 2021 or earlier, may still be out of reach for *billions* of people. CRISPR<sup>233</sup>-based tests may be promising<sup>234</sup> in the future (see Fig 19). BinaxNOW \$5 test<sup>235</sup> is at hand but may not be feasible for daily use in communities under economic constraints. The case of Hepatitis-C<sup>236</sup> is an example how even after nearly 50 years, anti-viral drugs are not within the buying power of billions of people. *Success of vaccine is not equal to access to vaccine.*

Death, destruction and the decay of civilization<sup>237</sup> may continue and may *continue to amplify* in certain regions of the world, long after the pandemic. *If* the current pandemic is substantially contained by the end of 2021, then the aggregated loss from mortality, morbidity, mental health conditions, and direct economic losses in the US alone is conservatively estimated at \$16 trillion<sup>238</sup>. The US economy is about a quarter of the global economy<sup>239</sup>, hence, extrapolation suggests that losses due to this pandemic may be an estimated \$64 trillion, globally (about 80% of the global GDP<sup>240</sup>).

This mundane proposal is an elusive quest for an alternative path, albeit temporary and vastly incomplete, perhaps through the use of aptamers (or other variations based on oligonucleotides<sup>241</sup>) to partially bridge the chasm of inequity<sup>242</sup> and cushion the blow from the mortality and morbidity, yet to be witnessed. Healthcare is a pillar (FEWSHE - food, energy, water, sanitation, healthcare, education) of life and living but it is prudent to avoid indulging in any illusion or delusion because neither aptamers nor vaccines or CRISPR tools, irrespective of their respective efficacies, are a panacea for the restoration of civilization, even if this pandemic subsides in a few years. The quintessential ingredients for public health and global rejuvenation are scientific credibility, color-blind magnanimity and ethical leadership.

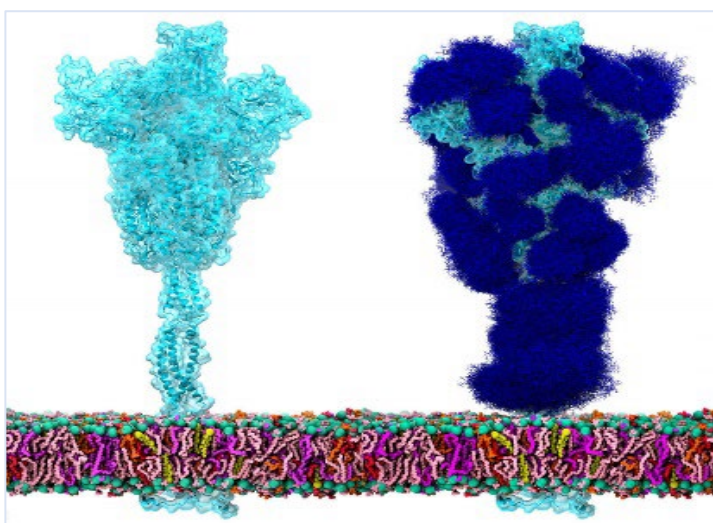


Figure 20: Similar to HIV, SARS-CoV-2 Spike protein uses a N-glycan shield<sup>243</sup> to thwart the host immune response (coating of N-glycans in cobalt blue, right). Mutations<sup>244</sup>, inborn errors of the immune system and other factors may make SARS-CoV-2 endemic<sup>245</sup>. Thus, it behooves us to explore other risk mitigation strategies. Anti-sense oligonucleotides, mini protein inhibitors<sup>246</sup> and aptamers (this proposal) are alternatives.



## 10. Post-Pandemic Public Health and Healthcare: Broad Spectrum use of Sensors with Smartphones

Transaction cost<sup>247</sup> of *humans-in-the-loop* detection or surveillance<sup>248</sup> is often astronomical<sup>249</sup> and the burden of cost<sup>250</sup> for public health and healthcare systems is prohibitive even for affluent nations. For resource constrained communities, the mortality and morbidity due to lack of access to primary care must be reduced. Can we effectively combine the ecosystem of sensors, smartphones and data informed decision analytics to deliver usable information at the point of care or point of need, in near real-time?

In general, part of the solution may be found in remote sensing and imaging tools (oil and gas pipelines<sup>251</sup>, leaves<sup>252</sup>, tree<sup>253</sup> canopy<sup>254,255</sup>, radiation<sup>256</sup>). SEE or “sense everything everywhere” (paint-based computation<sup>257</sup>, sensors in fabrics<sup>258</sup>) was a ‘touchy-feely’ mantra at the turn of the millennium buoyed by the principle of ubiquitous<sup>259</sup> computing<sup>260</sup> but stumbled in practice due to the cost<sup>261</sup> of computation<sup>262</sup>. Vinton Cerf’s “I P on Everything”<sup>263</sup> (*I pee* on everything) was the witty clarion call for embedding the IPv6<sup>264</sup> standard<sup>265</sup> in all things<sup>266</sup> to enable “bit dribbling” between “digital” objects. These ideas were preceded by “tangible bits”<sup>267</sup> from Hiroshi Ishi<sup>268</sup> and the “atoms to bits” paradigm<sup>269</sup> of “*Internet 0*” from Raffi Krikorian<sup>270</sup> and Neil Gershenfeld<sup>271</sup> followed by the origins<sup>272</sup> of internet of things<sup>273</sup> by Sanjay Sarma<sup>274, 275</sup> and others<sup>276</sup>. The borborygmi of radio frequency identification (RFID) and standardization of the electronic product code (EPC<sup>277</sup>) shifted the thinking from stationary goods and products with static bar codes to dynamic digital objects which can be uniquely identified in any process or supply chain and tracked and traced digitally between any number of transactions, globally.

Project Oxygen<sup>278</sup> offered extraordinary insight into the art of the possible<sup>279</sup> and represented a consilience and confluence<sup>280</sup> of ideas but it was cost-prohibitive for real world applications, circa 2000. With decreasing cost of computation<sup>281</sup>, memory<sup>282</sup>, data storage<sup>283</sup> and transmission<sup>284</sup>, these streams, which were occasionally bubbling since Isaac Asimov’s<sup>285</sup> *Sally*<sup>286</sup> in 1953, turned into a raging river bursting its banks. The convergence of these tools with initial thoughts about the networked physical world<sup>287</sup> were far more than the sum of the parts. It exploded to become the inescapable tsunami of IoT<sup>288</sup> which has infected every domain. The anastomosis<sup>289</sup> of IoT with cyberphysical systems<sup>290</sup> (CPS<sup>291</sup>) has penetrated almost every field from asteroids to zeolites and engulfed them within the new<sup>292</sup> laissez-faire world of DIKW<sup>293</sup> hierarchy. The mobile smartphone represents the grand conduit for the aggregated dissemination of distributed facets emanating from the DIKW pyramid. The mobile platform appears to be the global choice to access and implement all and any service which is possible, via the smartphone, in some form or the other, where the ubiquitous device serves as the platform for information<sup>294</sup> arbitrage.

ADD is a recognition element and a tiny part of this landscape. ADD enables the sensor, data is captured, analyzed, communicated and visualized on a smartphone. It may detect infectious molecules relevant to SARS-CoV-2 in saliva and nasal swabs for early detection to prevent the spread of the virus. This principle is applicable to *any* infectious agent as well as any physiological molecule of interest (see Figure 21). The potential of developing a “breathalyzer” (identifying molecules in breath, see Figure 18) may make it easier to detect any molecule or molecules which are either volatile or lighter than air. ADD may be the “*killer-app*” that IoT<sup>295</sup> was searching since its conception<sup>296</sup> circa 1999.

Based on the idea of swappable, modular, flash drives, sensors-on-a-chip in the form factor of flashdrives are not hypothetical but frontrunners as potential tools for dealing with infectious<sup>297</sup> diseases. Cameras, accelerometers (speed, movement), gyroscopes (tilt, orientation), magnetometers (compass), lidars (range, depth sensing from reflected laser signal), GPS and other “sensing” tools are increasingly “standard” with smartphones. These “detectors” makes it possible to use multiple mediums and phases for detection of signals from molecules, changes in dipole moments (electro-magnetic field) and perhaps even perturbation *ambient* electromagnetic waves (transmission and capture of reflected radio waves).



Figure 21: Billions of users in underserved geographies may access limited health services by using<sup>298</sup> ubiquitous tools that does not require installation of new infrastructure and re-uses “mobile lifestyle” devices to partially bridge the scarcity of resources. Smartphones may be catalytic for delivery of service, remote monitoring<sup>299</sup> and health surveillance, not restricted to infectious diseases but as physiological probes for health and homeostasis or detecting onset of disequilibrium (BNP, Brain Natriuretic Peptide).

Using information arbitrage to better contain the pandemic is the thrust of ADD. Expanding this principle as a routine for public health and healthcare, in general, is not a leap of imagination but natural progression. It bears reiteration that data informed decision analytics (DIDAS) must embrace sensor data plus smartphone (SDS) applications not as “pilot” projects but science in the service of society to catalyze the **SENSIBLE** system (**SENS**ors and **I**nformation arbitrage via **moBiLE** system). The marriage of DIDAS with SDS in the affluent world is a matter of social acceptance of SENSIBLE but the penchant for profit-first and lack of leadership are holding us back. In the rest of the world the barrier to diffusion of life-saving tools are, albeit with exceptions, greed, pursuit of unethical profitability, cost or paucity of infrastructure (engineering, energy, telecommunications) and rampant inequity in social cohesion.

One milestone for smartphone-based health surveillance may be the non-invasive<sup>300</sup> glucose<sup>301</sup> monitoring<sup>302</sup> system which the healthcare system failed to aggressively adopt<sup>303</sup> despite significant<sup>304</sup> advances<sup>305</sup>. The chest-thumping about diabetes pandemic<sup>306</sup> continues in parallel with avoidance of available<sup>307</sup> solutions<sup>308</sup>. Cholesterol<sup>309</sup> monitoring<sup>310</sup> using smartphones<sup>311</sup> may be a preventative measure for adults at increased risk for a slew cardiovascular diseases, a few of which may not show symptoms.

Decades after the discovery of atrial natriuretic peptide<sup>312</sup> (ANP, 1981), brain or B-type natriuretic peptide<sup>313</sup> (BNP, 1987) and C-type natriuretic peptide<sup>314</sup> (CNP, 1990), we *still* do not have SENSIBLE monitoring for BNP even though BNP sensors<sup>315</sup> including an aptamer-based<sup>316</sup> sensor for BNP-32 and cardiac Troponin I are available. These and other<sup>317</sup> biomarkers (CRP5/CRP6, TNF $\alpha$ ) are indicators of cardiovascular dysfunction including congestive heart failure (CHF) and state of the patient after myocardial infarction, in addition to other conditions. BNP and other markers are key to risk stratification, diagnosis, prognosis, disease monitoring, titration of therapy, and identification of therapeutic targets for cardiovascular disease. Brain Natriuretic Peptide concentrations >400 pg/mL and N-terminal (NT) pro-BNP >400-900 pg/mL (age related) are prognosticators of congestive heart failure. Analysis of 48,629 patients<sup>318</sup> of acute decompensated heart failure found linear correlation between BNP levels and in-hospital mortality. Failure of BNP to decline during hospitalization predicts death or rapid re-hospitalization. However, BNP levels of 250 picograms per mL (pg/mL) or less during discharge predicts potential for survival. Accelerating availability of sensors<sup>319</sup> and transforming innovations<sup>320</sup> to SENSIBLE systems for prevention of cardiovascular disease should neither suffer from paralysis due to analysis nor asphyxiated by the rancour over margin of profitability.

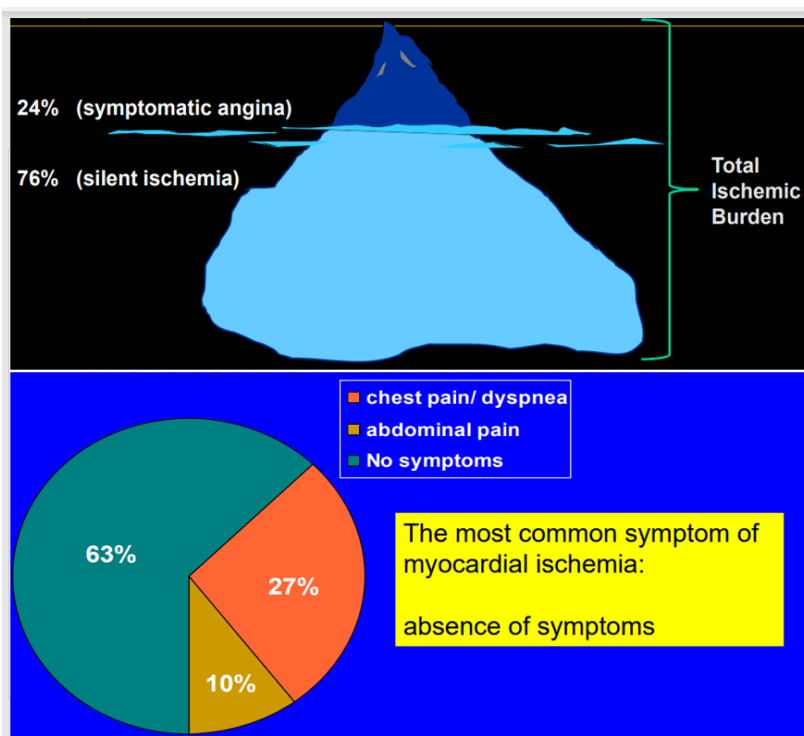


Figure 22: Tip of the Ischemic Iceberg<sup>321</sup> hides >63% of the individuals who lack symptoms but are increasingly at risk for CVD, ischemia, myocardial infarction, congestive heart failure. BNP and other biomarkers may reduce the risk using the SENSIBLE system. We know these facts<sup>322</sup> for ~40 years. Yet, the proponents of prevention *policies* wear that perpetual unctuous grin assimilating both the promises of a television evangelist and the sympathies of a funeral home director of marketing.

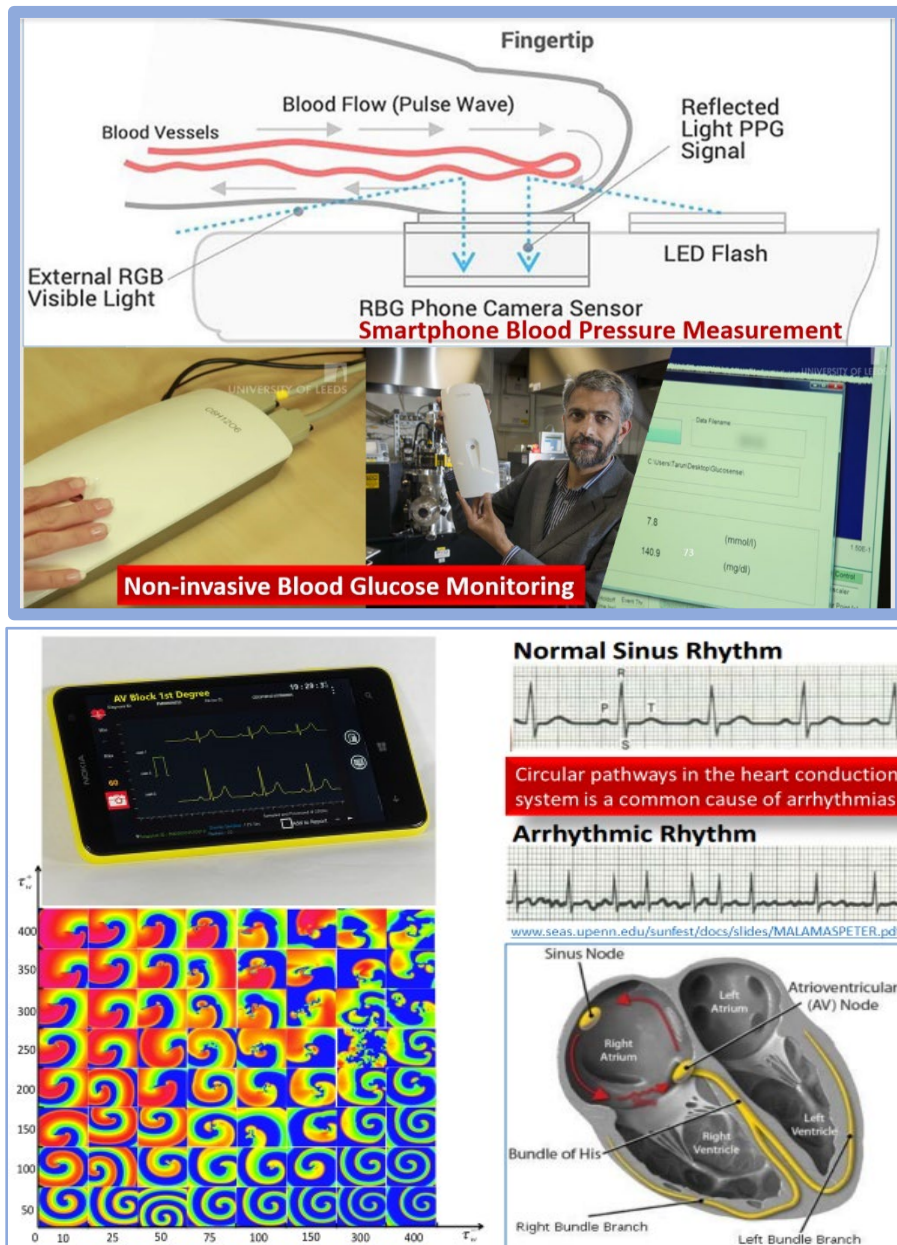


Figure 23a: Smartphone cameras, LEDs, LiDARs and a FEAST<sup>323</sup> of signal transduction tools (optical, Raman spectroscopy, electrochemical impedance spectroscopy, surface plasmon resonance) are now available as data carriers. Cartoon (top) shows smartphone-based blood pressure<sup>324</sup> and non-invasive blood glucose<sup>325</sup> monitoring<sup>326</sup>. The SENSIBLE system may be used to estimate blood cholesterol level, hemoglobin<sup>327, 328, 329</sup> and uric<sup>330</sup> acid<sup>331</sup> as indicators of health, albeit imperfect. Data from smartphone based optic disc<sup>332</sup> exam, photoplethysmography<sup>333</sup>, electrocardiograph for arrhythmias<sup>334</sup> (see bottom panels), general ECG<sup>335</sup>, heart<sup>336</sup> rate, respiratory<sup>337</sup> rate (*reflection* of radio waves), pulse oximetry<sup>338</sup> and other vitals, collectively, may create precision physiology portfolios (open data source interoperability).



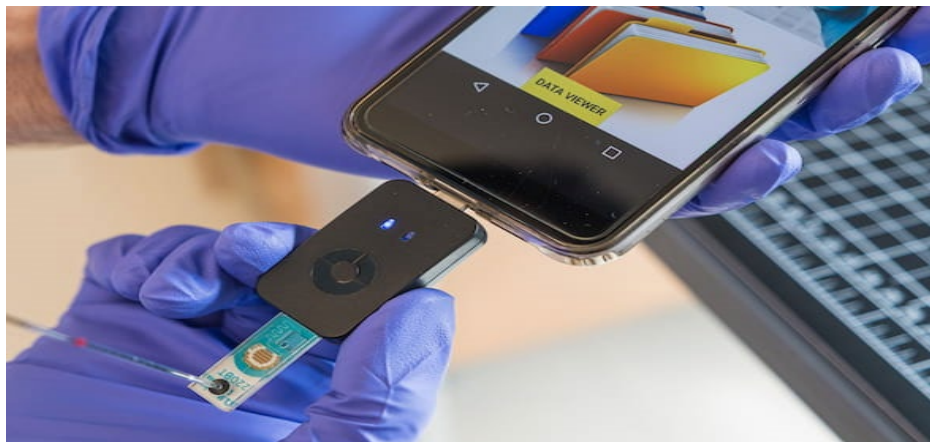
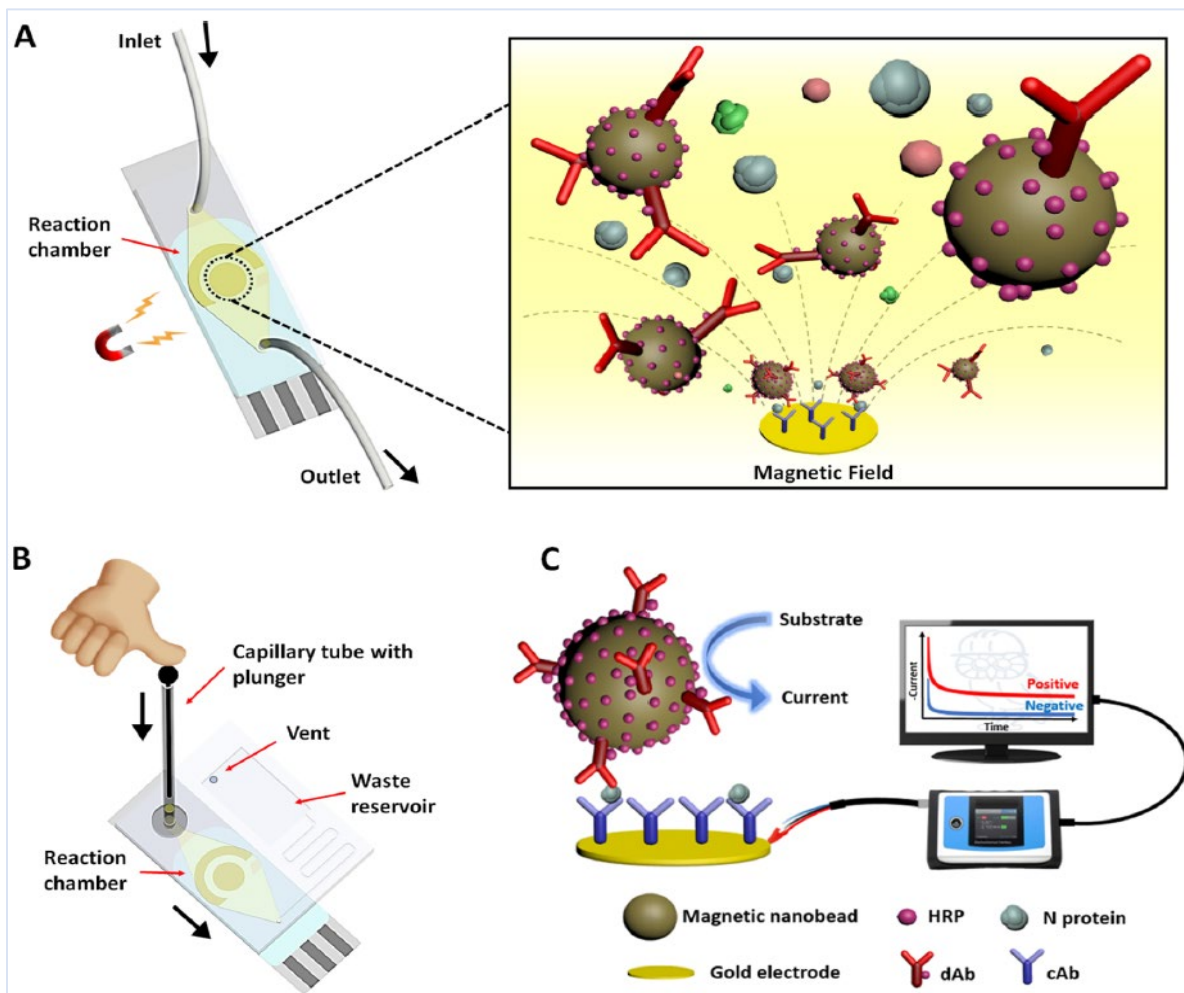


Figure 23b: Example of smartphone tool using magnetic nanobeads in a microfluidic immunosensor<sup>339</sup> which can detect SARS-CoV-2 nucleocapsid (N) protein in serum. The requirement for serum decreases its appeal as a tool for mass deployment. How many parents will consent to this invasive procedure for their children if the school requires testing? Smartphone tools must be non-invasive to be useful for all.

It is likely that *hundreds* of papers are published *weekly* on sensors, many of which may be adapted/adopted as a part of the SENSIBLE family. Biosensing using mobile devices at the point of use is a staple, for example, in the food industry (spoilage, contamination, security), soil, water, agriculture, manufacturing, chemical industry, transportation, energy, etc. But, lack of open data and restrictions on data interoperability makes the transformation of data to information quite difficult. Scientists are eager to drill deeper to develop yet another sophisticated<sup>340</sup> sensor but real-world user are in quest of *answers* at the point of use, and does not have the luxury to deal with numbers, compilers and programmers.

This discussion about the cacophony of available sensors is an embarrassment of riches from decades of research and development scattered as parts, in silos or locked by patents. The form and functional orchestration and integration necessary for sensors to contribute to precision physiology requires cross-pollination of ideas. Multi-disciplinary teams are necessary to create end-to-end working SENSIBLE systems which can be synchronized, if authorized, as a part of the public health information system. If that data is shared in real time, it may reduce mortality, morbidity, cost to society, decrease the burden on emergency medical professionals, and actually aid in preventing dysfunction. If this data is anonymized to serve molecular epidemiology, it may help precision public health and channel benefits to the community by revealing the environmental conditions or instances which need additional attention.

A plethora of brilliant experts with deep knowledge can fill any university hall but few have the breadth of ideas which, if synthesized, synergized and integrated, may helps to address or even solve a real problem in the public domain where non-experts are the end users. Solutions based approach must combine depth with breadth to deliver the fruits of science to society as global public goods. The latter may be missing in the academic context where chronic search for scholastic erudition is the norm. The concept of essential products and services as global public goods may not be appetizing in the corporate context due to their perpetual penchant to promote profit and profiteering, first. The cleavage between purpose and profit needs a new bridge and a new breed of thinkers and leaders with altruistic traits.

The laser-focus of biomedical professionals on saving the lives of those affected by COVID-19 and the public health community on preventing the spread of infection by SARS-CoV-2 is the only path, at present, to lift us out of the quagmire of the raging pandemic. Yet it may be crucial to use this disaster as a global opportunity to strengthen public private partnerships (academic-industry-government) for the ubiquitous deployment of global tools for early detection and prevention, not only for pandemics, but for public health and healthcare, in general. It is an enormous task and requires global leadership.

Ubiquity of smartphones is the available SENSIBLE platform to create at least one bridge over the chasm separating the *haves* from *have nots*. Inextricably linked economies of the under-developed, developing and developed nations makes it imperative that the leadership for global public health must be agnostic of prejudice. Trans-disciplinary cooperation and collaboration between corporations must rise above conventional economics<sup>341</sup>, narcissism, egocentricity and personal wealth creation. We need an overwhelming force for good, for a greater purpose, for the greatest number<sup>342</sup> (of people).

## 11. Comments

The complexity of the physiological dysfunctions associated with the SARS-CoV-2 pandemic calls for multiple routes for detection, prevention and therapy. ADD is a tool for detection and perhaps one of many other alternative<sup>343</sup> treatment (nanobodies<sup>344</sup>) that does not depend on the immune system. Immune response may be heterogeneous<sup>345</sup> based on individual genetic constitution (inborn errors) as well as natural decay<sup>346</sup> of antibodies and antigenic evolution<sup>347</sup> despite error correction in coronaviruses (see Figure 10, antigenic drift<sup>348</sup> may not be *exclusively* due to genomic mutations during replication).

The global diffusion of smartphones as ubiquitous devices is an opportunity for digital<sup>349</sup> public health to accelerate the use of smartphones for detection of any infectious agent. ADD is one tiny sub-component of the proposed SENSIBLE system. Realization of the SENSIBLE system is a difficult task. The deployment of the SENSIBLE system, with ADD and other sensor components, is an essential task, unless the veneer of health equity<sup>350</sup> is only that, that is talk, or in other words *lippenbekenntnis*<sup>351</sup>.

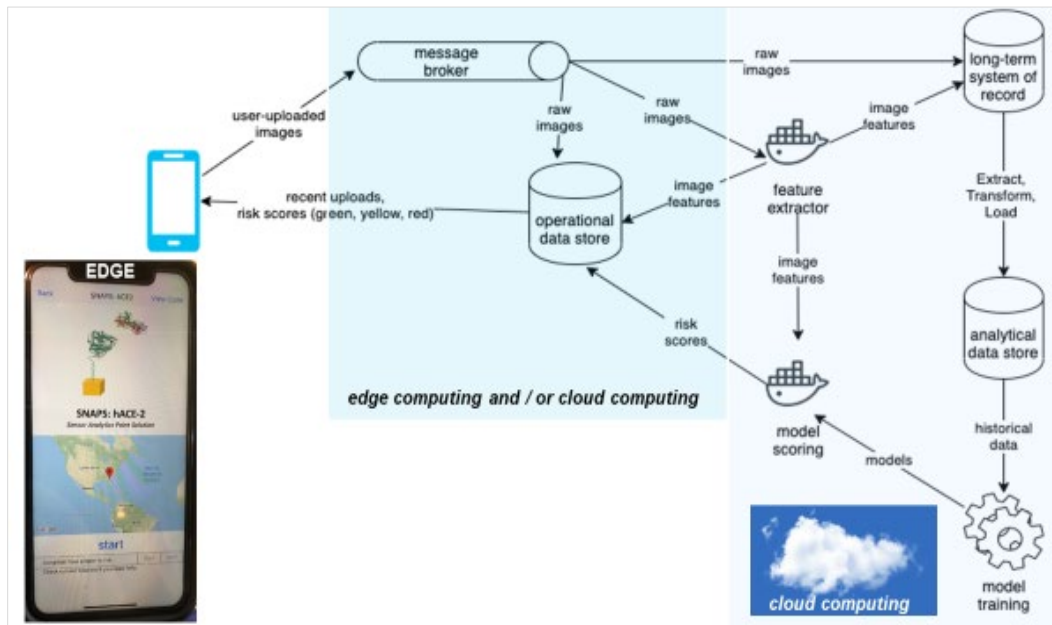
The caveat in this line of thinking is the over-emphasis on the sensor and the SENSIBLE system as if the value proposition is undeniable. The latter is true for the affluent economies of the world but detection without follow-up is an exercise in futility. The latter is common in communities under economic constraints where tools to detect (SENSIBLE system) are impotent because there are very few resources to attend to the public health or healthcare need identified by the SENSIBLE system. Just because the user can detect the presence of mercury in drinking water does not mean that the user has another alternative source of drinking water in under-served or dystopian communities. Is it more or less psychologically debilitating to drink water or consume food if the user is cognizant that the water is contaminated with mercury<sup>352</sup> or the food is laced with bacteria<sup>353</sup> beyond the level of food safety?

Incongruity between the *pursuit* of science, *implementation* of the fruits of science and science as a *measurable service* to society is a conundrum beyond the horizon of tools and technology, for example, the SENSIBLE system. Entrepreneurial innovation can create SENSIBLE but implementing SENSIBLE requires leadership imbued with a sense of the future, especially for low-income countries with ultra-low per capita *disposable* income. In the absence of charity, SENSIBLE for public health and healthcare must carry with it a pay-per-use price tag which may be a micro-payment or even a nano-payment but still it *must* be a non-zero payment for the system to be sustainable and survive long enough to deliver value.

Ephemeral gimmicks demonstrating SENSIBLE in geographies with GDP which may be less than an average household income in Europe is a deliberate act to deceive, dressed up in a marketing<sup>354</sup> garb by the glib, the smug and the smarmy. Enabling a SENSIBLE future and making it sustainable for most segments of the community, in greatest numbers, is a mission for a visionary leader who radiates the aura that kindness is a strength, not a weakness, humility is a virtue, not a lack of knowledge, that agreeing to disagree is a mark of civility<sup>355</sup> and dignity, not a character flaw, that fear is not a tool for maiming diversity, speech or peace, that progress of civilization is development of the freedom<sup>356</sup> to act on remediable injustices<sup>357</sup> and lift many boats, not a few yachts. The best man for *that* job is a *woman*.

## APPENDIX

## APPENDIX I - Components<sup>358</sup> for SARS-CoV-2 ADD Decision System (Fig 1) and Data Templates



### Message Broker

When users upload images (the data after scanning with the HoloLens app or equivalent mobile tool), the mobile application (on their phones) writes messages with the image content and other metadata to a message-broker, which may be cloud-based message queuing (MQ<sup>359</sup>) protocol (open source software). The message broker allows devices to quickly offload data and confirm “sent” to a user (if cloud based), thereby decoupling the user experience from the data store (even if it uses a temporary tinyDB on the device, if the network is unavailable to access the cloud in real-time at the point of use). Messages can be queued in topics and the system may enable autoscaling (as usage of the application increases, more users can be provisioned, process user uploads and get them stored). The uploads (data) are also sent by the message broker to the feature extractor and long-term storage database (may use the batch upload option when device is proximal to a high bandwidth gateway which can offer access to cloud services).

### Operational Data Store

The message broker transfers uploads to ODS (Operational Data Store<sup>360</sup>), which may be a cloud-based managed service or part of the tinyDB on the device, if cloud is inaccessible at the point of use. ODS must be able to store image data (supports binary blob column type) alongside time-index numerical and character data. It is intended to only serve “hot” (nascent) data to the application. Older data may be evicted (batch uploaded to cloud managed facilities) to optimize on-device service and prevent data amplification. ODS is tuned for fast random reads and serves requests made by mobile app when users view recent uploads and additional metadata about those uploads, including “risk scores”. ODS is optimized for fast writes and high efficiency time-series queries.



### **Feature Extractor**

Extracts additional metadata from images/data uploaded from the mobile app (uploads it to the long-term system of record<sup>361</sup> which includes raw data uploaded from the application, similar to “master data” in ERP<sup>362</sup>). Feature Extractor may convert the uploaded image into a numeric matrix<sup>363</sup> or create hash table or representation of a region<sup>364</sup> and correct for differences in resolution (for example, variation due to pixel density of cameras on different smartphones). Feature<sup>365</sup> vectors<sup>366</sup> may be maintained in the long-term system of record. It may be written to the operational data store to enable extraction/selection<sup>367</sup> of incoming data (uploads from message broker) relevant to these feature vectors.

### **Long-term System of Record**

Mobile applications may never access data directly from this data store<sup>368</sup>. Interactive-speed queries to this data store may not be supported. When necessary, objects stored in this “record” may be extracted and the data is loaded into an analytical data store. For object stores, this operation may be accomplished using query-over-files engines<sup>369</sup>. The thorniest problem that ferments within long-term data record is the inaccuracy of “accurate” data and the diabolical mayhem from “big data” if it is sourced and stored.

### **Analytical Data Store**

Scientists and data experts will need historical data (from uploaded samples) to train task-specific<sup>370</sup> machine learning (ML) models to assign risk scores to samples. Analytical data store (ADS database<sup>371</sup>) may be populated with data from the long-term system of record using scheduled batch data uploads.

### **Model Training**

In model training<sup>372</sup>, a statistical model is built from historical data. Models should be serializable<sup>373</sup> representations of the program generated by ML training. Serialization is essential for interoperability on different platforms. It is key to create composable models where models from different groups can be deconstructed to sub-elements which can be reconstructed to compose a new model (which may be greater than the sum of parts). Serialization enables the process of translating a data structure or object state into a format that can be stored or transmitted and reconstructed. Proprietary software vendors obfuscate or encrypt serialized data to prevent access. Standard architectures such as CORBA<sup>374</sup> define the serialization formats in detail to enable open access.

### **Model Scoring**

In model scoring, a model is called on input data, the model processes the input data and generates a prediction. The structure of this code depends on the *design choices* made during model training. For reliability of deployment, model scoring may run in a container<sup>375</sup> (an unit of software) which contains code (and all its dependencies) that uses a model to produce predictions on new input data. If model scoring runs in a container then the model can be arbitrary code in the developer’s language<sup>376</sup> of choice. Model scoring requires features created previously by the feature extractor (feature selection is critical).

## ADD DECISION SYSTEM IS FAR BEYOND OPTICAL SIGNAL DETECTION & IMAGE ANALYSIS

There is nothing new in proposing that we apply a patent-free 30 year old idea (aptamers) to a nascent problem (SARS-CoV-2). The expectation is that a credible scientific investigation may reveal if aptamers may indeed serve as an alternate to conventional wisdom (immune response). The scientific strength of this idea is based on a rigorous tenet of molecular biology which has repeatedly demonstrated that proteins bind to nucleic acids, as a fundamental mechanism of action in biological regulation. If this approach succeeds, it may help and save lives in the less affluent nations (80% of the global population), not only for detection but also in prevention and therapy (APPENDIX II).

The thrust of ADD decision system is far beyond the cartoon in Figure 1 suggesting optical data acquisition and image analysis to visualize outcome on a smartphone. Using optical signal is probably more user friendly for mass deployments due to familiarity of end users with smartphone cameras. But it is not the only tool for global diffusion and ADD decision system is a multipartite combination of many different sources of data, a few of which is discussed in this section.

The core of this section aims to explain the central significance of two issues, the importance of which cannot be overemphasized: [a] binding and [b] detection. Without binding the target (in this case SARS-CoV-2) there is no detection. Unless detected, the individual cannot be isolated (undetected but infected individuals are a threat by spreading CoVID-19). A different *modus operandi* for binding and detection is discussed in APPENDIX X.

Binding data in any scientific pursuit or research publication will require titration data from precision tools (for example, biolayer interferometry and surface plasmon resonance) to show that A and B are binding with measurable specificity. Scientific analysis of the kinetics of interaction between the aptamer and protein (target analyte) is essential for science. But, this data may NOT predict binding kinetics in a pragmatic use case, for example, binding clinically relevant target protein in saliva (sputum sample from humans and animals). Lower cost tools amenable to mass implementations must be tested with target analyte in a mix that uses non-invasive samples, for example, saliva (using blood or serum is ineffective, see Figure 23b).

If the sensitivity of the optical system (discussed earlier) fails to be sufficient (Fig 1), the next option is to test signal transduction using electrochemical impedance spectroscopy (EIS). Therefore, titration of binding must be performed with low cost Laser Inscribed graphene (Turbostrat) Sensors (LITS) using EIS for signal acquisition. Optimizing signal over noise and acquiring the EIS data in a smartphone-based system for analysis and visualization holds the key to potential for large-scale deployment in environments devoid of access to labs and hospitals (underserved communities, home use, schools, small-medium businesses, farm laborers, logistics personnel, waste collectors, the elderly).

Aptamers or antibodies or ACE2 attached to LITS (APLITS, ABLITS, ACLITS, respectively, see section 6, Figure 3, page 12) in conjunction with EIS emphasizes one low-cost approach for large scale deployment of tools for detection and lead to isolation, preferably early (TETRIS – test, treat, isolate).

True efficiency of mass testing may emerge if “testing” is an “IoT” lifestyle. The test, *hidden in plain sight*, will make users oblivious of its existence. For example, building access and security is now woven seamlessly into our lives through the ease of carrying near field communications (NFC<sup>377</sup>) devices in our clothing, wallets, key cards or embedded in mobile devices / smartphones.

But, the practice of mass testing will be impotent if limited to affluent nations because of antigenic drift (see APPENDIX III and APPENDIX IV). Any organism, including the SARS-CoV-2 virus, must replicate in order to mutate. Antigenic drift enables actively replicating viruses to make mistakes in its replication process and generate mutants<sup>378</sup>. We are observing global spread of SARS-CoV-2 mutants from geographically distant nations to far corners of the world. We must reduce the opportunity for the virus to replicate if we hope to restrict new mutants/variants of SARS-CoV-2.

The lack of drugs<sup>379</sup> makes it even more imperative that we find multiple ways to **reduce new infections**. If we can stop the virus from infecting the uninfected then the virus will not have access to new hosts and progeny viruses will not be produced in yet another host. ***Thus, by controlling the spread of transmission of the virus we will reduce its ability to mutate.*** We can only contain the spread if we know who is infected and then isolate the person. To accomplish this quintessential task, globally, we need low-cost tools because deployment must be global. It is toothless to pursue detection in affluent nations only because the more populous developing and under-developed nations will continue to serve as fertile grounds for the virus to create mutants which will spread due to travel involving humans, animals, goods, and global supply chain related essential services dependent on less affluent nations.

The herculean task of detection and isolation to reduce transmission, globally, cannot begin in the field without rigorous R&D. A few key processes for aptamers (APLITS) are outlined as follows:  
 [1] (Pharmaco)Dynamics of DNA aptamers that bind with efficiency (target access?), specificity and reproducible (quantitative) affinity. Which SARS-CoV-2 proteins are targets? (Pharmaco)Kinetics of binding, in terms of equilibrium dissociation constant [ $K_D$ ], where a smaller  $K_D$  (the ratio  $k_{OFF} / k_{ON}$ ) indicates greater binding affinity of the ligand for its target (SARS-CoV-2 target protein)], between aptamers and preferred target vs “nearest neighbor” protein competitor, based on *shape* and amino acid sequence (larger  $K_D$  value indicate weaker binding), must show reproducible and statistically significant difference of at least one order of magnitude. The design of the binding *assay* remains to be determined but must include kinetic data using different principles (surface plasmon resonance<sup>380</sup>, biolayer interferometry<sup>381</sup> and electrochemical impedance spectroscopy<sup>382</sup>) to enable incisive data analysis.



Rigorous determination of dissociation constant and association constant (reciprocal of dissociation constant) is the bedrock of biochemistry (quantitative, reproducible) at the heart of chemical equilibrium<sup>383</sup> which is the essential pharmacokinetic pillar to determine which aptamers and protein targets may be potentially useful for which purpose.

It remains to be explored if lectins<sup>384</sup> and *glycan specificity* (Figure 20) must inform our outcome (data). Will including lectin sensors<sup>385</sup> (LELITS as parallel positive controls, see APPENDIX VIII) enhance data accuracy (reduce false negatives)? Will Spike glycan shield influence the interaction which may lead to data uncertainty by (occasionally) perturbing epitope specific binding with the aptamers?

[2] Test DNA aptamer binding to sensor material (laser inscribed graphene, Au/Pt nano-materials, etc). Establish metrics for stability of binding, using linkers to attach aptamers (covalent), using lectins and/or chemical tails, eg, poly(N-isopropylacrylamide).

[3] Test stability of conjugation of nano-dots with aptamers (cadmium, carbon). Is the optical signal (with/without protein) vs noise reliable and reproducible under different conditions? Explore other signal transduction techniques (electrochemical impedance spectroscopy, surface plasmon resonance).

[4] Repeat [3] with aptamers linked/adsorbed on sensor material surface to choose the best outcome from materials and transduction. Is the signal vs noise quantitative or qualitative? Prefer quantitative because qualitative offers only “yes/no” (with reservations) and influenced by limit of detection (LoD).

[5] Combine outcomes. If signal over noise is statistically significant ( $P < 0.001$ ) after data acquisition by a mobile platform, then we have accomplished the scientific rigor to fuel the engineering basis for creating tools and applications for detection/diagnostics/screening/surveillance. The latter may involve innovation in engineering design to determine form factors, product development (breathalyzer) and imagination to transform the idea of USB connected modular, mobile, adaptable, *sensor-on-a-chip* to link to smartphones (any USB port) to create the (hypothetical) surveillance tool: **molecularphone**.

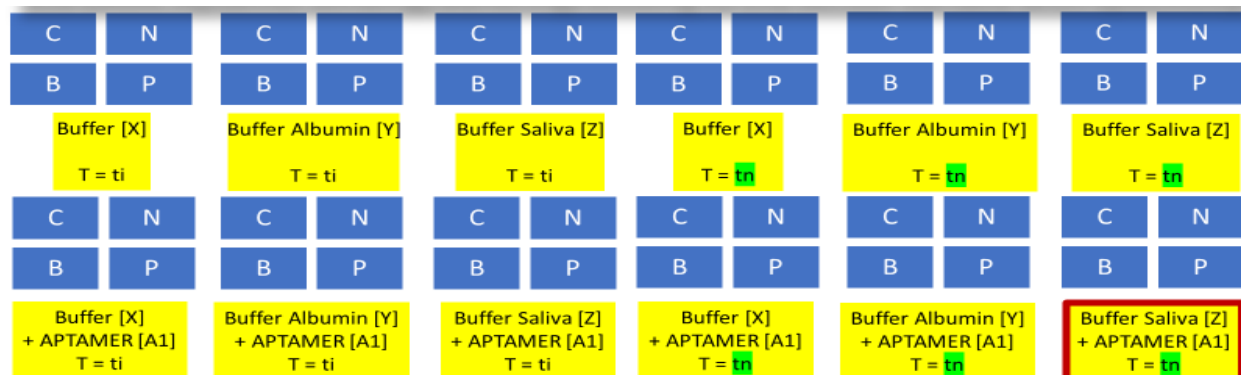
#### EXPERIMENTAL TEMPLATES: BIOCHEMISTRY AND BIOSENSORS (APLITS, ABLITS, ACLITS)

The most informative data for application purposes will be the reproducible data from the assay (see EXPERIMENTAL TEMPLATE 1 – bottom row, last column) where a pre-titrated concentration of saliva-mimicking substitute (indicated as Buffer Saliva [Z]) will generate consistent EIS signals under conditions which includes an aptamer at a titrated concentration (indicated as +APTAMER [A1]) binding to sensor surface (LITS) at time  $T = t_n$ . Each experimental panel must include different types of data plots and EIS measurements: [a] cyclic voltammetry (C) [b] Nyquist plot (N) [c] Bode plot (B) [d] phase shift (P). Variables include titrating for time of incubation ( $t_i$  through  $t_n$ ) and different aptamers (different DNA sequences) in concentrations (A1 through A<sub>n</sub>). Specificity of aptamer binding to LITS (APLITS) must be rigorous with each data set in triplicate analyzed critically for consistency.

It is imperative to understand and convey precisely the meaning of the data which is acquired in these plot and graphs. Cyclic voltammetry (C, CV) and Nyquist plot (N, NP) both rely on instruments which use software treatment of data including filtering options and chemometric processing prior to generating the plots. This “manufactured” data (x-axis) introduces potential for error-prone answers.

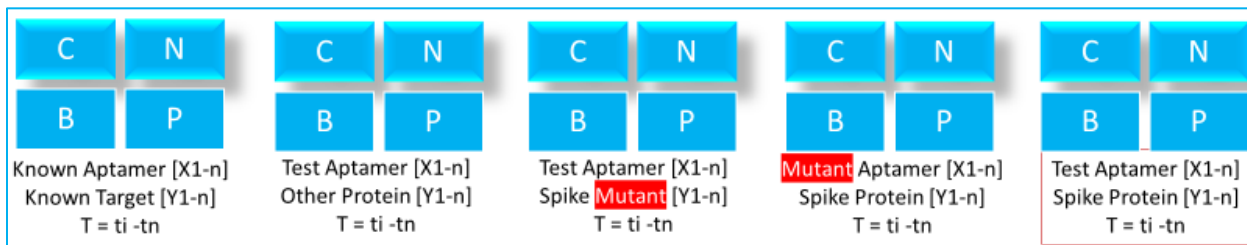


Bode Plot (B, BP) and Phase Shift (P, PS) data are actual measurements of frequency, resistance and impedance values which are presented without data processing by embedded software in devices.



EXPERIMENTAL TEMPLATE 1 – Expected EIS data to demonstrate without any reasonable doubt that the aptamer used in the assay binds (adsorption) to LITS (sensor surface) with specificity. Titration of aptamer concentration (A1 through An) is implied but not explicitly shown in the template cartoon.

Armed with bonafide APLITS, the most useful non-IRB data (internal review board, IRB, permission is necessary for using human saliva samples) for clinical application purposes will be the reproducible data from the last panel (right) in EXPERIMENTAL TEMPLATE 2 where a pre-titrated concentration of saliva-mimicking substitute generates consistent EIS signals under conditions which includes test aptamer sensor (APLITS) and SARS-CoV-2 Spike protein (or variations of the viral Spike protein, such as Spike S1 only, Spike S2 only, combined Spike subunits S1+S2, select peptides based on Spike protein) at a pre-titrated concentration incubated for optimized time (T, t).



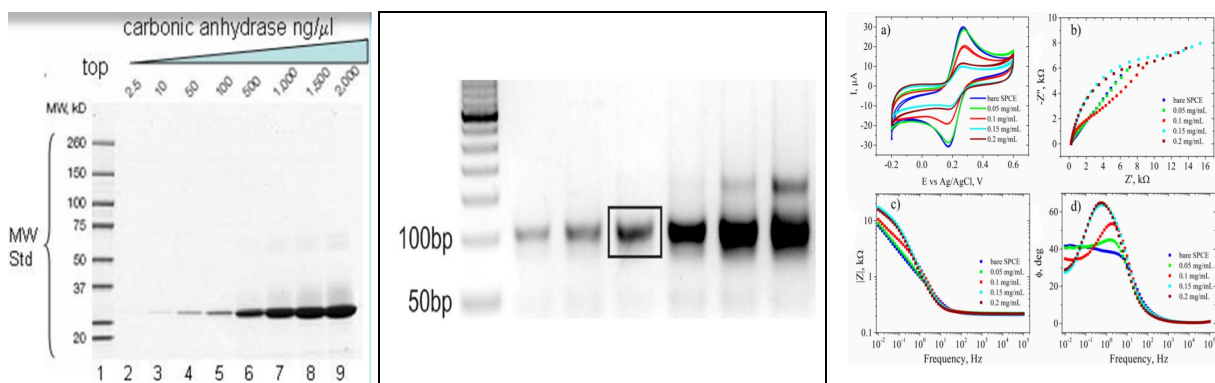
EXPERIMENTAL TEMPLATE 2 – Expected EIS data to demonstrate without reasonable doubt that the aptamer (APLITS) binds specifically to the target SARS-CoV-2 Spike protein. All panels except the last panel are controls. The value of the data in the last panel is null and void (unscientific) without data from controls. All experiments (panels) shown in the templates are performed as a set. Each panel indicates expected EIS data: [a] cyclic voltammetry (C) [b] Nyquist plot (N) [c] Bode plot (B) [d] phase shift (P). Variables include titration for time of incubation ( $t_i$  through  $t_n$ ), different APLITS (aptamer sensors with different sequences of single stranded DNA) in concentrations (X1 through Xn) and relevant proteins (concentrations Y1 through Yn). If successful, data from the last panel (right) may serve as the starting point for testing human saliva, obtained with permission from the internal review board (IRB).

CV	NY	CV	NY	CV	NY	CV	NY	CV	NY	CV	NY
BO	PS	BO	PS	BO	PS	BO	PS	BO	PS	BO	PS
Buffer [X]		Buffer Albumin [Y]		Buffer Saliva [Z]		Buffer [X]		Buffer Albumin [Y]		Buffer Saliva [Z]	
T = ti		T = ti		T = ti		T = tn		T = tn		T = tn	
CV	NY	CV	NY	CV	NY	CV	NY	CV	NY	CV	NY
BO	PS	BO	PS	BO	PS	BO	PS	BO	PS	BO	PS
Buffer [X] + Antibody [A1]		Buffer Albumin [Y] + Antibody [A1]		Buffer Saliva [Z] + Antibody [A1]		Buffer [X] + Antibody [A1]		Buffer Albumin [Y] + Antibody [A1]		Buffer Saliva [Z] + Antibody [A1]	
T = ti		T = ti		T = ti		T = tn		T = tn		T = tn	
CV	NY	CV	NY	CV	NY	CV	NY	CV	NY	CV	NY
BO	PS	BO	PS	BO	PS	BO	PS	BO	PS	BO	PS
Buffer [X] + Antibody [An]		Buffer Albumin [Y] + Antibody [An]		Buffer Saliva [Z] + Antibody [An]		Buffer [X] + Antibody [An]		Buffer Albumin [Y] + Antibody [An]		Buffer Saliva [Z] + Antibody [An]	
T = ti		T = ti		T = ti		T = tn		T = tn		T = tn	

EXPERIMENTAL TEMPLATE 3 – Titration to optimize antibody binding to LITS (ABLITS). Template also applicable to titrating ACE2 receptor with LITS to create ACLITS. Only panels with pre-IRB saliva substitute and antibody (concentrations A1 through An) are true experimental panels. All other panels are controls. Without controls, *each time*, data from true experiments are unacceptable. Each rectangle (blue) represents expected data from: [a] cyclic voltammetry (CV) [b] Nyquist plot (NY) [c] Bode plot (BO) [d] phase shift (PS). Positive Control with pre-tested and pre-calibrated known antibody-antigen is included. EIS data must demonstrate without any reasonable doubt that the antibody (or ACE2 protein) used in the assay, binds (adsorps) to LITS surface with specificity (ABLITS, ACLITS). Titration of antibody (or ACE2) concentration (A1 through An) and time of incubation (T = ti through tn) is shown.

CV	NY	CV	NY	CV	NY	CV	NY	CV	NY
BO	PS	BO	PS	BO	PS	BO	PS	BO	PS
Other Ab (mAb) Known Antigen [N0...Nn]		Spike Antibody (mAb) Other Protein [N0...Nn]		Spike Antibody (mAb) Spike Mutant [N0...Nn]		Other Antibody (mAb) Spike Protein [N0...Nn]		Spike Antibody[A] Spike Protein [N0...Nn]	
T = ti...tn		T = ti...tn		T = ti...tn		T = ti...tn		T = ti...tn	

EXPERIMENTAL TEMPLATE 4 – EIS data template applicable to test ABLITS and ACLITS. First four panels are controls. Last panel (right) is the outcome of interest (irrelevant without data from all four controls). N0 refers to zero concentration. Different monoclonal antibodies (mAb) may be tested (mAb preferred over polyclonal antibodies, pAbs). Different versions of SARS-CoV-2 Spike protein (S, S1, S2, mutants) as well as Spike-derived synthetic peptides may be tested. Combinations of Spike mAb and Spike proteins/peptides may be tested to probe epitope specificity or overlapping epitopes in view of Spike protein mutants and Spike protein antibodies from previous SARS-CoV infections (SARS, MERS).



EXPERIMENTAL TEMPLATE 5 – Characterization of proteins (left panel) and DNA (middle) using gel electrophoresis (or other techniques) to show molecular weight distribution and concentration gradient. Right – EIS (unrelated examples): [a] cyclic voltammetry [b] Nyquist plot [c] Bode plot [d] phase shift.

## APPENDIX II – APTAMER-AS-A-DRUG (AAAD) FOR THE LESS AFFLUENT UNDERSERVED WORLD?

Can aptamers serve as alternates or supplements to traditional vaccines? Small molecule-like "inhibition" by aptamers (*in vivo*) may offer low-cost (?) therapeutic paths<sup>386</sup> for less affluent nations. However, the general mechanism of action of small molecules versus aptamers may be quite different. Aptamer binding to a specific region of a target protein may induce some changes, perhaps a change in conformation, but it *may not* disable the 'active site' of the target protein (if it has enzymatic functions) or dissuade the protein from its usual activities even if it suffices to reduce its efficiency and/or efficacy.

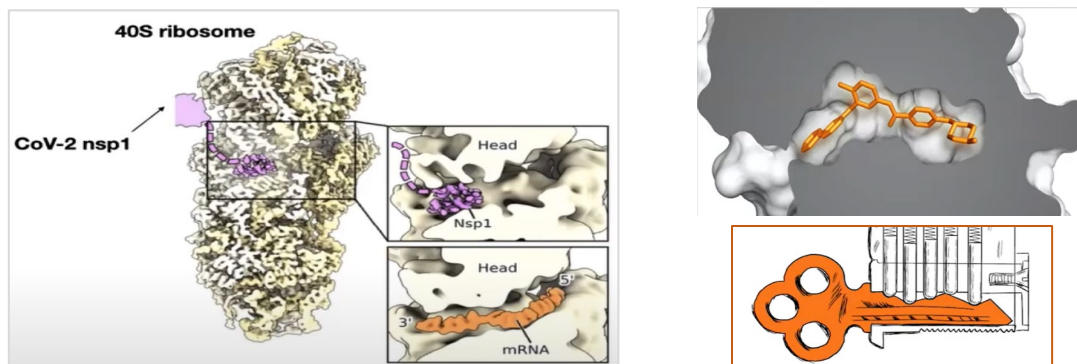
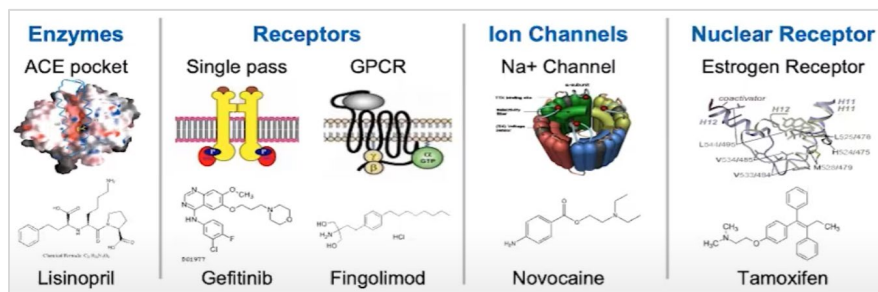


Figure 24: Natural and synthetic organic small molecules are important pharmaceuticals (top panel). SARS-CoV-2 Nsp1 (Thoms *et al*, see Fig 7) acts as a small molecule to "fit" in the "groove" of the 40S ribosome (bottom, left) to arrest host translational systems, as shown in the models<sup>387</sup> (bottom, right).

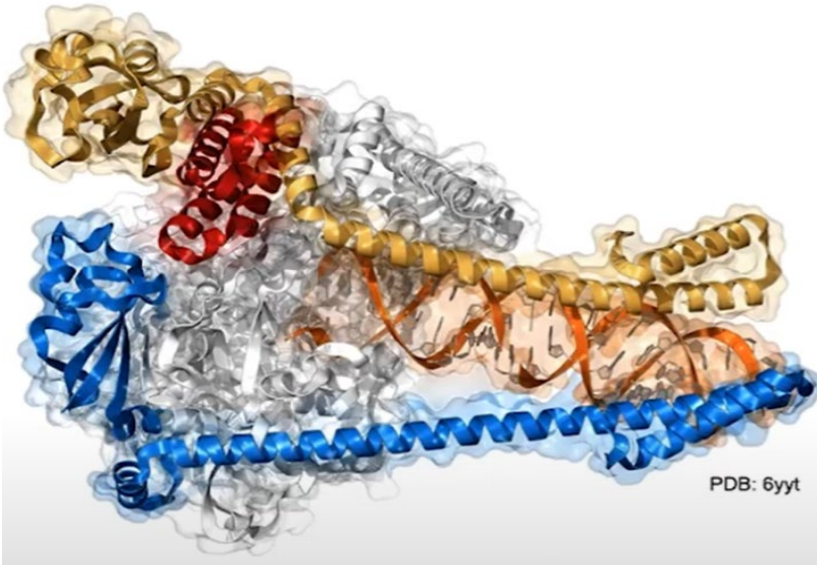
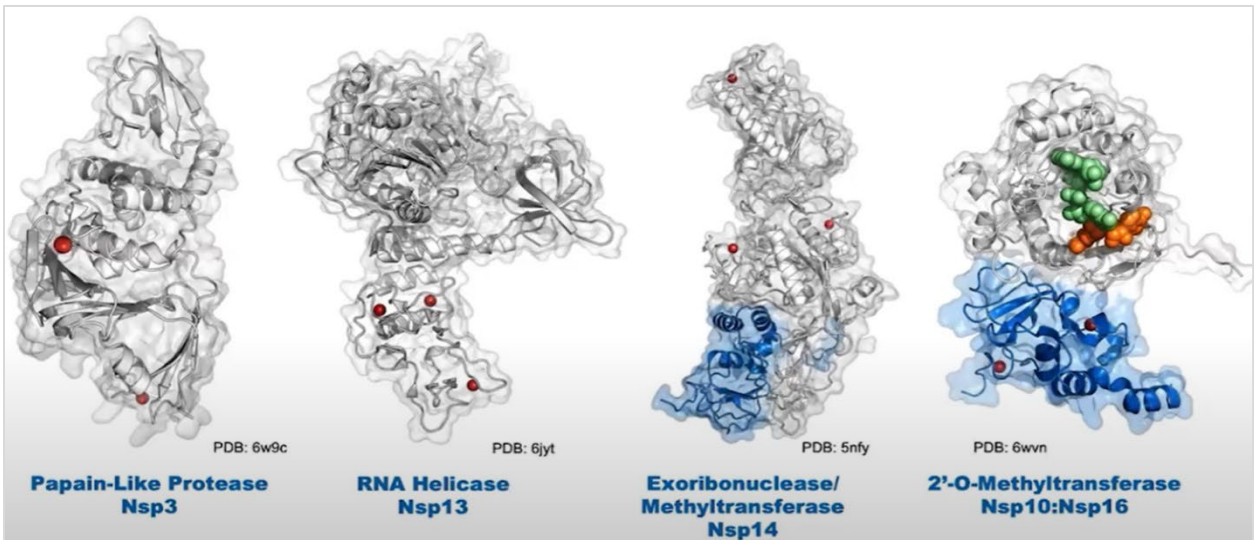
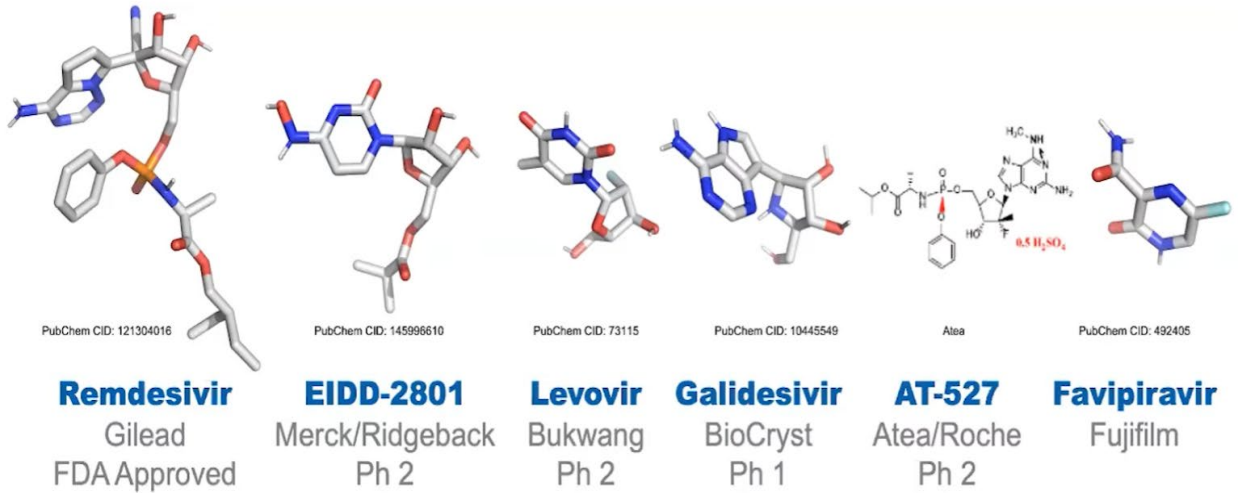


Figure 25: RdRp (left panel) may be the primary target for aptamer-as-a-drug (AAAD). Remdesivir (middle panel) is ineffective for SARS-CoV-2 RdRp (other small molecules are being tested). Additional targets (bottom panel) may include proteases and the individual proteins of the RdRp (RNA-dependent RNA polymerase) complex. Source: Bradner, James (2020).





Uncompromising rigor of (pharmaco)dynamics and (pharmaco)kinetics [step 1] tested under a spectrum of conditions (pH, salinity, salivary enzymes, blood, plasma) will determine if AAAD is even a possibility. A positive outcome (small number of aptamers binding select viral proteins with stability, **epitope** specificity and selectivity in a variety of body fluids, pH range) may be the first indication that it may be worthwhile to test AAAD candidates for *in vivo* activity (assay to test strength of inhibition?).

Initial interactions in buffer and laboratory conditions are good indicators but it *cannot predict* what will happen *in vivo* because aptamer-protein binding is determined by the *3D shape* (secondary structure) the aptamer (single stranded DNA string) will assume (under a set of conditions) and protein (epitope) binding, as a *consequence of that structure* (shape). To partially mimic *in vivo* conditions, testing aptamers in cell culture may be the ‘quick and dirty’ first choice. The *dynamic conformation* of aptamers under various conditions influences the binding to target proteins. It is a source of *uncertainty*.

Extrapolating results from *in vitro* tissue culture and *in vivo* animal models (next mandatory step) to humans is neither prudent nor a *bona fide* scientific process. But step-wise success may help to justify the path forward. Any one of many factors could be the *nail on the coffin* of the AAAD idea. These factors include but may not be limited to stability, bio-availability, delivery, absorption, permeability, distribution, metabolism, elimination, cross-reactivity, general cytotoxicity, organ specific toxicity (for example, cardio toxicity, renal toxicity, neurotoxicity, blood-brain barrier). Finally, only unequivocal success in most stringent human clinical trials may help to transform the idea of AAAD into reality.

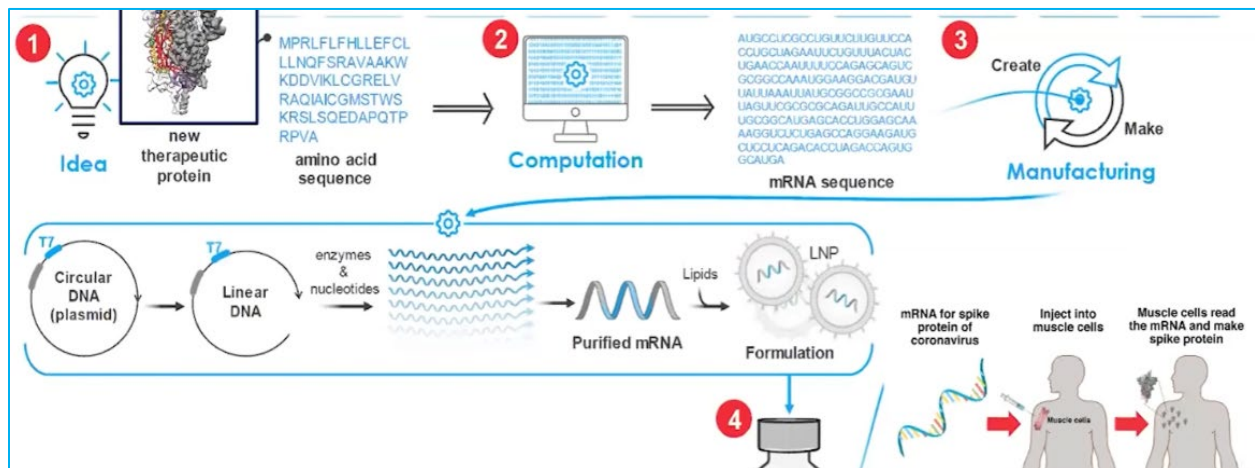


Figure 26: AAAD could use lipid nanoparticle delivery as in the Moderna mRNA-1273 (1273 amino acid SARS-CoV-2 Spike protein) vaccine protocol<sup>388</sup> (LNP formulation by Langer *et al*). Delivery of AAAD using LNP may improve absorption, for example, if used as a topical nasal spray to prevent spread of infection. All mRNAs in human cells are encoded by 2% of the entire genome. 98% of the genome is transcribed into cellular RNAs whose activities still remain to be discovered (new RNA therapies?). Thus, most of the human genome does not code for proteins, most of the disease-causing genetic variants are located in non-coding regions, most transcripts produced by genes are non-coding and non-coding genes may not have a functional annotation (their biological role is unknown). But, clues about the biological role of long non-coding RNAs in the human genome are beginning to emerge<sup>389</sup>.

## APPENDIX III – THE MOLECULAR BASIS OF DISEASE

### Variability in Immune Response to SARS-CoV-2 Infection: Conundrum, Conjecture, Common Themes

The spectrum of immunological diversity<sup>390</sup> presented by CoVID-19 reinforces the value of thinking broadly<sup>391</sup> and thinking differently. It may not be unwise to forward hypothesis or conjectures which may or may not provide clues to understand or unravel the biological basis of this conundrum. On the other hand, based on common themes in molecular biology and genetics, perhaps what we are observing is not a conundrum at all. This discussion brings together what we think we may know.

Let us commence with the observation that bacteria belonging to even one strain, for example, *Escherichia coli* (O104:H4, O157:H7, O121) if sequenced (DNA genome), will reveal that their genomes, in terms of DNA sequence are not exactly identical. One explanation based on the molecular biology of CRISPR (clustered regularly interspaced short palindromic repeats) indicates acquisition of new spacer sequences<sup>392</sup> from foreign DNA necessary to adapt CRISPR-Cas<sup>393</sup> system to confer adaptive immunity. The human genome<sup>394</sup> revealed our genomes<sup>395</sup> are similar but not identical (even between twins), due to unequally distributed single nucleotide polymorphisms (SNPs) in coding and non-coding sequences.

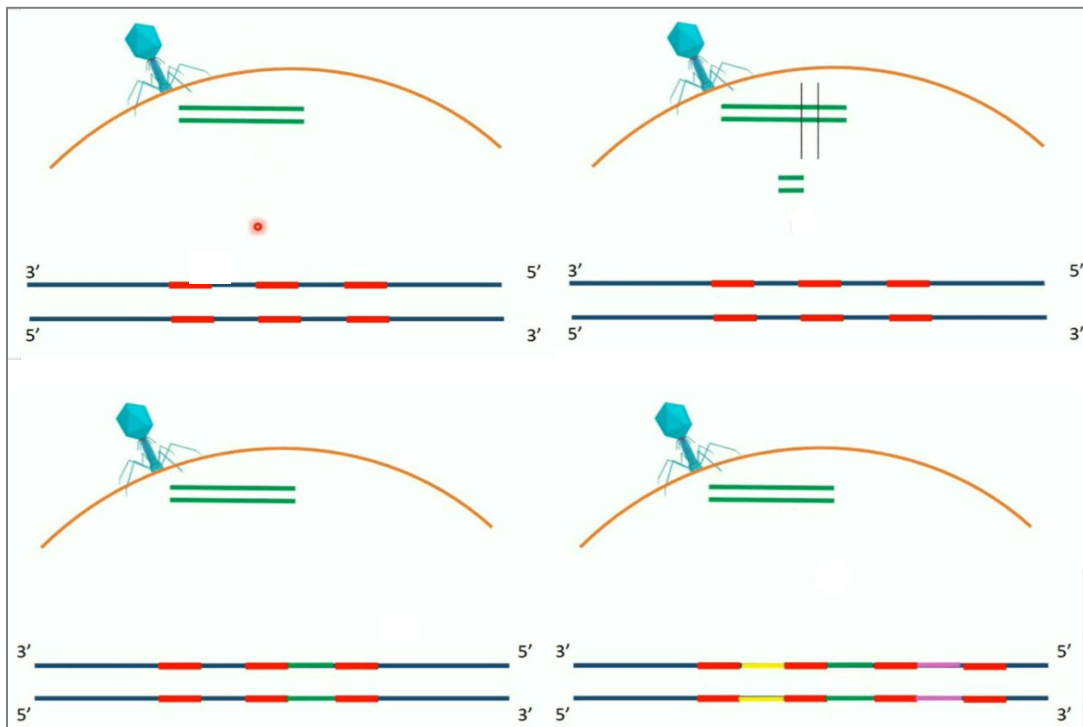


Figure 27: Acquired spacer sequences in bacteria is one reason why genomic sequences differ even within the same strain. What is the impact of the integration and the heterogeneity of the type and number of spacers on bacterial gene expression, protein expression, physiology and metabolism?

The conundrum about the variation in human immune response to CoVID-19 may be *natural* if we consider that each human is genetically unique. We have known for 2 decades that approx. 3 million nucleotides (including about 2 million SNPs) are different between our genomes (why we are genetically unique) which contain ~6.4 billion base pairs<sup>396</sup> (6.4 billion nucleotides - A, T, G, C – in diploid human genome or 3.2 billion base pairs, monoploid). These differences are significant when evaluating drugs. Hence, genetic<sup>397</sup> stratification of humans in clinical trials is now routine. The efficacy, effectiveness or resistance<sup>398</sup> of the same drug may be quite different between individuals. The latter partially explains the observed variation<sup>399</sup> in immune<sup>400</sup> responses<sup>401</sup> to CoVID-19. SARS-CoV-2 induces a multi-factorial<sup>402</sup> physiological<sup>403</sup> cascade of events<sup>404</sup> involving systems<sup>405</sup> and network of factors<sup>406</sup> linked to genetic predispositions and co-morbidities which may influence phenotypic expression, to different degrees, directly or indirectly, in each human. The 823 epitopes<sup>407</sup> mapped in the SARS-CoV-2 proteome, were not all equally recognized by antibodies in all individuals, indicating the complexity of stratification.

It is not only genomics but **regulation** of gene<sup>408</sup> expression<sup>409</sup> (transcriptomics), proteomics and metabolism (rates of anabolism and catabolism, metabolomics). Omics may be affected by epigenetic factors (food, air, water, environment<sup>410</sup>) and immune cell dynamics are modulated by microbiomes<sup>411</sup> (viromes). Taken together, these factors are likely to affect detectable symptoms and clinical outcomes.

In humans, multiple processes and DNA sequences flanking the immunoglobulin genes (V, D, J) influences the genetic rearrangement of the gene segments followed by somatic hypermutation<sup>412</sup> which contributes to the great diversity of our immunoglobulin repertoire. It is one of the key tools available to the immune system to design antibodies and respond appropriately upon presentation of an antigen. Somatic mutations vastly differentiate and enhances the scope of response which may be otherwise restricted if the system were to depend only on the inherited genetic components (germ line theory<sup>413</sup>).

The machinery available to antibody-producing cells for executing somatic changes in genes and gene expression is an evolutionary process. Creative application of this machinery may generate quite a variation in phenotypic response in CoVID-19. Somatic reshuffling in combination with differences between SNPs may result in an inordinate number of different permutations and combinations. Hence, the spectrum of CoVID-19 symptoms. SARS-CoV-2 proteins<sup>414</sup> may induce somatic hypermutation in cells and tissues to result in perturbation of homeostasis. Epigenetic modifications triggered by sequence cassettes<sup>415</sup> may affect basic processes (transcription, translation, post-translational modifications, etc.).



Figure 28: Discovery<sup>416</sup> of transposons in maize<sup>417</sup> revealed that segments of genes “jump” from one genome to another (see left, kernel colors<sup>418</sup>). Variations of this “dynamic” concept are found in influenza<sup>419</sup>, Trypanosomes<sup>420</sup>, Plasmodium<sup>421</sup> and other organisms. SARS-CoV-2 may hijack this mechanism, create *ad hoc* changes and alter therapeutic targets.

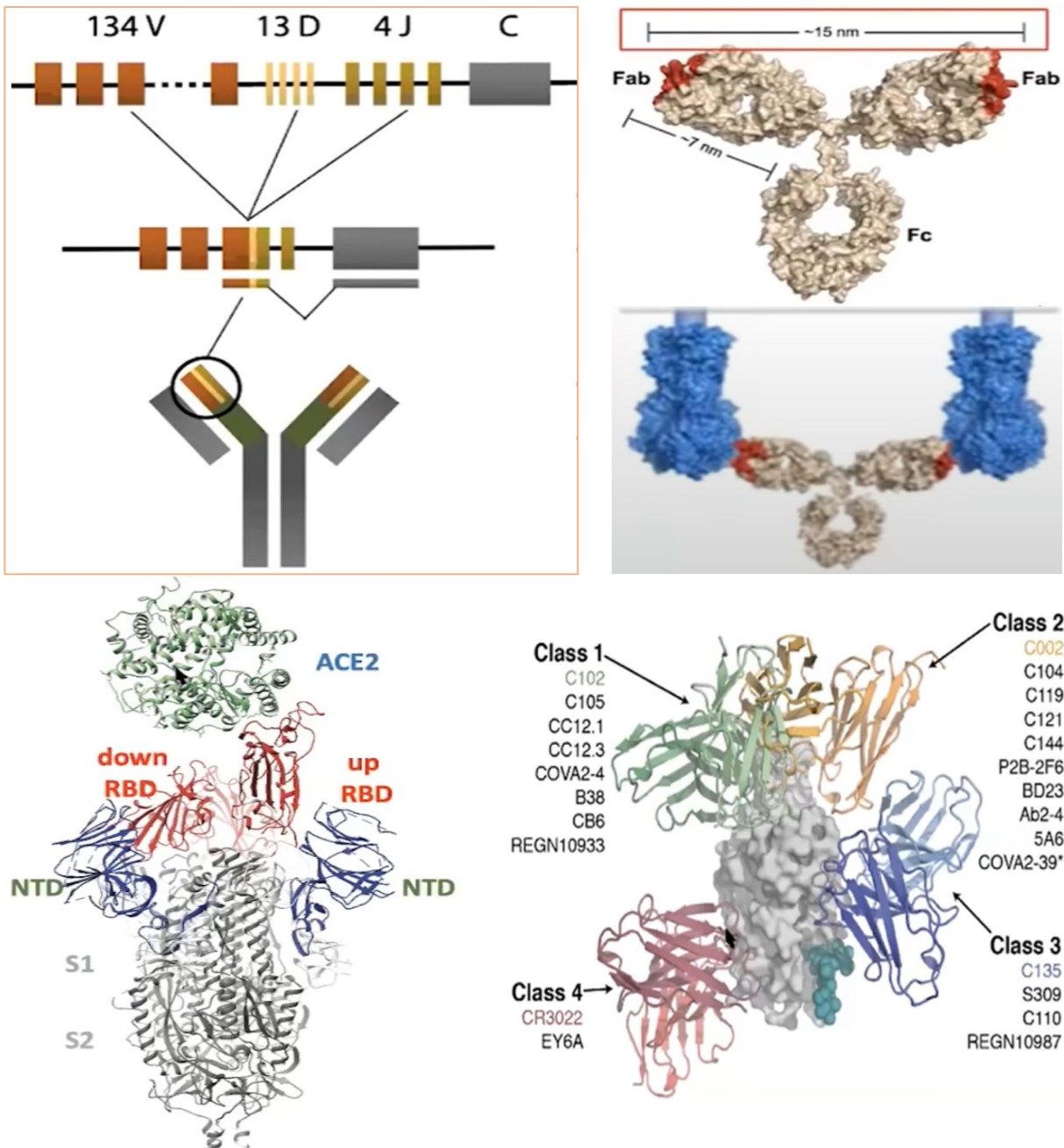


Figure 29: Variations in antibodies and binding affinities<sup>422</sup> are due to VDJ (upper, left) recombination (combinatorial diversity  $\sim 2.5 \times 10^6$  & junctional diversity  $> 10^{14}$ ) and somatic mutations (Tonegawa *et al*, 1974) mainly in B cells ( $\sim 10^{11}$  in humans). Understanding recurrent features of antibodies<sup>423</sup> binding to SARS-CoV-2 Spike protein (lower, left) helps to identify antibodies with therapeutic<sup>424</sup> potential. Four classes of human neutralizing monoclonal antibodies are shown (lower, right). The virus will mutate to evade mAbs, e.g., E406W [Glu<sup>406</sup>→Trp] escapes<sup>425</sup> binding to known antibodies. On the other hand, antibodies from pre-SARS-CoV-2 (2003) neutralized<sup>426</sup> SARS-CoV-2 due to overlapping epitopes.



## BACK TO BASIC SCIENCES

Molecular dissection of the SARS-CoV-2 genome<sup>427</sup> to delineate the functional role of each viral protein is fundamental. Testing infectivity of single gene knockouts (and multi-gene combinations) may be one essential step. Physiological exploration of the >332 human proteins (Gordon *et al*, 2020) which may interact with SARS-CoV-2 proteins is crucial. A global collaboration may be necessary to analyze the *value of each nucleotide* in the ~30KB single stranded RNA genome<sup>428</sup> of SARS-CoV-2 because even a single amino acid structural change can have *profound impact*<sup>429</sup> on function. Precedence for this rigor may have started with in the incisive minutiae of mutagenesis<sup>430</sup> applied to beta lactamase. Thoughtful design and detailed execution of this megaproject may provide clues to what constitutes virulence<sup>431</sup>. Metrics of virulence is pivotal to deconstructing its cryptic complexity and reconstructing the role of molecular medicine in healthcare for humans and animals infected with virulent agents.

In the interim, undergraduates in molecular biology may undertake the theoretical analysis of SARS-CoV-2 proteins. For each known viral protein, it may be useful to list expected modifications in primary amino acid sequence (if any) due to changes in the 3rd position of the RNA codon (Fig 26, L). For example, if AGU mutates to AGC, the amino acid Serine is still the same (silent mutation) due to the degeneracy of the triplet<sup>432</sup> genetic code. If AGU/AGC mutates to AGA/AGG then Serine is replaced by Arginine. Using the Ramachandra Plot (Fig 26, R) students may explore which values of the  $\psi$  and  $\phi$  angles are possible for *that* amino acid residue which *changed* in the viral protein. Can the change in the codon create an amino acid substitution which can influence the conformation of the viral protein? Structure and function are inextricably integrated in biological activity. This exercise may uncover targets for experimental analysis and predict which changes in the codon and primary sequence, may be of consequence with respect to interaction between viral proteins and their putative human targets.

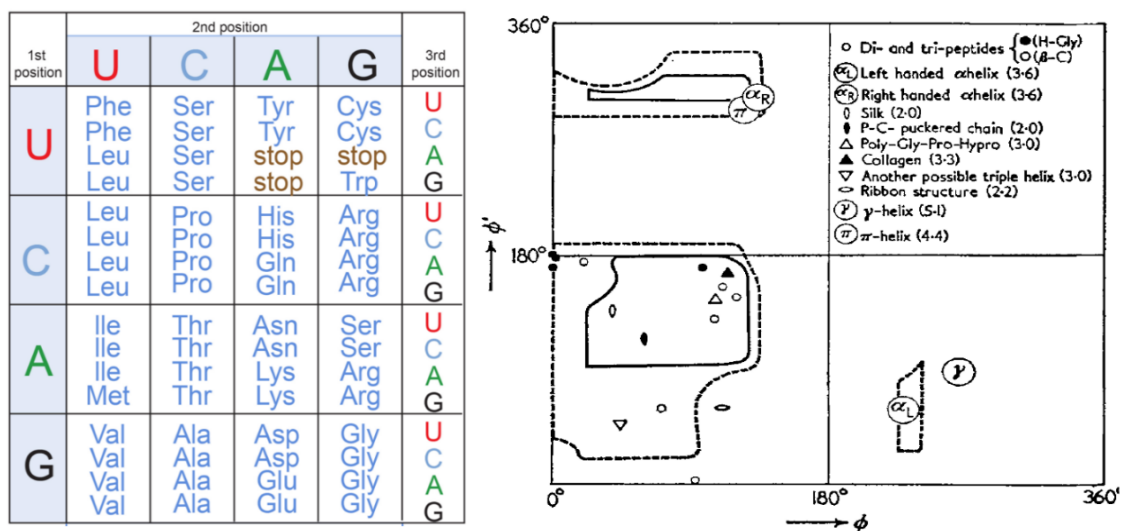


Figure 30: RNA Codon<sup>433</sup> table<sup>434</sup> (L). Ramachandran Plot<sup>435</sup> of allowed values of the  $\psi$  and  $\phi$  angles (R).

The diversity and acuity of symptoms suggests SARS-CoV-2 proteins may have access to genetic circuitry<sup>436</sup> of developmental<sup>437</sup> clocks<sup>438</sup> and may be “playing” with master<sup>439</sup> switches<sup>440</sup> or re-wiring<sup>441</sup> circuits or “time<sup>442</sup> spoofing” the expression in some form<sup>443</sup> to modulate<sup>444</sup> differentiation. It is possible that developmentally re-programmed genetic circuits or regressively differentiated cells may express proteins and/or *other molecules* which are not in our ‘*data dictionary*’ because it is not a part of our *differentiated* physiology. Therapeutic targets and approved drugs<sup>445</sup> may not be effective<sup>446</sup> because the virus may be creating their own *decoy*<sup>447</sup> molecules to wreak havoc. The virus appears to be *changing our targets* to escape host defense (immune system) and offense (drugs) as well as perturbing cellular signals for biomarkers associated with CoVID-19 mortality (IFN- $\alpha$ <sup>448</sup>, IL-18<sup>449</sup>, IL-10<sup>450</sup>).

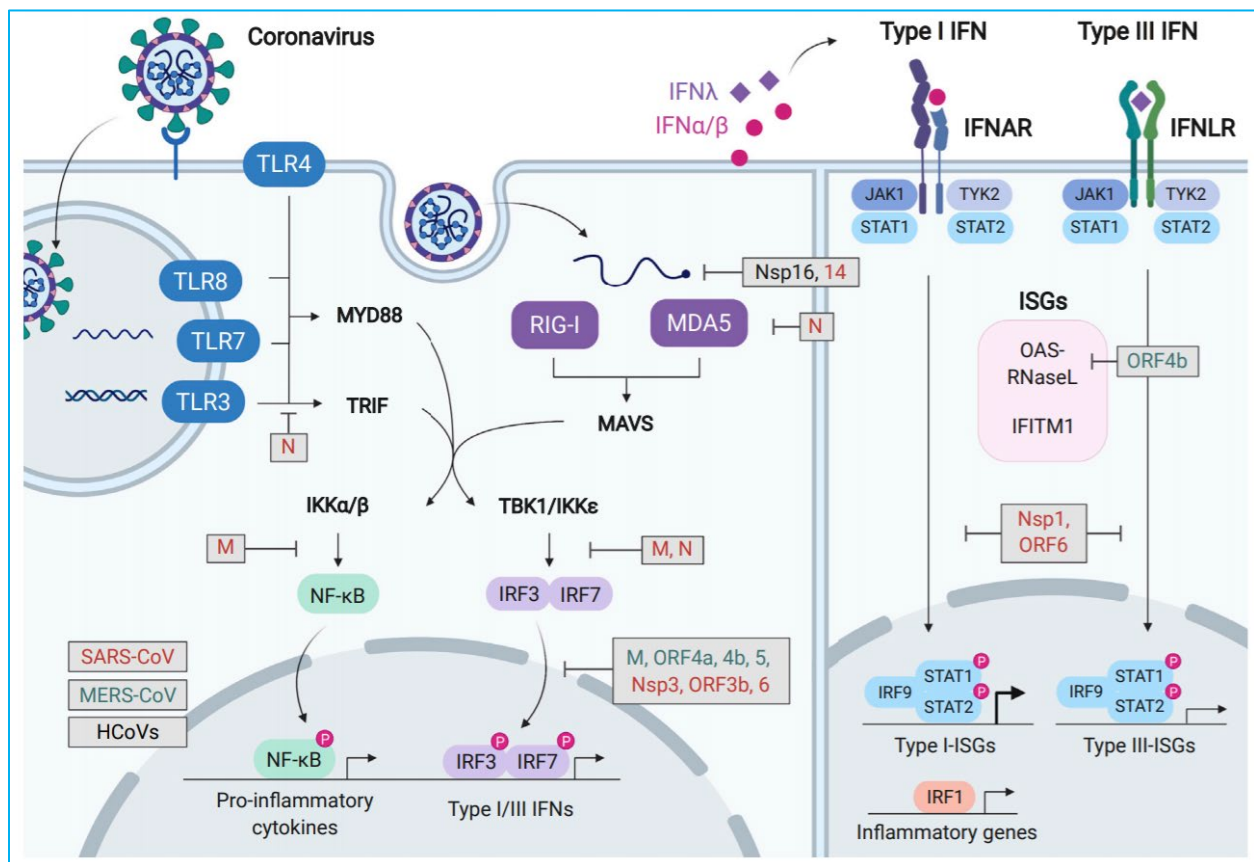


Figure 31: Immune evasion strategies<sup>451</sup> by coronaviruses include antagonization/disruption of various pathogen recognition receptors, TLRs<sup>452</sup> (TLR3, TLR4, TLR7, TLR8; blue) and RLRs<sup>453</sup> (RIG-I, MDA5; purple), transcription factors nuclear factor kappaB (NF- $\kappa$ B<sup>454</sup>) and interferon regulatory factors 3 and 7 (IRF3, IRF7) which are (*normally*) supposed to stimulate the production of pro-inflammatory cytokines and type I and III interferons (IFNs), respectively. IFNs (autocrine and paracrine secretion) induce expression of interferon-stimulated genes (ISGs) via the JAKSTAT signaling pathway.

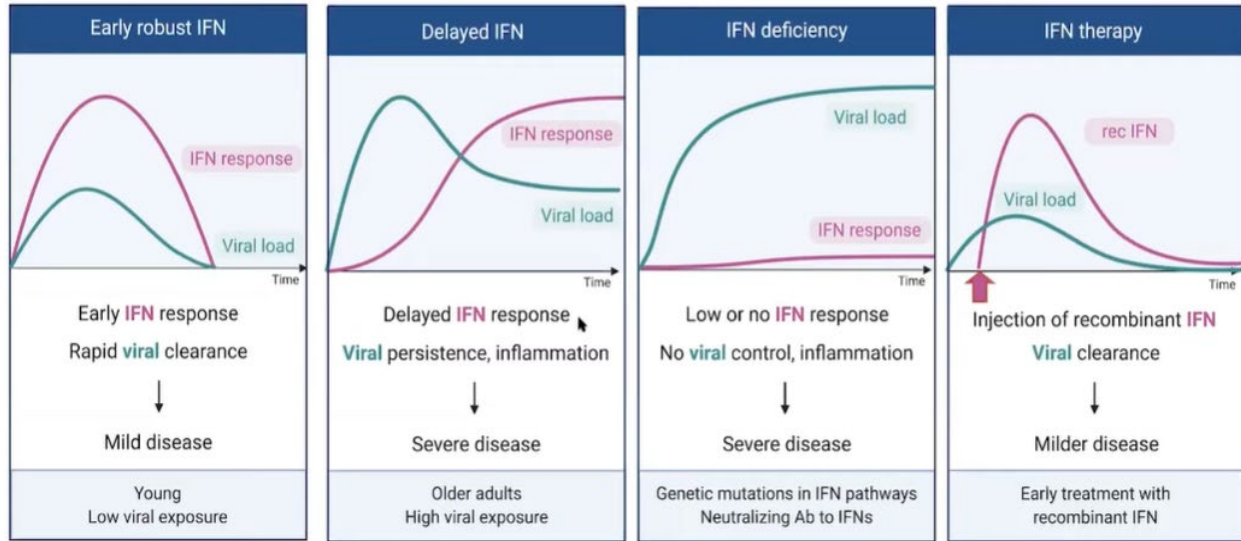


Figure 32: Promise of interferon<sup>455</sup> and its role<sup>456</sup> as a potential therapeutic agent. A reason to believe<sup>457</sup>? Hypothetical Interferon Therapy illustration<sup>458</sup> provided by Kizzmekia Corbett, NIH.

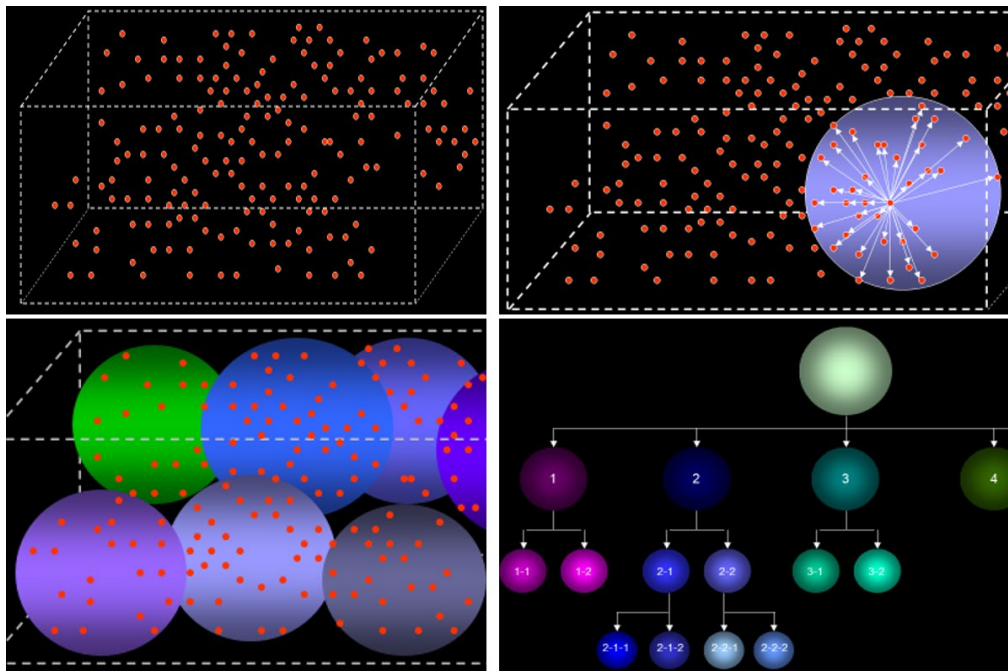


Figure 33: With >50 million infected by SARS-CoV-2 (actual number may be >500 million) the clinical deluge to deal with CoVID-19 patients may leave little time to pursue molecular stratification. Access to vaccines, neutralizing<sup>459</sup> monoclonal antibodies and promise of interferon therapy (Figure 32) could make this discussion moot. But, currently the best we can expect is cluster treatment (top, right). If optimism begins to fade, then research must go beyond the hierarchical model (bottom, right) to understand the molecular basis of disease.

Investing in basic science to probe the molecular basis of virulence may be complex and tedious. The answers may not inform us sufficiently. It may leave room for doubt but it may also create room for consilience and build extensions for imagination and innovation. Science, engineering and economics<sup>460</sup> may instruct us to “*build back better*”<sup>461</sup> the predictive compass we may need for our tryst with destiny. Whether we can reach that fateful destination or not may be shaped, in part, by the plight of our ability or inability to reduce the incredible inequity which surrounds access to health<sup>462</sup> science and healthcare.

To improve our (global one<sup>463</sup> health) preparation for future epidemics and pandemics we need metrics (quantitative comparisons) to better grasp the variations in molecular structure and function associated with physiological dysfunctions, degree of virulence with respect to infections, rate of replication of infectious agents and factors affecting mortality. Genetic perturbation screens and GWAS (genome-wide association studies<sup>464</sup>) are already generating molecules of interest for further analysis.

These are tasks for dedicated bench scientists who may toil for long hours to contribute even an infinitesimal iota of data to inform our understanding. Science in the service of society is a purveyor for the progress of civilization. Credible advances in science may not be achieved by flaky<sup>465</sup>, fluffy<sup>466</sup> and fuzzy<sup>467</sup> flash of publicity, chicanery and malarkey<sup>468</sup>.

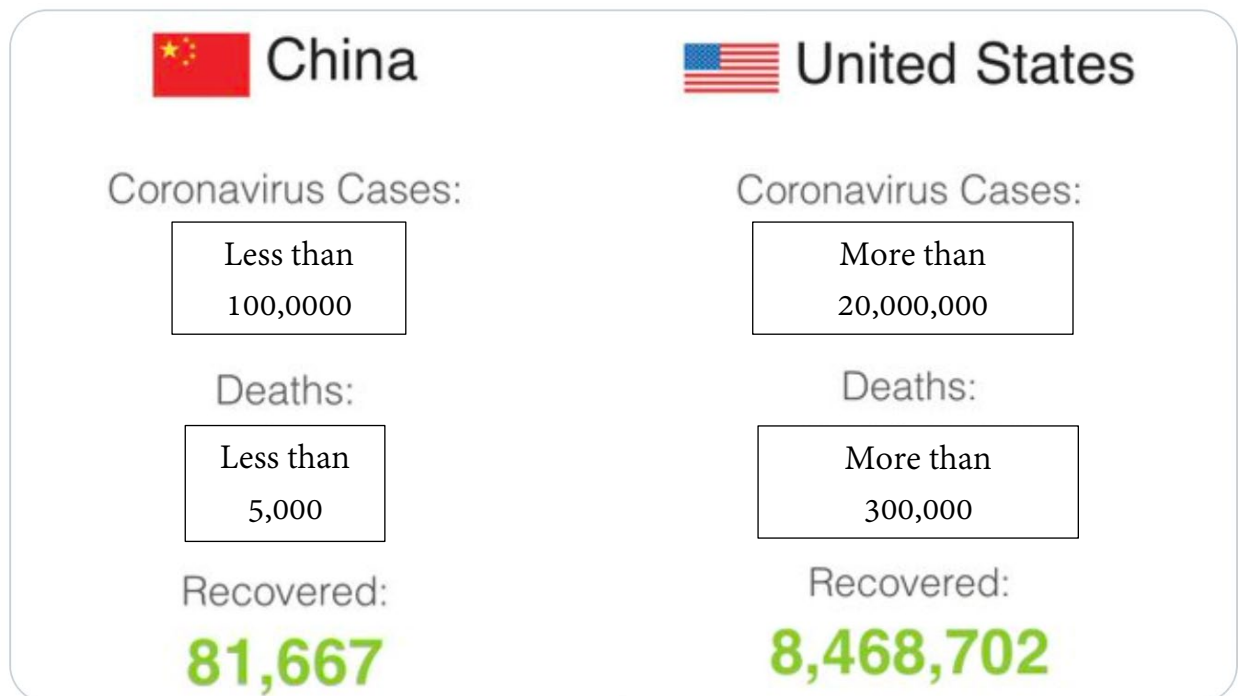


Figure 34: Let’s do the numbers<sup>469</sup>: can we explain this *incredible* difference? True, false or artifact?

## APPENDIX IV – APTAMERS as ADJUVANTS and/or PARALLEL ALTERNATIVE to ANTIBODIES

Global public health and healthcare needs immune-agnostic and temperature-agnostic low-cost therapy

Dedicated bench scientists from diverse fields in molecular biology and biochemical genetics worked for years before the successful *in vitro* transfection<sup>470</sup> of nucleic acids (DNA, RNA) into human cells in tissue culture. It was an even greater ordeal to establish that *in vivo* only those messenger RNAs (synthetic mRNAs) with a modified nucleoside<sup>471</sup> may avoid immune recognition. Free RNA in higher animals rapidly activate host immune response. SARS-CoV-2 replicates its RNA inside coated vesicles<sup>472</sup> in infected cells to evade immune surveillance. The promise of synthetic mRNA immunotherapies<sup>473</sup> and its success as a CoVID-19 vaccine displays excellence in intellectual pursuit, plight and perseverance.

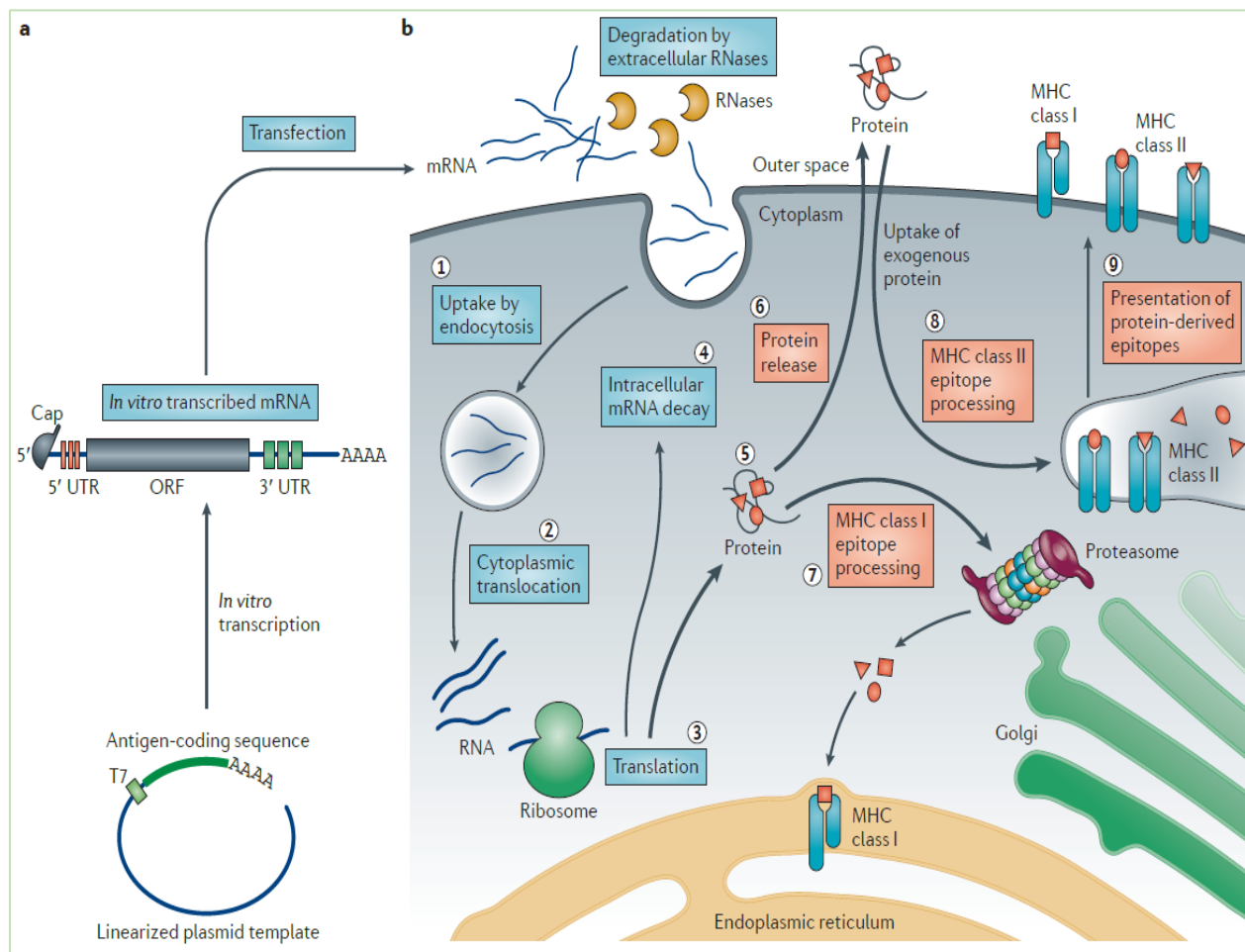


Figure 35: Synthetic messenger RNA encodes the target antigen which is packaged in a lipid nanoparticle and injected into individuals. Following uptake by cells, the mRNA<sup>474</sup> is translated in the cytosol<sup>475</sup>. The foreign protein may be released (induces B cell immune responses) and/or the antigen-derived epitopes are presented on the cell surface by major histocompatibility complex proteins (MHC) class I and II.



The title of this sub-section springs from the chasm between invention and implementation. The latter is a combination of resources and logistics. Implementation depends on cost of goods, production, facilities, distribution, personnel to facilitate access and administration. mRNA vaccine is a milestone for science but is it accessible, globally? Are aptamers affordable as a global alternative or wishful thinking?

One could have left this discussion at the doorstep of social disequilibrium if not for recent developments<sup>476</sup>. Emerging reports<sup>477</sup> suggests that the virus seems quite eager to take a bite out of the vaccine strategy. The *usual* suspect is antigenic drift<sup>478</sup>, which has hindered the creation of a successful vaccine against the influenza virus<sup>479</sup>, despite the devastation due to the 1918 global flu pandemic<sup>480</sup>.

This usual (antigenic drift) occurrence may be *unusual* in case of SARS-CoV-2 (and coronavirus family) because error correction in coronaviruses (during viral RNA replication) should, bio-*logically*, reduce the frequency of mutations which contributes to antigenic drift (see Figure 10). However, antigenic and epitope evolution<sup>481</sup> is not solely due to antigenic drift (mutations). Assuming the latter still holds true for recent SARS-CoV-2 mutants, then, once again, we may have stepped into yet another unknown abyss in our effort to understand the molecular evolution of antigenic drift in SARS-CoV-2 and implications for future vaccines<sup>482</sup>.

If this paradox (antigenic drift despite error correction) is a sign of what may eventually become a paradigm for SARS-CoV-*n* then the mRNA vaccine strategy may be forced to continually invent and reinvent itself to keep up with the antigen/epitope evolution if it chooses to serve as an effective vaccine. It is a mimicry of “flu vaccines” which are of questionable efficacy.

SARS-CoV-*n* may evolve as the aetiologic agent of future CoVID-*yyyy* and start trending in the annals of the 21<sup>st</sup> century medical anthropology, with usual public health and wellness concerns. *But*, if the mortality and morbidity from SARS-CoV-*n*/CoVID-*yyyy* approaches SARS-CoV-2/CoVID-2019, then the economics of re-inventing mRNA vaccine cycles (for antigenic drift) and vaccination, may be prohibitive *even* for the general population in most affluent nations. Pursuing multiple<sup>483</sup> targets<sup>484</sup> for vaccines is prudent. Including aptamers as an alternate may be even wiser.

Despite its existence for about thirty years, patent-free classic aptamers are still uninfluential in molecular medicine except for one FDA-approved drug. Aptamers in molecular diagnostics including proteomic profiling (patented<sup>485</sup> modified aptamers) may not have reached beyond R&D domains, yet. This discussion about classic aptamers for non-invasive rapid detection and prevention (as an “agile” drug which may be configured and re-configured/re-composed with relative ease compared to syn-mRNA vaccines) builds on *one success*, the RNA aptamer based FDA-approved drug Pegaptanib (Macugen) as a treatment for choroidal neovascularization associated with age-related macular degeneration<sup>486</sup>. However, progress<sup>487</sup> in the use of aptamers are not in short supply. But, it should be no surprise if critics choose to label the advocacy of aptamers as a case of *chacun voit midi à sa porte*.

The short-term rebuttal to immune-agnostic suggestions (including the current advocacy for aptamers) is the fact that epitope evolution due to antigenic drift in SARS-CoV-2 variants may not affect the molecular strategy adopted by the synthetic mRNA vaccine directed toward the SARS-CoV-2 Spike protein (including the receptor binding domain, RBD). Epidemiology of S protein variants supports<sup>488</sup> this short-term view. The long term risk mitigating strategy calls for alternative therapies.

Why are we observing an increasing number of variants in a virus family supposedly equipped with error correction<sup>489</sup> system? Is the replication coupled error correction<sup>490</sup> erroneous? Inaccuracies during replication by the RNA-dependent RNA polymerases (RdRp, nsp12, Fig 8) are error-corrected by nsp14, a bifunctional enzyme possessing RNA cap guanine N7-methyltransferase (MTase) and 3'-5' exoribonuclease (ExoN) activities (proofreading function). Mutated<sup>491</sup> or inactivated nsp14 may be lethal for viral replication<sup>492</sup>. Error correction is the reason for RNA genome size expansion (Figure 16). Nsp14 mediated excision of erroneous mutagenic nucleotides inserted by nsp12 is also the molecular basis<sup>493</sup> for potential nucleoside drug resistance exhibited by SARS-CoV-2 family of pathogens<sup>494</sup>.

With accumulating genome wide errors<sup>495</sup> in coding and non-coding sequences of emerging<sup>496</sup> variants, the impact of errors on SARS-CoV-2 proteins may influence the mortality and morbidity due to SARS-CoV-2 infection. Spike protein RBD is an element in this set and may remain as the prominent antigen of choice for mRNA vaccines, albeit in the short-term. The nsp14 protein works with the non-enzymatic<sup>497</sup> nsp10<sup>498</sup> in an error correction complex to restore replication fidelity. Both are susceptible to mutations. Reduced efficacy of either protein may deliver deleterious outcome for humans if the new mutations introduce changes in the progeny virus which may be harmful to humans.

Structure and function (genomic/proteomic) comparison of nsp14 and nsp10 (independently and in complex) between sister clades of SARS-CoV-2 variants (D614G<sup>499, 500</sup>, N501Y<sup>501</sup>, etc.<sup>502</sup>) may offer clues, if we can identify specific mutations in the parent genome which affected nsp14 (exoribonuclease for proofreading) and/or nsp10, resulting in progeny which causes more harm.

The generality of evolution<sup>503</sup> and its punctuated equilibrium<sup>504</sup> introduces uncertainty in the sudden/dynamic appearance of variants which manifests<sup>505</sup> differently depending on the host. The latter warrants development of a *portfolio* of therapies. Creating a portfolio of synthetic mRNA vaccines is perhaps a better and/or efficient immune-dependent pharmacological strategy compared to the classical approach of using attenuated virus in a viral vector<sup>506</sup>. But, high energy<sup>507</sup> consuming products (vaccines) are incompatible with distribution to energy-deprived regions, nations and continents.

Appendix II outlines the rigor of kinetics necessary to establish the criteria for DNA aptamers. The foundation of the science in terms of molecular biophysics is without question. The silent success of SomaLogic<sup>508</sup> may be a “radio silence” because it was not intended for publicity (for example, the media-cultivated baseless drum beat of “Smart Cities” that is about to bite the dust<sup>509</sup>).

Trials and tribulations<sup>510</sup> involved in the mRNA vaccine strategy may have a parallel story<sup>511</sup> in the innumerable<sup>512</sup> failures (200+ experiments) that preceded *one* successful protocol for creating LNP (lipid nanoparticles, see Figure 26) to deliver charged macromolecules to cells, *in vivo*. Use of aptamers as an alternate path for 80% of the global population may have to endure pain, shame and ignominy on the fatiguing climb, hopefully, to claim success, albeit partial. This “*dismissed tool*” (DNA aptamers) may lead to alternative diagnostics and therapeutics to serve the globally underserved but it will not be easy.

Unique and degradation-resistant aptamers with enhanced binding affinities may unleash new approaches. But attempts to expand our genetic vocabulary has met with limited success. Modified<sup>513</sup> nucleotides “artificial” to nature are available. Will the inclusion of modified bases in aptamers result in favourable changes in terms of kinetics and dynamics?

Name	Sequences	K <sub>d</sub> (nM)	Percentage in pool
LZH1	~CCAATAAATCT <b>P</b> GC <b>Z</b> PGGTATCGG~	14±3	0.40%
LZH2	~GGAAGTGACGGTAGC <b>P</b> TTTTGGAGG~	24±3	0.28%
LZH3	~CGACC <b>Z</b> GACTTTTAGC <b>P</b> TCGAATAG~	24±5	7.84%
LZH4	~GGATAAGTCT <b>P</b> AC <b>Z</b> PGGTATCATG~	36±5	0.50%
LZH5	~GCTAC <b>P</b> TGGGCCCTGGT <b>P</b> TCTGTGC~	41±7	0.66%
LZH6	~TATCAGCCCGATTAAACT <b>P</b> ZATGG~	47±5	2.15%
LZH7	~CAATAATTCT <b>P</b> GC <b>Z</b> GCGGTATTGGG~	55±8	23.04%
LZH8	~TATTAGTACGGCTTAACCC <b>P</b> CATGG~	96±10	0.65%
LZH9	~TATC <b>P</b> GTTGCCCTTAAAGGCTATGG~	214±26	25.67%
LZH10	~CGCCACGGAAGAGTCTCTGCGGCC~	234±53	1.35%
LZH11	~CGCCCGC <b>Z</b> GAGCAGG <b>P</b> CCCCCCCCG~	247±25	0.27%
LZH12	~CGGCTTGACAGAC <b>P</b> GCAT <b>Z</b> GATCAG~	298±37	0.14%
LZH13	~GTGCGGCCACCATACCCTCTGGGC~	326±53	0.91%
LZH14	~CCAACCTGCGACCCACAACCCTATG~	346±50	1.27%
LZH15	~TTGCGCATGCCACCACCTACCAGGC~	727±84	1.25%
LZH16	~TCCCTACATGCGAGTACCACCCTG~	>1000	0.74%
LZH17	~CCACCTAAGCTCTGGTTCCCGTGG~	>1000	0.54%

Table II: This table (Biondi and Benner, 2018) shows DNA aptamers containing artificial nucleotides in “red” letters (**P** and **Z**). Aptamers with the lowest K<sub>d</sub> (highest affinity, slow turnover) contained Z and/or P residues (LZH1 K<sub>d</sub> 14 nM) while natural AGCT-only aptamers had K<sub>d</sub> ranging from 326 nM (LZH13) to >1 μM (LZH16, LZH17). Percentage in the pool indicates relative amounts of a particular sequence to the total sequences analyzed from cycle 13. Six<sup>514</sup> nucleotide DNA alphabet containing AGCT (Adenine, Guanine, Cytosine, Thymine), **Z** {6-amino-5-nitro-3-(1'-β-D-2'-deoxyribo-furanosyl)-2(1H)-pyridine} and **P** {2-amino-8-(1'-β-D-2'-deoxyribofuranosyl)-imidazo[1,2-a]-1,3,5-triazin-4(8H)-one} was guided by Watson-Crick base pairing ability. Perhaps a larger number of “artificial” DNA and RNA alphabets may be available for use in single stranded aptamers to improve specificity and sensitivity.

## APPENDIX V – ENGINEERING RNASE RESISTANT RNA APTAMERS: A PARALLEL APPROACH?

DNA aptamers may be easier to work with compared to RNA aptamers. Which one is better? There is no single answer. Unless created and evaluated, we do not know which synthetic aptamers may be lower cost alternatives for diagnostics and therapeutics for 80% of the less affluent global population. One shoe does not fit all. We need portfolio of tools from an epidemiologic<sup>515</sup> perspective because bias<sup>516</sup> is omnipresent in science and variations in human physiology are far beyond the grasp of medicine.

Ubiquitous presence of copious amounts of mammalian (extracellular) ribonucleases (RNase A<sup>517</sup>, RNase P<sup>518</sup>) and to a lesser degree the intracellular RNase L<sup>519</sup>, if present in cell-free<sup>520</sup> extracts, induces fear in scientists. It makes it harder to work with RNA tools for diagnostics and therapeutics. On the other hand, the configurational versatility of RNA makes it an efficient “lasso” to bind proteins. We focused on ssDNA aptamers because the deoxyribonuclease families may be less of a problem<sup>521</sup>.

Thirty years ago this conundrum was visited by none other than Paul Zamecnik<sup>522</sup>. More than a decade ago the seminal paper by Katalin Karikó (Karikó *et al* 2005) was the key which later<sup>523</sup> enabled Moderna and Pfizer/BioNTech’s mRNA vaccine. Advances<sup>524</sup> in therapeutics<sup>525</sup> over the past few years provide optimism. Use of modified pyrimidines<sup>526</sup> and purines<sup>527</sup> to create synthetic RNase-resistant single stranded RNA aptamers may not be an insurmountable challenge<sup>528</sup> for nucleic acid chemists.

As outlined in Appendix I, establishing rigorous pharmacokinetics and pharmacodynamics of synthetic RNase-resistant single stranded RNA aptamers for specific protein targets (communicable diseases and non-communicable<sup>529</sup> diseases) may be similar but with at least one additional qualification. The pre-requisite is a two part pre-phase where the range of resistance must be determined (profile of RNase attack on naked RNA aptamers in cell-free extracts *in vitro*) to serve as a qualitative indicator. Verification of the stability of the RNA aptamer to remain intact (structure, function) in cell-free extracts *in vitro* compared with saliva samples (from uninfected/normal human and animal) may be critical as a metric or index of comparative stability to serve as a quantitative guide. These tasks are not easy.

## APPENDIX VI – SCIENCE, SOCIETY VS SENSITIVITY, FREQUENCY OF TESTING AND COST

Discussion with global experts (see “acknowledgements”) points to the relative naïveté (?) of this proposal with respect to aptamers as potential diagnostic (ADD) and therapeutic (AAAD) tools in detection and prevention of SARS-CoV-2. Thirty years of advances in aptamers may appear to suggest that this proposal is inadequate to approach, meet or exceed the existing excellent body of science which has been accomplished with respect to specificity and sensitivity of aptamer-protein interactions.

Sub-attomolar detection limits<sup>530</sup> are unlikely to be attained by ADD tools. Do we need such sensitivity for ADD as a global public health tool? Is detection at  $10^{-18}$  moles per liter (concentration) necessary to detect SARS-CoV-2 in (asymptomatic) individuals to reduce transmission? What is more important: frequent, low-cost, low sensitivity tests or infrequent, high-cost, highly sensitive qRT-PCR?

How many viable SARS-CoV-2 must invade a human in order for the individual to be infected? *The answer is unknown.* In ferrets and cats the infectious dose is  $10^5$  colony forming units (cfu)<sup>531</sup> which suggests that attomolar, femtomolar or nanomolar levels may be unnecessary for detection of viral RNA or proteins. Can micromolar sensitivity (*twelve orders of magnitude* lower than attomolar) suffice to detect infection by viruses, such as SARS-CoV-2? Just because high performance Koenigsegg *Agera* is the vehicle of choice for a few in Chicago or Cologne or Cannes, does not mean that it a global standard for automobiles or must be *the* adopted make and model for driving in Calcutta or Cairo or Cartagena<sup>532</sup>.

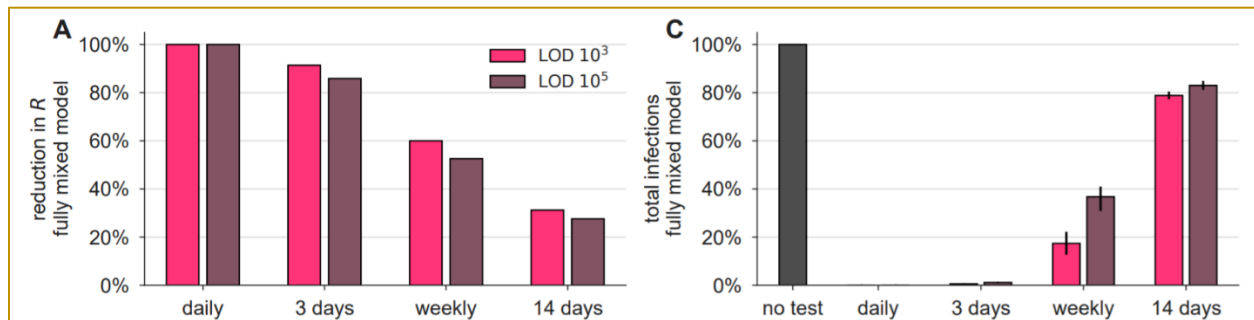
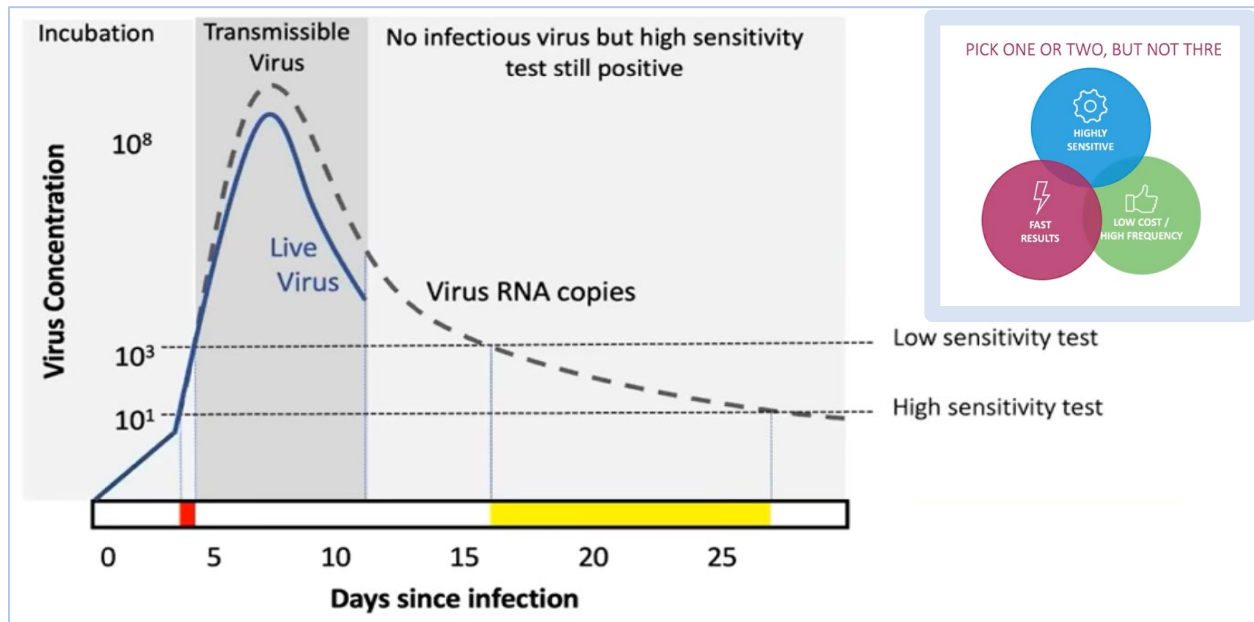


Figure 36: Sensitivity is *not* the Holy Grail. The evidence to maximize sensitivity may not be the golden rule<sup>533</sup>. Sensitivity is a *shoe where one size does not fit all*. The difference in virus concentration (limit of detection, LOD) between high and low sensitivity tests may be two orders of magnitude (**TOP** panel, note **red** zone, horizontal bar). The **time** for the viral load to leap from 10 or 100 ( $10^1$  or  $10^2$ ) to 1,000 or 10,000 ( $10^3$  or  $10^4$ ) is a matter of hours (*viral* lifecycle). What is detectable by gold standard qRT-PCR ( $10^1$  or  $10^2$ ) is **ALSO** detectable with a lower sensitivity test ( $10^3$  or  $10^4$  virus concentration), a few hours later. For control of transmission, degree of sensitivity is neither critical nor rate limiting with respect to LOD (**BOTTOM** panel). The *frequency*<sup>534</sup> of testing is key to controlling the pandemic. Low-cost, rapid, lower-sensitivity tests are essential to establish the epidemiology<sup>535</sup> of the population already<sup>536</sup> infected.



Therefore, the notion that attomolar or femtomolar is the “best” or the “standard” of sensitivity for enabling access to public health is a *misguided bias* in the Western world. For a different purpose, a different level of sensitivity may work for a community if the tool delivers **value** by reducing mortality and morbidity, perhaps not for all, but may be for 80% of the population. ADD tool will deliver value.

The criteria of good and best is subject to modification by socio-economic yardsticks when the “best” is out of reach of the people and government healthcare budgets. The value of the service must not be compromised just because it is used by people who are less litigious and less affluent. ADD is also expected to serve the underserved nations where 80% of the global population resides. This is not only about science in pursuit of the truth but also the role of science as a service to society to lift many boats, not just a few yachts.

that the University of Cambridge, which has done best at teaching mathematics, is the one from amongst whose graduates have come more of the English poets, while Oxford, which has specialized in the humanities, has tended to turn out writers who have attained, on the whole, a high level of mediocrity. I suppose by the time a man has discussed literature with a learned and witty tutor two or three times a week over a period of years he has rather talked it out instead of writing it. Then he knows too well how much good work has been done and how good it is, and is too respectful toward it: ‘Who am I that I should do better?’ ”

Quote [A]: Three decades of grand scientific accomplishments in the field of aptamers may induce a novice to hear the echo of the quote “who am I that I should do better” attributed to Alfred North Whitehead<sup>537</sup>. Similar presuppositions may have fueled attempts by Harvard (established 1636) to “absorb” MIT (established 1861) in six failed merger attempts<sup>538</sup> (1862 through 1905). Unquestionable excellence in the domain of aptamers may also serve the underserved in the post-pandemic world but it may not happen if we are trying to *gild the lily* or without the guidance and support from the experts.

Comparing sensitivity of detection between tools may lead to imprecision sensitivity (the conventional wisdom surrounding “apples versus oranges” problem). The elements that may be measured in an ADD-type detection tools shares few common features with tests using quantitative RT-PCR (reverse transcriptase – polymerase chain reaction). Perhaps comparing the *outcome* of the tests may offer a common feature as another dimension of sensitivity. If two tools were used to test the same group of individuals and the tests were scored, with respect to the ratio of positive over false positive versus negative over false negative, then this ratio (or an appropriate combination of positive, negative, false positive, false negative) may serve as an indicator of meaningful sensitivity (compared to the gold standard).

The caveat in this framework is that the notion of false negative as a function of the true viral load is a difficult metric. Nevertheless, the distribution of the *outcome data* (several tools compared to the gold standard) may reflect the range or the spectrum which is useful for diagnostic purposes. The latter is the key to determining the efficiency of each tool, for example, if the score falls within a certain range then they are useful. For ADD, if optical signal transduction from quantum nano dots (to smartphone<sup>539</sup>) is inefficient in terms of signal versus noise, then other options<sup>540</sup> are available (electrochemical impedance spectroscopy<sup>541</sup>, surface plasmon resonance<sup>542</sup>, etc.) to obtain the data.

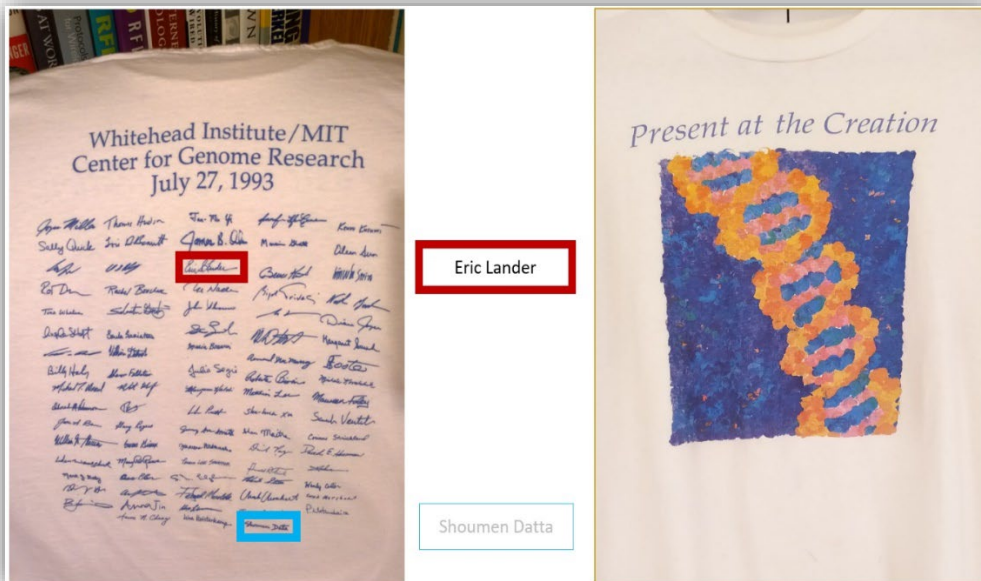


Figure 37: “Frameworks” (Eric Lander<sup>543</sup>). False positives and false negatives as a function of true viral load are mired in numerical controversy due to LOD and inherent errors in biological measurements<sup>544</sup>.

Specificity may turn out to be equally critical in the context of testing individuals who may be affected by multiple viruses<sup>545</sup> at the same time. Patients with persistent<sup>546</sup> infections may be exposed to yet another virus. Context-aware diagnostic specificity must differentiate *between* viruses in the event that the recognition element shares homology or something related. However, “re-purposing” motifs may offer advantages through molecular mimicry (see APPENDIX – IX on page 64 of this document).

Epidemiologic indicators (for example, R<sub>0</sub>) are statistical tools which are entirely dependent on correct data. The quagmire of data sharing, fake data, suppressed data and data-driven misinformation makes a mockery of such KPIs (key performance indicators). Using qRT-PCR as the ultra-sensitive gold standard tool to provide data for R naught estimation of transmission in local communities may be limited to a few cities in the world while the pandemic continues its march of unreason, globally.

CoVID-19 pandemic may morph into a globally endemic infectious disease (EID). The resulting situation may be akin to a social IED (improvised explosive device) which could transform geo-political economy. Demand for alternatives to vaccines may be served by small molecules, including aptamers, for diagnostics and therapeutics. The grave disequilibrium facing 80% of the global population calls for tools to deliver **value** rather than pursue six sigma<sup>547</sup> R&D to reach the *luminous summit* where “best, highest or lowest” resides within the *hall of fame* for metrics of excellence with respect to characteristics, attributes and features.

Contrary to what one might conclude, this proposal neither promotes nor includes that which is shoddy and second grade masquerading as good enough. Re-visiting aptamers for ADD and AAAD with “new” eyes is a structured exploration and convergence of tools and protocols to deliver value *optimized* to help *billions of users*. But, it may fail to be useful. Yet, the pursuit must continue “*not because the work is easy but because the work may be hard*”<sup>548</sup>. The plight of ADD and AAAD may be a tryst with destiny.

#### APPENDIX VII – ROOM FOR *IN SILICO* CREATIVITY IN PREDICTING APTAMER DESIGN?

Nucleic acid polymers binding to a protein or peptide with *specificity in body fluids* is at the scientific heart of the principle salient both for detection (ADD, binds in an external environment) and therapy (AAAD, *in vivo* anti-viral activity). To create a set of these highly specific aptamers, the usual approach may start with a library of aptamers (say 10<sup>19</sup> or 10<sup>20</sup>) and through sequential enrichment (principle of column chromatography, one molecule is immobilized) the aptamer pool may be reduced (10<sup>1</sup> < x < 10<sup>2</sup>) using a range of K<sub>d</sub> values as a metric (see Table II in Appendix IV).

Molecular design *in silico* may be worth exploring to predict aptamer design and consequently focus on testing a smaller pool of synthetic aptamers. The caveat in this *modus operandi* is the departure from pristine principles of chemistry (the assumptions in the design process). The criteria for binding is determined by shape and may be similar in principle to some variation of “induced fit” which is the underlying mechanism for enzyme-substrate interactions prior to catalysis<sup>549</sup>.

The shape or configuration of a single stranded DNA aptamer is guided by base-pairing between distant stretches which could generate “hairpin” structures. The *inter*-hairpin hydrogen bonding<sup>550</sup> between base pairs and/or other electrostatic interactions<sup>551</sup> may generate tertiary structures. Can a pool of ssDNA give rise to tessellated, concatenated or catenated DNA? Are the structures/hairpins stable enough to interact with specificity with the protein target and generate a signal upon binding? Can the *in silico* approach predict the structure, a measure of its stability and the probability of binding?

As mentioned in the previous paragraph, the attributes associated with these *in silico* models assumes that nucleotides are interacting in a *standard* state. The *in silico* approach largely depends who programmed the principles of physics and chemistry associated with each molecule. One assumes the interaction is occurring in some form of optimized buffer with known hydrophobic and hydrophilic profiles, specified van der Waals radii, hydration values, pH, ionic content and a mixture of salts.

The situation is exponentially complicated when we move from ssDNA aptamers in “ideal” solutions to aptamers co-existing with large/small proteins/peptides in a colloidal solution of variable pH with different factors (for example, mucus in saliva). The amino acid sidechains play a defining role in binding. Factors that perturb secondary and tertiary structure of protein/peptide will influence binding (specificity,  $K_d$ ). Protein structures are dynamic (think resonance structures of organic molecules) and the zwitterionic status of amino acid sidechains may be sensitive to presence of other charged molecules (aptamers), pH, salinity and viscosity.

Viscosity is not so subtle an influencer because the state of fluidity (fluid dynamics, kinetics) influences Brownian motion of molecules in solution and the frequency of collision between molecules. The effect of temperature on kinetics is an equally important parameter we are avoiding, at present, because the environment of external interaction (ambient temperature at the point of use) and *in vivo* applications (body temperature in humans/animals) are unlikely to fluctuate beyond a few degrees.

Taken together, predictions from the *in silico* model under “ideal” conditions may *immediately* fall apart when aptamers (which were predicted to be “good” *candidates in silico*) are evaluated for binding strength (aspirational low  $K_d$ ) and specificity in body fluids, for example, saliva and blood, which may diverge from the computational “ideal” state (fluids may also vary between individuals).

But, this is not a fact. This is a pessimistic *hypothesis* that the *in silico* prediction may not even come close to mimicking the experimental milieu when evaluating aptamer binding to target proteins in saliva, blood or other fluids (*in vivo*). Nevertheless, *in silico* modeling and prediction is a scientific design tool worth evaluating before we conclude that the “ideal” state predictions are at complete odds when testing aptamer binding in body fluids.

For SARS-CoV-2, there is a “baseline” interaction which could serve as a template for *in silico* design and prediction tool. ssDNA aptamers were identified to bind to the Spike (S) protein RBD (Song, Yanling, *et al.*, 2020) and nucleocapsid (N) protein (Chen, Z. *et al.*, 2020) with affinity below 5 nM (one  $K_d$  0.49 nM in Zhang, Liyun, *et al.*, 2020). The *in silico* approach may model the parameters of the N protein using data/attributes relevant to the aptamers identified. The model must include primary sequence of the binding region on the N protein followed by the putative secondary structure of the region (both as a peptide and as a region in the intact protein). Linking *in silico* models with protein structure data is necessary to understand how the secondary structure may change due to mutations in the primary structure. This connectivity is required to test if variants of SARS-CoV-2 may be defective in binding certain aptamers. The ADD “sensible” system will encounter individuals harboring variants other than the “model” virus on which the *in silico* approach may be trained.

The *in silico* logic tools must connect with the knowledge base with *attributes* of the “alphabets” (DNA alphabets A, T, G, C) which are *featured* in the sequence of the ssDNA. Displaying the theoretical structure of that sequence in solution (hairpins) is a required step. Then proceed to predict which part of the ssDNA aptamer secondary structure interacts with the secondary structure of SARS-CoV-2 protein. The “*glacage sur le gateau*” will be the ability of the *in silico* tool to display the tertiary structure of the specific region of the SARS-CoV-2 N-protein bound by the aptamer. Creating a numerical framework of values must accompany the predictions. The tool needs a knowledge base to import  $K_d$  values and other equilibrium constants associated with this interaction for quantitative predictive analytics.

Can this “modelling” predict negative changes in  $K_d$  (higher  $K_d$  value) if we change the ssDNA aptamer sequence to unfavorably perturb the “model” secondary structure or change one or more of the amino acids at the “active” site of the aptamer-protein tertiary structure? If this *in silico* tool can provide the anticipated response (in this “known” scenario) then it suggests that the model is using a relatively reasonable set of attributes. The data store of attributes may be enriched by connecting or feeding the *in silico* system with *feature-specific* data to populate selected features in the model. These characteristics of feature engineering are not essential because data volume related to aptamer-protein binding is limited.

However, what is much more important, for the tool to be immediately useful, is the ability of the model to predict the impact of changes in aptamers and aptamer-protein *binding* due to changes in protonation and deprotonation from fluctuations in pH. The pKa of amino acids are susceptible to pH changes and that could modify configuration which could affect binding, either positively (lower  $K_d$ ) or negatively (higher  $K_d$ ). Creating model dependencies between pKa,  $K_d$  and Ramachandran parameters (see Figure 30) may be an important part of the analytical engine in this *in silico* model/tool. Experts may point to many other factors we have omitted. But, if the issues in the above discussion are addressed with some degree of confidence, then the *in silico* model may be ready for a design challenge. The overarching question is the ability of the tool to predict the design of aptamers which could bind target proteins with high specificity.



Using the SARS-CoV-2 N protein and the ssDNA aptamers which binds to the N protein as the base model in the *in silico* tool, could we substitute the N protein with the SARS-CoV-2 Spike (S) protein RBD (receptor binding domain) and then ask the *in silico* tool to *predict*, based on its *learning*, which ssDNA *aptamer shapes* are appropriate candidates for binding to the S protein RBD (S1)? The next expectation is a list of ssDNA *sequences* that could generate the aptamer *shapes* (predicted *a priori*) with an accompanying list of **predicted  $K_d$**  for each sequence-shape combination.

Comparing the outcome of this challenge with the data from the reported (Song, Yanling, *et al.*, 2020) binding of DNA aptamer with the S protein RBD, we can estimate the fit and reliability of the *in silico* model which used the data from the N protein as the template. What if the prediction of the *in silico* tool is incompatible with the reported aptamer that binds with the S protein? Perhaps there are at least three options: [i] abandon the *in silico* approach or [ii] start over with a different template which has rigorous data or [iii] use the untrustworthy, trial and error prone back propagation algorithm<sup>552</sup>, an error correction routine licensed to practice arbitrary numerical malfeasance.

In the name of *supervised* learning the attempt is to “train” artificial neural networks (ANN) to deliver what the trainer *wants the output to deliver*. For example, if the “training” failed to recognize a hot dog, after training with millions of photographs of hot dogs (if the ANN output is a baseball bat), then the algorithm, goes back through each step to push and pull and adjust arbitrary “weights” associated with each step, in order to *drive* the ANN to output “hot dog” as the outcome of the *supervised* learning.

To apply the algorithm in this case, the gulf between the predicted versus observed S protein binding aptamer needs to be narrowed down by pushing and pulling the *in silico* machine learning training tool to deliver the reported (Song, Yanling, *et al.*, 2020) S protein binding aptamer sequence (based on the initial training of the *in silico* model using the N protein binding aptamer). Die-hard believers observe that “force fitting” in back propagation algorithm (BPA) is sort of similar to curve fitting of data to models<sup>553</sup> and mimics the induced fit mechanism of enzyme action (Koshland, 1958) representing biomimicry.

**If “made” useful**, this machine learning<sup>554</sup> embedded *in silico* analytical tool may be trained to be deployed in a *retrosynthetic*<sup>555</sup> mode<sup>556</sup>. The key variables (aptamer and protein) leads to the  $K_d$  value. Using an oversimplified analogy, Ax (aptamer) + By (protein) = Cz ( $K_d$ ) in some form or the other. If we “fix” the desired value of  $K_d$  (or a range or set of values) and pre-set the target protein, then, can we ask the tool to predict the best-fitting shapes and corresponding optimized ssDNA sequences? Can we simulate<sup>557</sup> scenarios using a drag and drop menu ( $K_d$ , protein, etc.) and enter values in the *in silico* simulation<sup>558</sup> tool in a manner<sup>559</sup> similar to variant configuration in computer-assisted design?

Automation of small molecule discovery is the bread and butter of drug design<sup>560</sup> in the pharma industry. Convergence of bio-physics, chemistry and computer science embedded as principles in logic tools are trained to predict and guide the synthesis of designer molecules. There is a healthy place and need for some forms of machine learning techniques which may be valuable. Not all routines are as ill-conceived and over-hyped as back propagation algorithm and its recent<sup>561</sup> attempt to re-invent itself.

The *learning* that repurposed the antibiotic<sup>562</sup> Halicin, is one good example. Training the MPNN (message passing neural networks<sup>563</sup>) was structured and deployed hyperparameter<sup>564</sup> optimization, without any artificial intelligence<sup>565</sup>, as explained elsewhere<sup>566</sup>. Ensembling<sup>567</sup> was applied to improve outcomes *in silico* but predictions were **biologically** tested. Even after steps to minimize errors, the authors were cognizant to note: “*It is important to emphasize that machine learning is imperfect. Therefore, the success of deep neural network model-guided antibiotic discovery rests heavily on coupling these approaches to appropriate experimental designs.*” (Stokes *et al*, page 698)

Keeping in the mind these cautionary notes, can *in silico* tools<sup>568</sup> reliably predict sequence of ssDNA aptamers which may bind with low  $K_d$ ? Of course, the predicted aptamers must be rigorously tested in saliva samples to answer the question. But, if we can **translate the prediction** to reality *in vitro* (testing in saliva using an aptamer-based sensor), then we may accomplish an *in silico* milestone. Even if it is only partially effective, this tool may still predict, guide and design the synthesis of aptamers which may make it less arduous to explore the therapeutic application of aptamer-as-a-drug (AAAD). The demand for binding specificity for AAAD is not only more stringent but is intensely complicated because the binding must *remain* functional *in vivo*. Training the *in silico* tool to include **P** and **Z** (Table II) as “alphabets” in the prediction engine may enable the tool to predict ssDNA aptamers containing artificial nucleotides, which appear to improve binding (see aptamers with lowest  $K_d$  in Table II).

Thus far we have discussed binding *as if* the specific viral proteins are **available** to bind. The latter may be true when using purified recombinant protein as test molecules. Figure 20 removes all doubt that **bio-availability** of the **exposed protein** for binding is far from what a deterministic model could predict. Therefore, a negative outcome does not necessarily indicate absence of the target protein (by extrapolation, absence of the virus). The inability of an aptamer to bind to the target protein in an actual test with body fluids (saliva sample with live virus) may be obstructed by the glycan shield which prevents the protein from being exposed or available to the aptamer for binding.

Could we alleviate this problem by using ENGases (Section 2 and Fairbanks, Antony J., 2017)? Experiment alone can shed light whether ENGases can punctuate glycan shields sufficiently to enhance the ability of the aptamers to bind and elicit a signal (over noise) which can be transduced for acquisition of data, as proof of binding. Although the focus is on the SARS-CoV-2 Spike<sup>569</sup> protein it behooves us to explore other external proteins of SARS-CoV-2 as well as the N protein (nucleocapsid<sup>570</sup> [N], membrane<sup>571</sup> [M] and envelope<sup>572</sup> [E] proteins) as targets for detection.

## APPENDIX VIII – FIELD TESTING THE APTAMER SENSOR, ALONE, MAY NOT BE PRUDENT

The *rubber meets the road* in the outcome, that is, the data, which must indicate with **confidence** if an infectious agent (for example, SARS-CoV-2) is present or absent. For return on investment (ROI), this data is the Holy Grail. Ambiguity, uncertainty and false negatives will obliterate the value of this data and may annihilate the ROI. False positives may hurt the reproducibility and reliability of ADD. Selling for-profit tests lacking the foundation of robust kinetic data and scientific field testing without proper controls are criminal practices. University of Oxford considered forty devices but only nine met some of the performance criteria, and only one was advanced enough for assessment<sup>573</sup>.

	Positive Control	Negative Control	Aptamer Test	Confidence in Outcome (Data)
A	+	-	+	Presence of Virus: Confirmed.
B	+	-	-	Less Ambiguous. Repeat testing.
C	-	-	+	Ambiguous. Repeat testing.
D	-	+	+	More Ambiguous. Repeat testing.
E	+	+	+	Likely Presence of Virus. Repeat.
F	-	-	-	Failed to Detect Virus (< LOD)

Table III: Design of field trial may start with independent scoring of each category before the combined scoring. Positive control is tested with 100 samples which are qRT-PCR positive for SARS-CoV-2. Let us assume that the score from the positive control is 90% (compared to qRT-PCR which has a lower LOD). The **same 100 samples** are tested with the negative control and scores 99% (1/100 samples “detected” what the negative control is supposed to detect, but generally the target is never found in human saliva). The **same 100 samples** are tested with aptamer test with a score of 75%. Therefore, is the “ambiguous” in row B [table] **less ambiguous** than the “ambiguous” in row C, if we take into account the positive control is correct 90% of the time versus the aptamer test which is correct 75% when compared to qRT-PCR? Are we comparing apples vs oranges? Can this test serve as a low-cost tool to control transmission? In the real world, tests<sup>574</sup> must be performed daily to isolate individuals who test positive, early enough, at least to reduce transmission in the absence of treatment (TETRIS – TEst, TReat, ISolate). Criticism of the aptamer test may ask whether the aptamer is functional. Positive aptamer function test (AFT) may include an aptamer which binds to a known component of saliva (for example, salivary amylase). As a control for AFT, inclusion of an aptamer for the vasoactive peptide hormone BNP (Sudoh *et al* 1988, Sudoh *et al* 1990) and/or the human ACE2-short isoform (Blume *et al* 2021) may be of clinical interest. Inclusion of the former (BNP) may uncover physiological co-morbidities independent of SARS-CoV-2 and/or it may be significant in view of cardiovascular (CVD) symptoms<sup>575</sup> due to CoVID-19.

Proper implementation of field trials are crucial. Aptamer tests (ADD) must include positive and negative controls. A positive control must use a completely different medium to test the same sample for the presence of the infectious agent. A negative control should test the sample using the *same medium as the positive control* which serves as a control to confirm that the “medium” is not faulty and recognition is functional. The negative control tests for an agent that *may not be found* in saliva. Future testing may be able to create a **3-in-1** single test system with *different modes of signal transduction*<sup>576</sup> generating different streams of data from optical signal, electrochemical impedance spectroscopy and surface plasmon resonance. The data can be analyzed and visualized on a smartphone.

For resource unlimited scenarios, testing kits may include another set of positive and negative controls. The positive control (P2) medium remains identical to P1 but tests for an agent *not uncommon* in the oral microbiome<sup>577</sup>. The second negative control (N2) medium is identical (as in P1 and N1) but tests for a rare but not impossible agent, for example, HIV1. In a completely different approach, we can use immobilized hACE2 on the sensor surface as the medium (SARS-CoV-2 positive) and use hACE2-short isoform<sup>578</sup> as a control because hACE2-short isoform lacks the virus binding domains.

## THE CASE FOR LECTIN SENSORS AS CONTROLS

To change the *medium of sensing* we turn from nucleic acids (DNA aptamers) binding to target proteins (for example, Spike protein in SARS-CoV-2) to proteins (lectins) binding to target sugar groups (glycans) which may be on the same target protein (for example, glycan shield on the Spike protein of SARS-CoV-2).

Lectins are carbohydrate-binding proteins that are specific for sugar groups on other molecules. Host-derived sugar moieties on viral proteins are tools for immune evasion<sup>579</sup> whereas lectins (opsonin) trigger immune recognition<sup>580</sup>. The glycan shield on SARS-CoV-2 Spike protein (Casalino *et al*, 2020) is an excellent target for lectins, for example, mannose-binding lectin (MBL), which was reported a decade ago, to neutralize SARS-CoV mediated viral infection<sup>581</sup>. Macrophage galactose binding lectin (MGL) belonging to the Calcium-dependent C-type lectins<sup>582</sup> and subsets of I-type lectins (sialic acid-binding immunoglobulin-type lectins), Siglec-9 and Siglec-10, specifically binds<sup>583</sup> to SARS-CoV-2 Spike protein.

The role of lectins and the science behind mammalian and non-mammalian lectins are well<sup>584</sup> studied. Lectin-based adjuvants to vaccines<sup>585</sup>, lectin antibodies<sup>586</sup> (lectibodies<sup>587</sup>) and lectin-based<sup>588</sup> biosensors<sup>589</sup> are well documented<sup>590</sup>. Creating SARS-CoV-2 specific lectin sensors may not be trivial but certainly not an insurmountable barrier. Use of a lectin sensor as the positive control (Table III) needs no further emphasis. The negative control may be a mannose-binding lectin (MBL) sensor or something even more specific for detecting *Mycobacterium leprae*<sup>591</sup> which prefers to grow in cooler extremities of the human body (around ~30°C<sup>592</sup>) and unlikely to be present in human saliva samples.

Lectin	Source	Sugar affinity	Anti-SARS-CoV-2 action	Anti-SARS-CoV action	Anti-MERS-CoV action
FRIL	<i>Lablab purpureus</i>	Man/Glu	In vitro	—	—
APA	<i>Allium porrum</i>	Man	—	In vitro	—
Morniga M II	<i>Morus nigra</i>	Man	—	In vitro	—
EHA	<i>Epipactis helleborine</i>	Man	—	In vitro	—
UDA	<i>Urtica dioica</i>	GlcNAc	—	In vivo and in vitro	—
NICTABA	<i>Nicotiana tabacum</i>	GlcNAc	—	In vitro	—
Con A	<i>Canavalia ensiformis</i>	Man/Glu	—	—	—
HHA	<i>Hippeastrum hybrid</i>	Man	—	In vitro	—
GNA (or GNL)	<i>Galanthus nivalis</i>	Man	—	In vitro	—
Cyanovirin-N	<i>Nostoc ellipsosporum</i>	Man	—	—	—
Griffithsin	<i>Griffithsia sp.</i>	Man	—	In vivo and in vitro	In vitro

Lectin	Source	Sugar affinity	Type of application
Con A	<i>Canavalia ensiformis</i>	Man/Glu	Biosensor for viral detection
			Biosensor for detection of serum glycoproteins
Cramoll	<i>Cratylia mollis</i>	Man/Glu	Biosensor for detection of serum glycoproteins
BmoLL	<i>Bauhinia monandra</i>	Gal	Biosensor for detection of serum glycoproteins
GNA (or GNL)	<i>Galanthus nivalis</i>	Man	LAP

Table IV: The choice of lectin sensors as controls to improve confidence in data from aptamer tests. Non-mammalian lectins with anti-viral activities<sup>593</sup> (top). Use in biosensors (bottom). Man, mannose; GlcNAc, N-acetylglucosamine; Glu, glucose; Gal, Galactose; LAP, lectin affinity plasmapheresis.



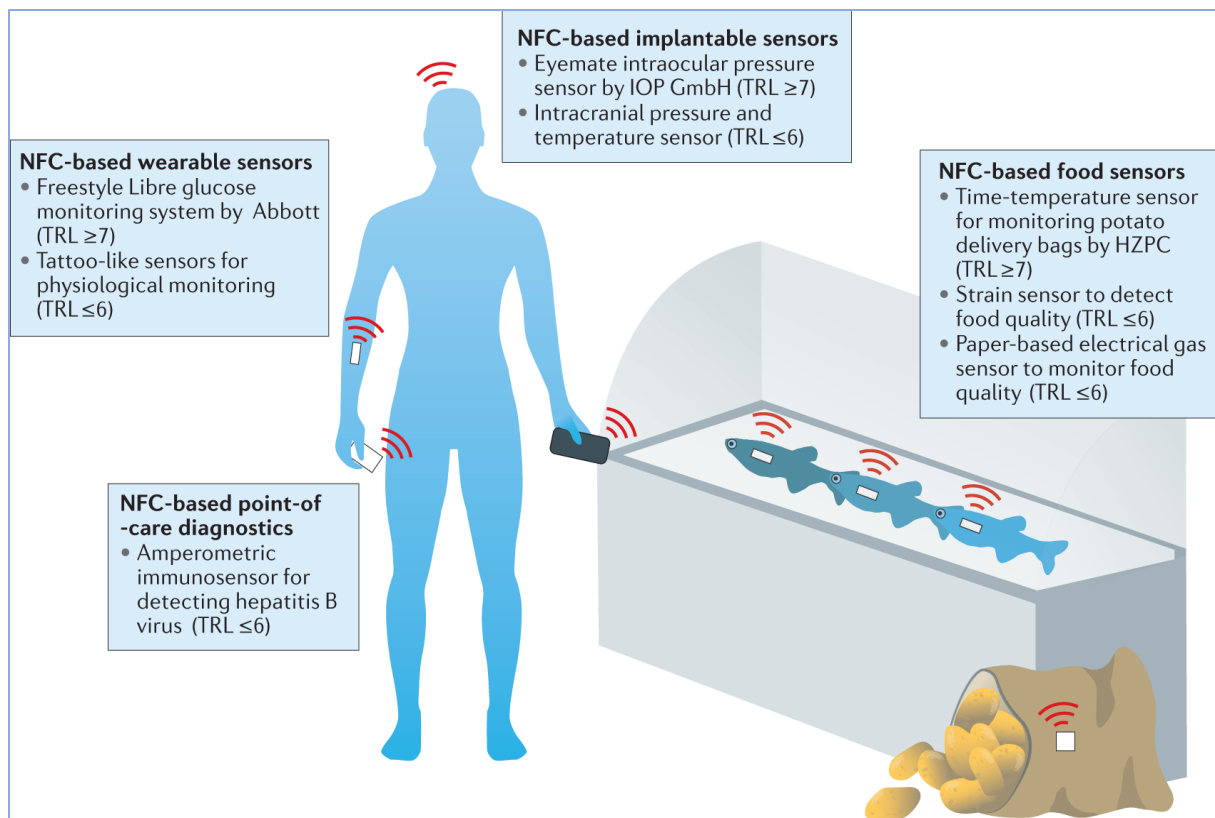


Figure 38: The confluence of ideas in this discussion consists of 2 major threads, sensors and detection. The application of both in the context of the current pandemic is viewed as a tool to stem transmission by early detection (and in future, explore the potential of aptamers as small molecule drugs as parallels and alternates or adjuvants to vaccines and antibodies). The pandemic has made detection tools highly context-aware because the global population is immersed in this quagmire. What happens if and when complacency creeps in along with the potentially forthcoming endemic? What happens if individuals are weaned off the heightened context-awareness and detection (tools, process, data, reporting) transforms into a chore to be procrastinated rather than a necessity for personal and public health? The cartoon in this figure is a sense of the future that we ought to avidly pursue in order to build a portfolio of detection modes as a part of our lifestyle (APPENDIX I, page 39, 1st paragraph). Near-field communications<sup>594</sup> (NFC) is one tool which may successfully contribute to build connectivity and functionalize the digital by design metaphor of IoT (internet of things). What if a NFC-card sensor (see Figure 21, on page 29) in a smartphone (when close to the individual, ~5 cm) could “sense” an infectious agent or a dysfunctional physiological molecule or volatile fatty acids (VFA) from a fish that once was fresh? In addition to radio frequency (RFID) readers (interrogators) or ultra-wide band (UWB) beacons or SDR (software-defined radio) in shops, malls, homes, factories and airports what if low cost NFC sensors were placed to “sense” (continuously) in a crowd-sourcing mode to detect and transmit data (inform) about analytes in their micro-environments? If underserved communities can bear the cost of NFC tools, then the explosion of detection may be exponentially beneficial. However, technology readiness level (TRL) must approach or exceed TRL9 for the NFC tool/device to be deployable. Is TRL6 or TRL7 too far away from cloud nine?

## APPENDIX IX – IS INNATE IMMUNITY TOO FAR FETCHED EVEN TO BE A RATIONAL IDEA ?

It is well nigh impossible to ignore the significant difference in data from India and USA with respect to new cases and new deaths. It is especially remarkable when compared with the population of USA (~300 million) which is one-fifth of the population of India (~1.5 billion). It is also impossible not to speculate if India's lower mortality and morbidity rate may be due to some form of innate immunity.

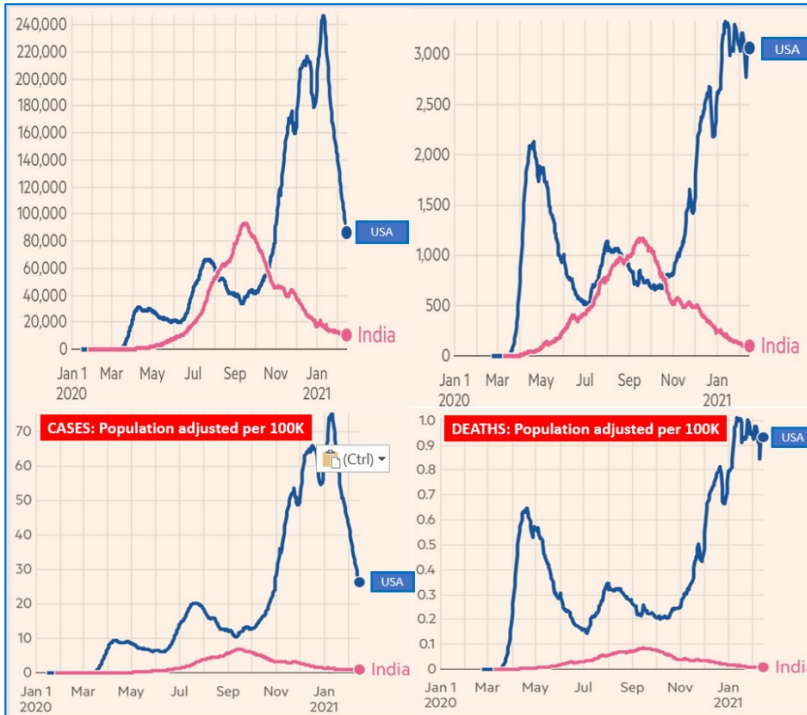


Figure 39: As of Feb 16, 2021, seven-day rolling average of confirmed new cases (left, top) and new deaths (right, top) due to CoVID-19. If adjusted for population, the difference in morbidity (left, bottom) and mortality (right, bottom) is even more striking. Is the “hygiene hypothesis” real or is the data inadequate due to inefficient or lack of testing? FT<sup>595</sup> cartoon based on data from CoVID-19 dashboard of the Johns Hopkins University, WHO, UK and Sweden.

Harrowing but it is still a fact of life in India that children are used to pick up garbage at a great risk<sup>596</sup> to their health. Has this risk turned into a reward? Waste is often found to be contaminated with fecal material and leads to infection of the gastro-intestinal and respiratory system. Those who survive in India, may have developed immunity from exposure to vast number of pathogens. Open defecation and contamination of soil and water with human and animal feces is rampant in India. Clean and hygienic Western nations with managed sanitation services may have failed to challenge to our immune system.

These squalid facts may partially explain the low numbers of CoVID-19 in India. At least one piece of scientific data suggests that this idea may not be entirely anecdotal if one considers that “Staph” infections are quite common. Molecular mimicry is blamed for diseases such as lupus erythematosus or Hashimoto’s thyroiditis or discomfort from rheumatoid arthritis, where prior infections may provoke auto-immunity. But, mimicry due to *re-purposed* motifs may have a silver lining, at least for individuals pre-infected with *Staphylococcus* sp. The superantigen-like motif of Staphylococcal enterotoxin B (SEB) generates anti-SEB monoclonal antibody, 6D3, which binds to SARS-CoV-2 Spike protein containing the same neurotoxin<sup>597</sup> motif S<sub>680</sub>PRRAR<sub>685</sub> and inhibits infection by blocking the access of the host cell proteases, TMPRSS2 or furin, to the cleavage site<sup>598</sup>. Is it possible that the SEB antibody may serve as an alternative low-cost global vaccine against SARS-CoV-2? Is this why India has fewer CoVID-19 cases?

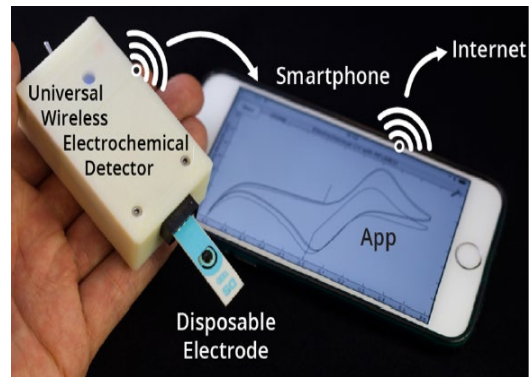
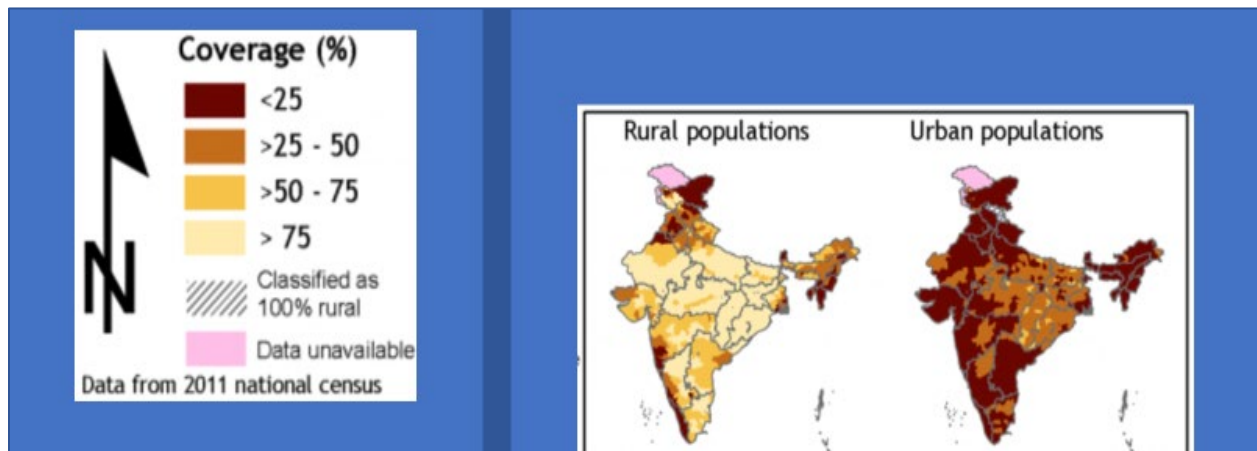


Figure 40: Open defecation<sup>599</sup> (top) & sanitation insecurity<sup>600</sup> is inescapable in rural (middle section<sup>601</sup>) India. Children eating<sup>602</sup> from rubbish dumps<sup>603</sup> are common (bottom<sup>604</sup>) in the Indian-subcontinent<sup>605</sup>. When this discussion, typical of a smartphone<sup>606</sup> linked system, using aptamers (ADD) for detection was presented to a VC (in the USA), her response was that there wasn't a good enough business case (she didn't think the focus of ADD on low-cost sensors for the underserved masses was good for her profit).



## APPENDIX X – REFOCUS ON DETECTION: LOW-COST SENSORS FOR THE UNDERSERVED

Refocusing on the central thrust of detection needs no emphasis in view of the SARS-CoV-2 mutants re-igniting the pandemic, a year later after the initial declaration of pandemic<sup>607</sup> by WHO.

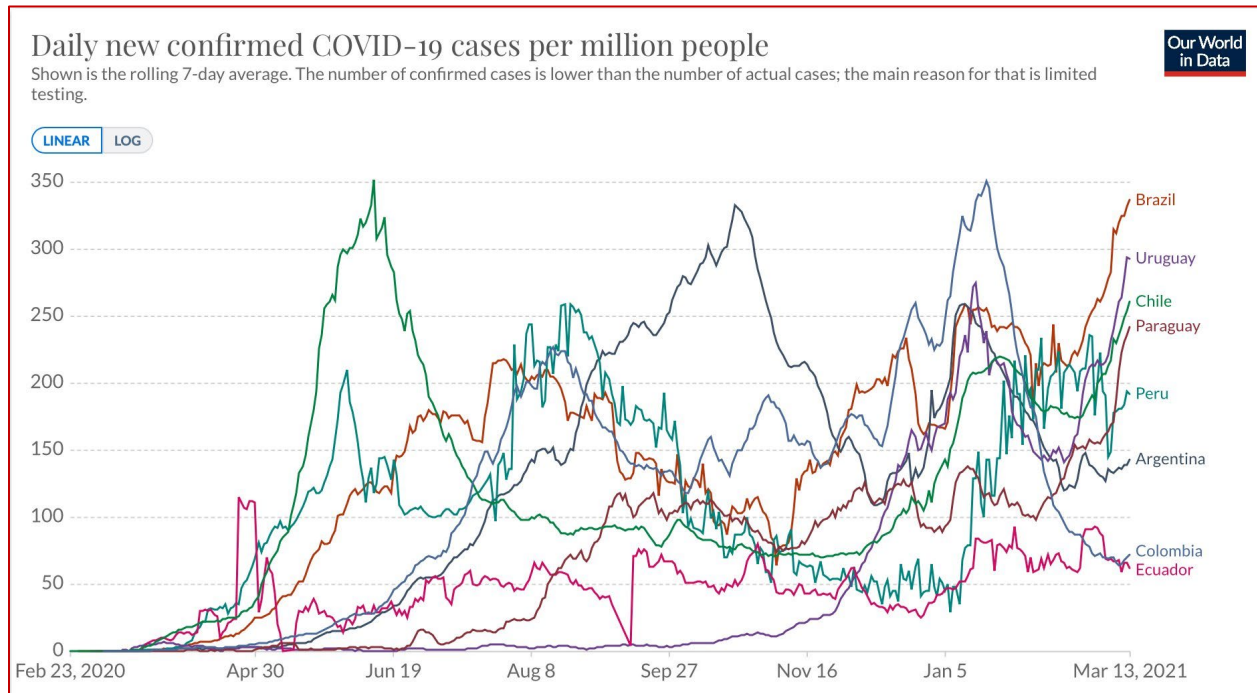
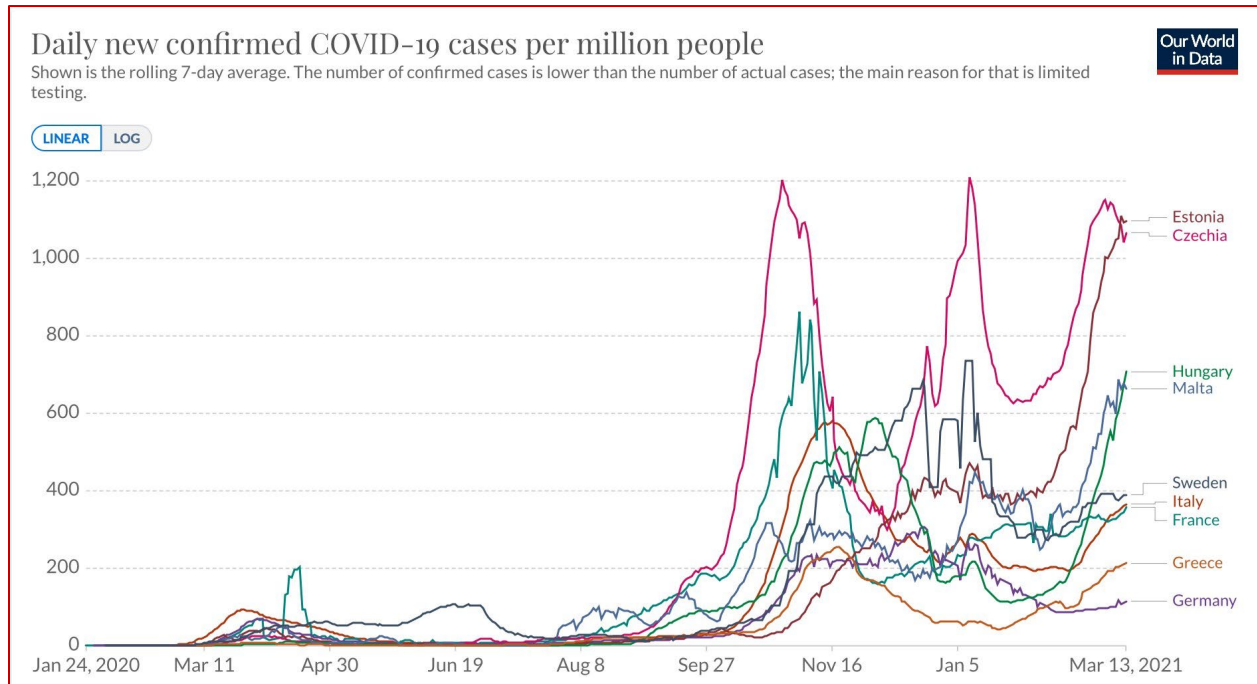


Figure 41: One year into the pandemic, the resurgence of new waves<sup>608</sup> of CoVID-19 is grim news for many geographies except a temporary but uneasy<sup>609</sup> exception in the USA.





## MECHANISM: ASSAY TO DETECT AN ENZYMATIC PRODUCT – PHENYLALANINE

Conversion of Angiotensin II to Angiotensin releases a free amino acid, Phenylalanine (F). The conversion can only happen if ACE2 is available to catalyze the reaction (enzymatic cleavage). Section 6 illustrated tethered ACE2 on the surface of low-cost laser inscribed graphene electrodes (turbostrat sensors). The idea proposed in section 6 was the *direct detection* of SARS-CoV-2 *virus* if the viral Spike (S) protein (from the saliva sample) binds to the immobilized ACE2 on the low-cost sensor (ACLITS).

The idea proposed in the current discussion is to include Angiotensin II (free floating mix) with ACLITS and use a “third-analyte” colorimetric *assay for Phenylalanine*. In an ideal world, the *absence* of virus in the sample will trigger cleavage by ACE2 to *generate Phenylalanine* residue. The color change is visible to the naked eye. Smartphone camera may detect intensity and quantify the color in an app.

If the sample contains the virus, the binding between the viral Spike protein and ACE2 (ACLITS) prevents the enzymatic action (assumption). Thus, the conversion of Angiotensin II to Angiotensin cannot proceed. In the absence of Phenylalanine, the assay to detect the amino acid generates a negative result (no color change) which indicates *presence of virus* in the sample (if color change -ve, then presence of virus +ve).

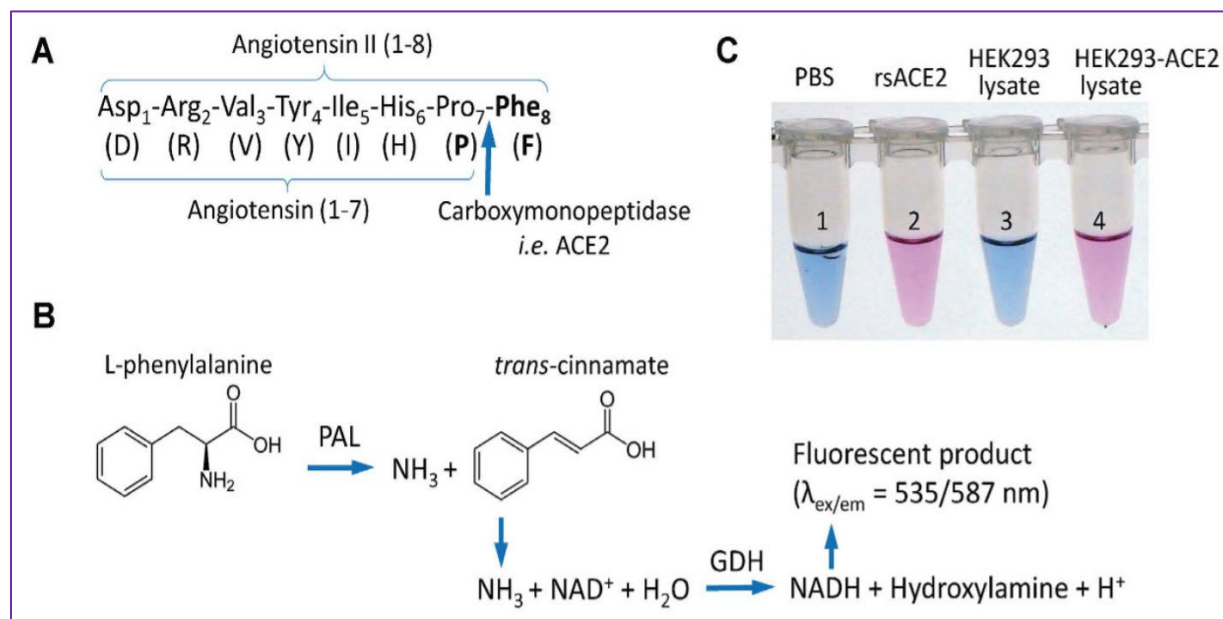


Figure 43: Phenylalanine (Phe) Assay. ACE2 cleaves (arrow) to release Phe (A). Assay<sup>614</sup> measures rate of Ang II cleavage. (B) Exposed Phe amino group undergoes deamidation by yeast phenylalanine ammonia lyase (PAL, EC 4.3.1.24<sup>615</sup>), also known as phenylalanine deaminase. In a series of reactions, fluorogenic substrate (trans-cinnamate<sup>616</sup>) is converted. GDH, glutamate dehydrogenase<sup>617</sup>. (C) Quantifiable visible color change to pink in the presence of ACE2 illustrated in tube 2 and tube 4 (emission at 587 nm).

The theoretical lucidity of the *in vitro* Phenylalanine assay is reason for cautious optimism if the cascade can be optimized to perform in the context of SARS-CoV-2 virus detection with immobilized ACE2. Reproducible success in detecting viruses on-site with saliva samples may need optimization in terms of immobilization strategies (ACLITS, magnetic nanobeads, quantum dots). Perhaps *free* ACE2 in the detection mix may perform better if steric hindrance (if any) is reduced by not tethering ACE2 at all.

Surface-free mobile ACE2 in a saliva sample *without virus will* generate enzymatic activity dependent color change when the substrate Angiotensin II is added if all other reagents are in the mix.

Surface-free mobile ACE2 in a saliva sample *with virus will NOT* generate enzymatic activity dependent color change when the substrate Angiotensin II is added if all other reagents are in the mix.

The most important source of contamination (background noise in data) is the release of Phe from other peptides [with C terminal Proline-Phenylalanine] in human or animal saliva. Many proteins, specifically salivary amylases<sup>618</sup>, contain Proline-Phenylalanine moieties (human alpha-amylase 1B<sup>619</sup>) but they are not known to release the Phenylalanine (required for the assay, Figure 43) which is not present in the terminal position in these instances (in these proteins, the human salivary amylases).

Use of [a] mutant ACE2, [b] another similar sized protein [c] mutated Angiotensin II peptide [d] another short peptide (BNP<sup>620</sup>) without terminal Phenylalanine are a few of the **essential controls** to validate specificity of interaction, in each set of experiments. Rigorous baseline data from virus-free saliva must be tested to quantify background noise (due to Bradykinin, etc). Statistically significant comparative values will be necessary to quantify noise levels stratified by demographics (age, gender, ethnicity, cardiovascular co-morbidities, drug use for hypertension, kidney function, dialysis, etc).

Peptide	Sequence
Angiotensin II-(1-8)	D-R-V-Y-I-H- <b>P-F</b>
Apelin-13	Q-R-P-R-L-S-H-K-G-P-M- <b>P-F</b>
Apelin-17	K-F-R-R-Q-R-P-R-L-S-H-K-G-P-M- <b>P-F</b>
Apelin-36	. . . . Q-R-P-R-L-S-H-K-G-P-M- <b>P-F</b>
Bradykinin (1-8)	R-P-P-G-F-S- <b>P-F</b>

Figure 44: Bradykinin is a vasoactive peptide found in saliva<sup>621</sup> and may contribute to background noise. Apelin<sup>622</sup> was not reported in saliva. Label-free colorimetric analysis using untethered ACE2 may be another low-cost solution if the reagents are stable, easily available and do not require high energy consuming low temperature storage. Quantifying color change using a smartphone camera and an app embedded with analytical tools may be a low-cost milestone for citizen science and surveillance, globally.

**INCONCLUSIVE CONCLUSION: The scientific success of mRNA vaccine may not be enough (for all)**

Concluding this article/essay is impossible in the midst of a raging pandemic despite the awe-inspiring success due to mRNA<sup>623</sup> vaccines. Having said that, one may add that at least 2 things are clear (and applicable to the 80% of the world population excluding US, EU and other affluent nations):

1. Ubiquitous deployment of low-cost, reliable, at-home detection tools are quintessential for prevention and as a preparation to mitigate future<sup>624</sup> (forthcoming?) pandemics. But, quality control and reliability of sensors made with low-cost materials may not be optimum for accuracy of low-cost sensors.

2. Alternate vaccines or alternate *to* vaccines (for example, small molecules, aptamers) must be produced by the poor nations for the poor nations (80% of the world population) because the affluent nations may not share vaccines or vaccine manufacturing protocols. They may not help poor nations manufacture vaccines despite grand gestures (and immense profits for Pfizer, Moderna, J&J, etc.).

The US CDC claimed that immunization resulted in elimination of polio in the US<sup>625</sup> by 1979 but the world is still waiting even after 40 years and still dealing with polio<sup>626</sup> outbreaks in certain poor countries, perhaps due to less successful vaccination programs. A related and similar story of inequality is evident in exploring the US<sup>627</sup> versus world history of pox viruses (VARV<sup>628</sup> and VZV<sup>629</sup>). But the prize goes to the vaccine for malaria which is finally approved<sup>630</sup> (WHO 2021) after more than 100 years since the etiology of malaria<sup>631</sup> was established in the 19<sup>th</sup> century (1897) by Ronald Ross<sup>632</sup>, working in India.

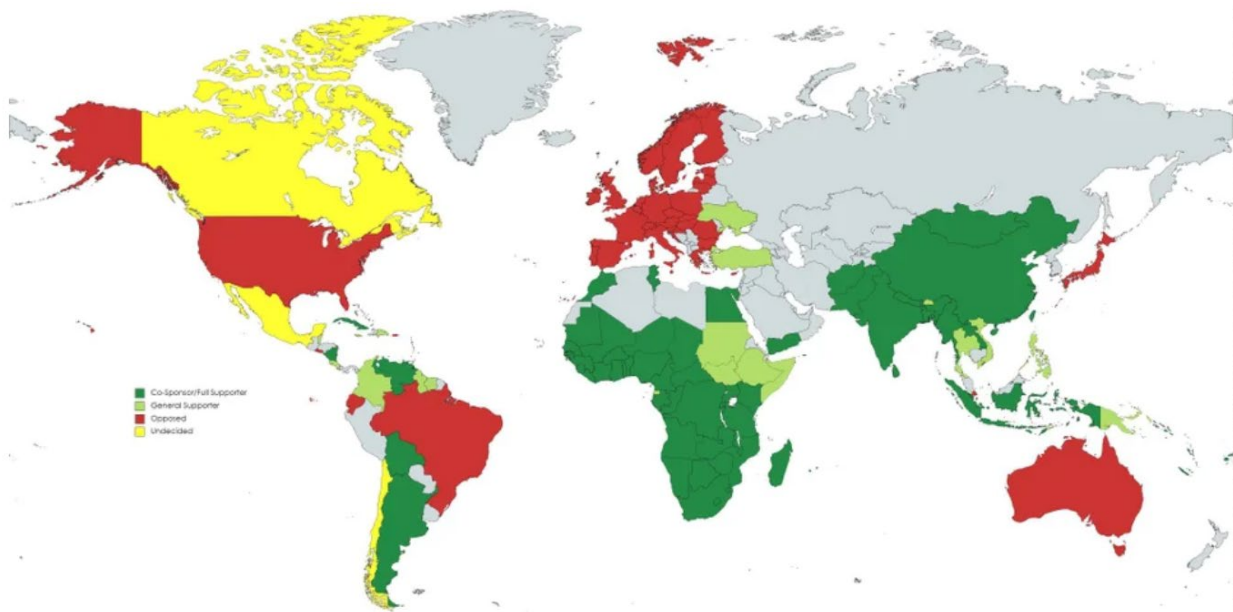


Figure 45: Countries shown in RED are the nations which OPPOSED<sup>633</sup> and *voted against* the UN call for waiving patent law for life-saving CoVID-19 vaccines. Countries in RED blocked the proposal which called for the right to manufacture and import affordable CoVID-19 vaccines. The proposal<sup>634</sup> was led by India and South Africa. Countries shown in yellow are “undecided” after ~250 million CoVID-19 cases and ~5 million deaths (perhaps a lot more?) due to CoVID-19, globally.

It could take a good part of a century for vaccines and other developments in affluent nations even to reach the poor nations. It takes much longer to make a dent in their disease burden even before these nations can begin to think about economic benefits. SARS-CoV-2 pandemic may not be a very different story and the pandemonium from future potential pandemics may be even worse than chaos.

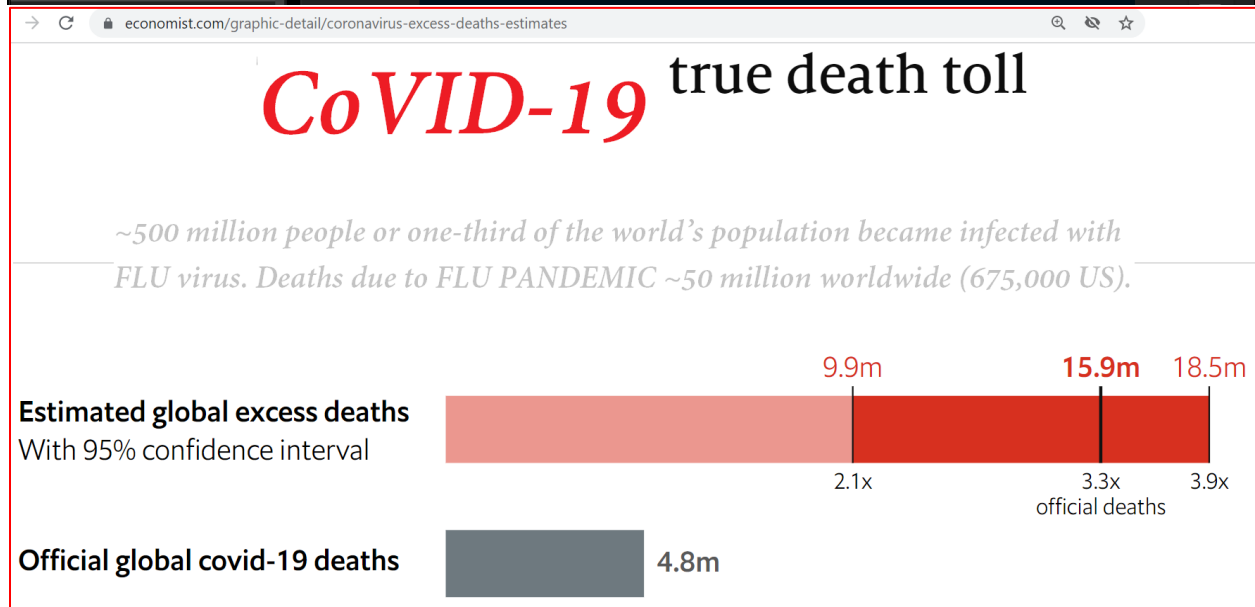
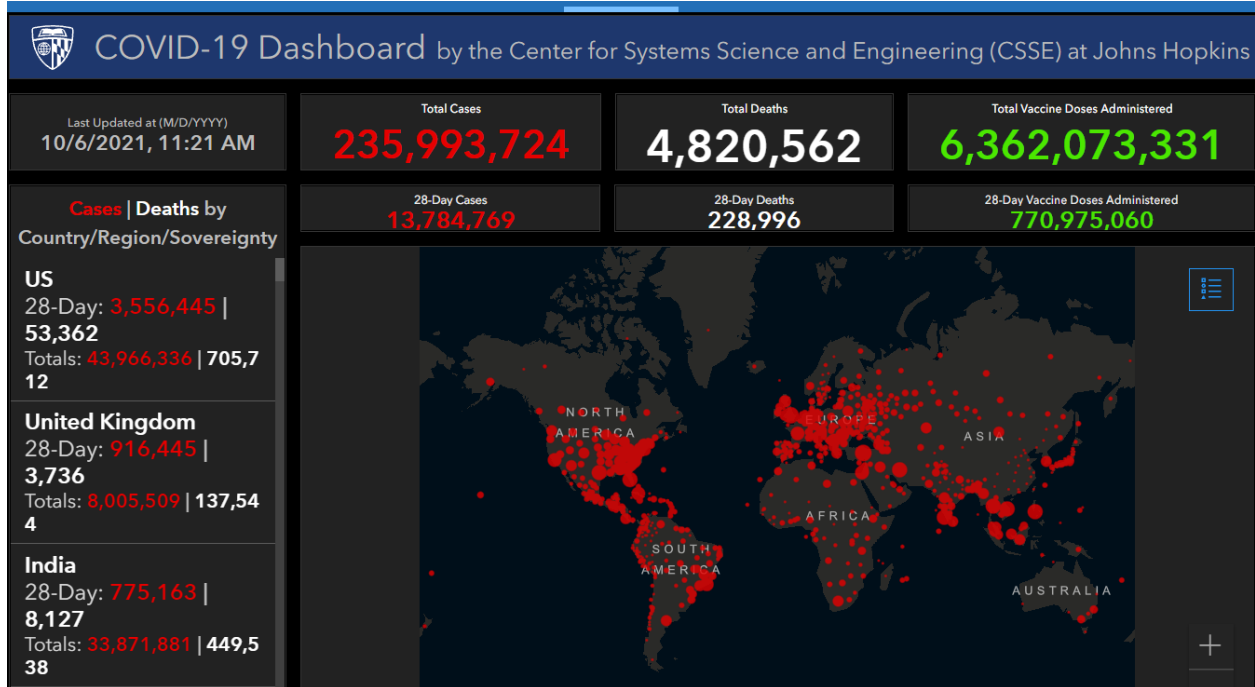


Figure 46: It is unlikely that parts of the world<sup>635</sup> are spared from the pandemic even though nations and governments may not reveal actual numbers of infected cases or may not have resources to test. Thus, it is not quite implausible to guess that the true death<sup>636</sup> toll from CoVID-19 may soon exceed 20 million.

Reiterating what was discussed elsewhere (including Appendix II and Appendix IV) we circle back to the need to pursue alternatives to vaccines which may be affordable for 80% of the world. The correct answer, still, is vaccination if manufacturers are compassionate and cares for the less fortunate.

In an alternate approach, the discovery of small molecules (see Fig 49) may extend to aptamers (somamers), nanobodies, ligand decoys and other tools to target and disrupt or lock the configuration change necessary for viral fusion proteins to initiate infection. These fusion proteins are dynamic<sup>637</sup> macromolecules and disrupting their structure may interfere with function. Mutations<sup>638</sup> affecting conformational<sup>639</sup> dynamics influences viral transmissibility.

Part of the rationale for this *modus operandi* is based on the observation that an antibody<sup>640</sup> (D25) bound to a quaternary epitope at the apex of the respiratory syncytial virus fusion F glycoprotein (RSV F) trimer could “lock” the RSV F glycoprotein in its pre-fusion state (preventing the fusion of RSV with host cell and disabling the cycle of infection). Immunization with post-fusion<sup>641</sup> RSV F<sup>642</sup> did not prevent RSV-associated acute respiratory illness<sup>643</sup> (ARI) in older adults. The pivotal importance of this lesson from the RSV helped to create the pre-fusion stabilized SARS-CoV-2 Spike protein (2P, using two Prolines<sup>644</sup>). The legendary<sup>645</sup> insertion of two Prolines<sup>646</sup> was crucial for the success of mRNA vaccines (directed against SARS-CoV-2 Spike protein). Small molecule strategy did work for RSV (Figure 49).

What if a small molecule (aptamer, nanobody, etc.) was created as a complementary “wedge” or “door stop” to bind the pre-fusion configuration and prevent the transition of SARS-CoV-2 Spike to the post-fusion configuration in vivo? Using clues from the structure of the antibody binding site (RSV) or the bioengineered “HexPro” Spike protein (SARS-CoV-2) could we manufacture a molecule to fit and block the trigger site that induces the transition of S from pre- to post-fusion conformation? Flooded (?) with this small molecule (delivered as a nasal spray, sublingual pill, throat spray or epidermal patch) the designer molecule may not prevent the virus from **binding** to the host cell receptor (eg ACE2) but if the molecule can “jam” the conformational transition then the virus will be incapacitated to inject (fuse) the genomic material. Rendered impotent to infect, the virus may cease to multiply, prevent symptomatic infection, transmission and reduce antigenic drift (show-stoppers are cytotoxicity and bioavailability).

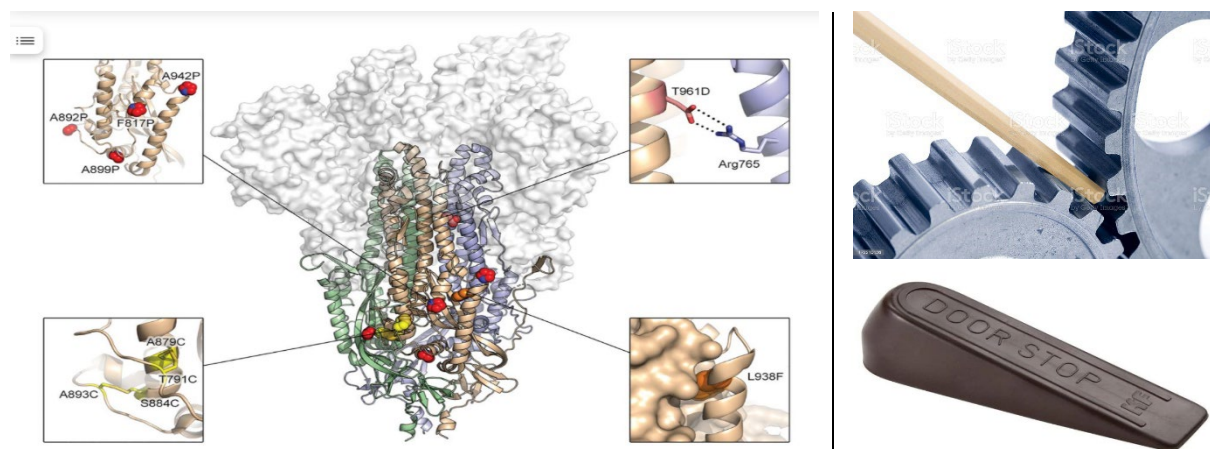


Figure 47: Clues from HexPro<sup>647</sup> stabilized SARS-CoV-2 S protein may be a guide to create molecules to “jam” the transition from pre- to post-fusion. Wedged gear and “door stop” are molecular metaphors.



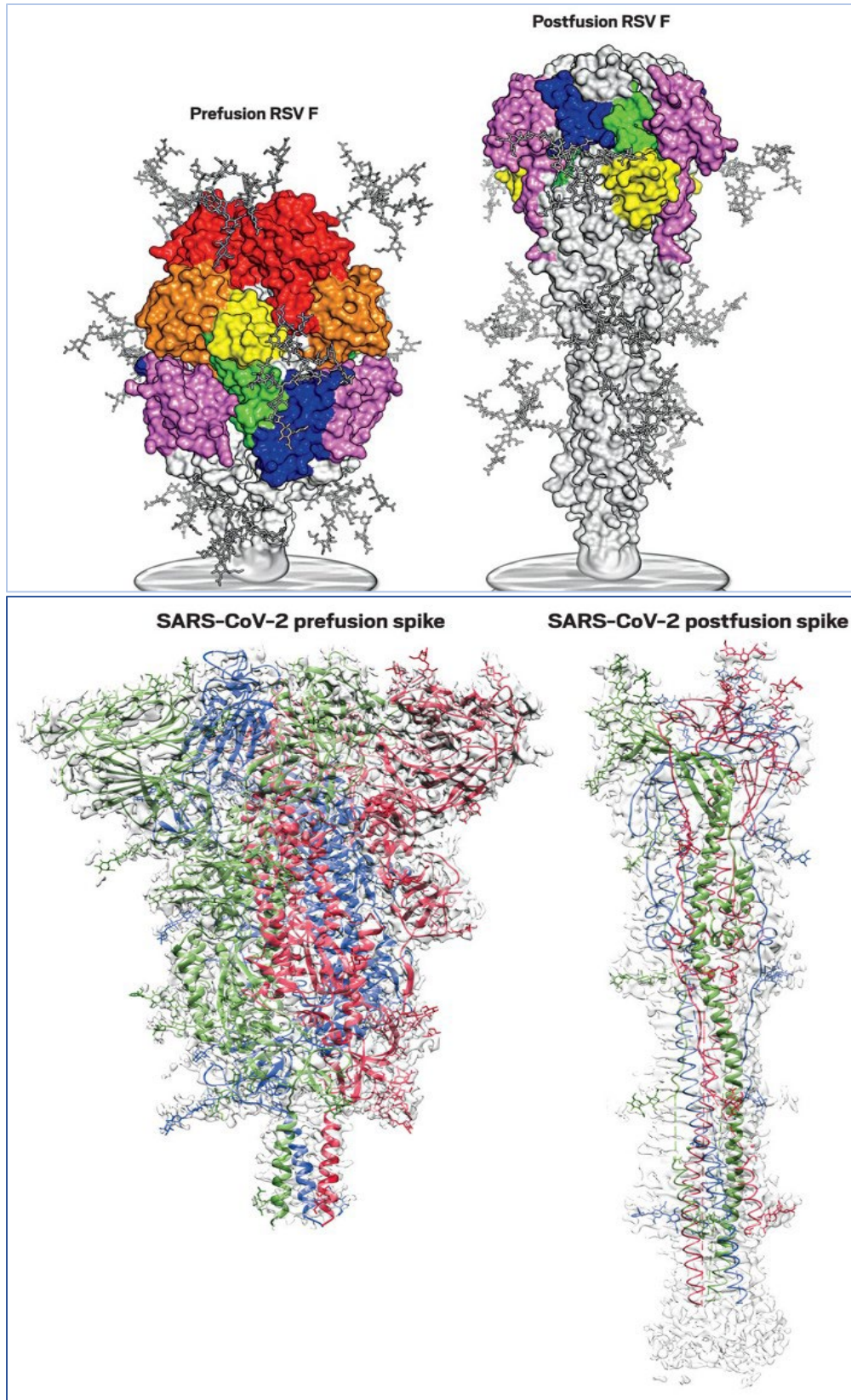
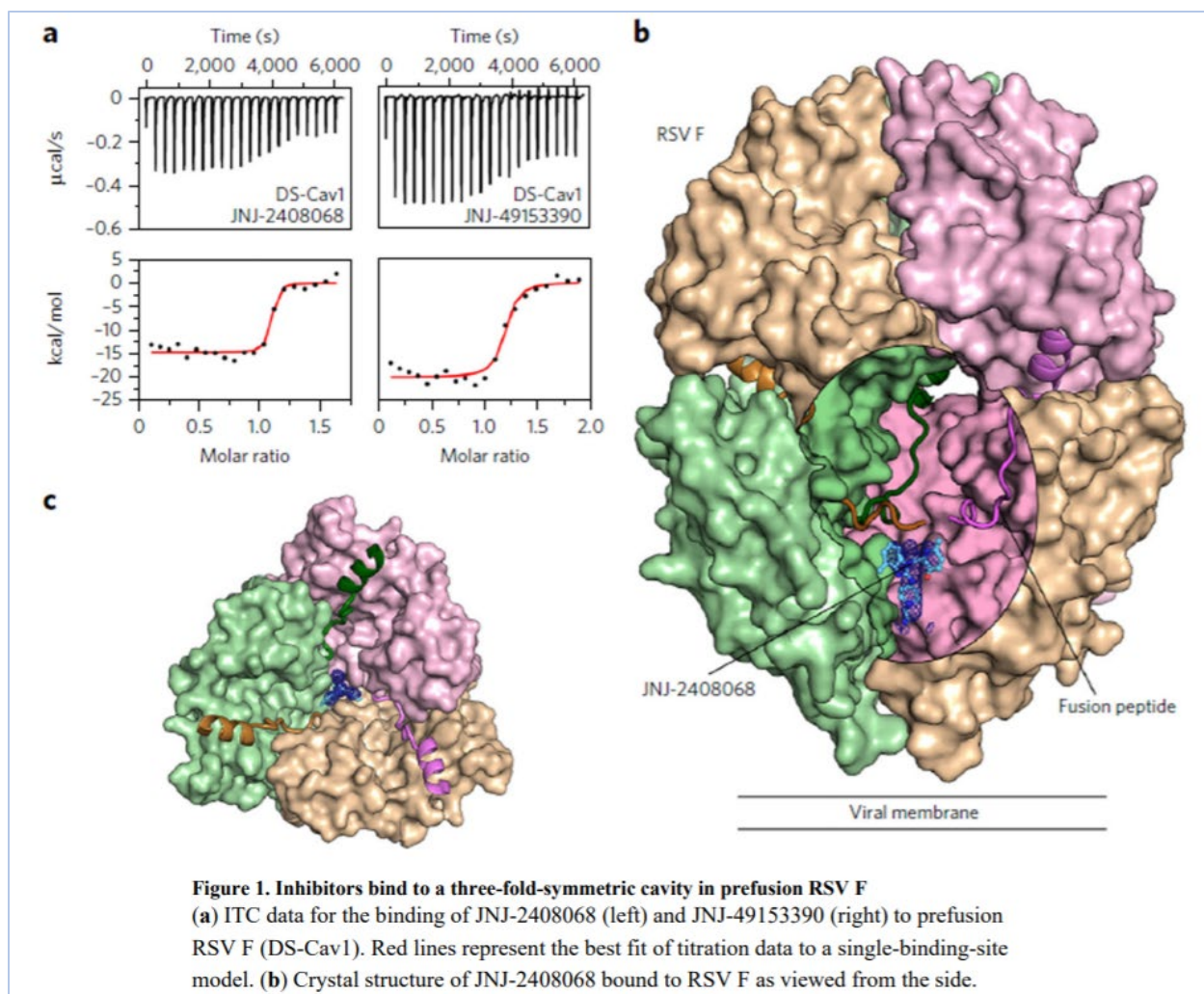


Figure 48: The dramatic conformational changes in RSV and SARS-CoV-2 are key and potential clues.



**Figure 1. Inhibitors bind to a three-fold-symmetric cavity in prefusion RSV F**  
**(a)** ITC data for the binding of JNJ-2408068 (left) and JNJ-49153390 (right) to prefusion RSV F (DS-Cav1). Red lines represent the best fit of titration data to a single-binding-site model. **(b)** Crystal structure of JNJ-2408068 bound to RSV F as viewed from the side.

In addition to those for RSV, small-molecule fusion inhibitors against other class I viral fusion glycoproteins such as influenza hemagglutinin (HA) and HIV-1 Env have been previously described. The majority of these inhibitors block either receptor binding<sup>32–35</sup> or the conformational changes required for fusion<sup>36–40</sup>. Of the latter group, the influenza fusion inhibitor umifenovir (Arbidol) may be most similar to the RSV compounds described here in that docking experiments suggest that Arbidol binds a cavity along the three-fold trimeric axis of influenza HA and interacts with the fusion peptide<sup>41</sup>. Interestingly, among class I fusion proteins for which structures are available, influenza HA has an interior fusion peptide arrangement most similar to RSV F (Supplementary Fig. 8). Thus, for RSV F, influenza HA and structurally related class I fusion proteins, cavities adjacent to the fusion peptide may represent promising targets for the development of small-molecule antivirals.

Figure 49: **JNJ-2408068<sup>648</sup> is a small molecule which prevented the transition of RSV F from pre-fusion to post-fusion.** The existence of other “*small-molecule fusion inhibitors against other class I viral fusion glycoproteins*” suggests that this may be a paradigm or platform approach for affordable (low-cost) drugs which may serve as an alternative or used as an adjuvant to (high-cost) vaccines for less affluent nations.

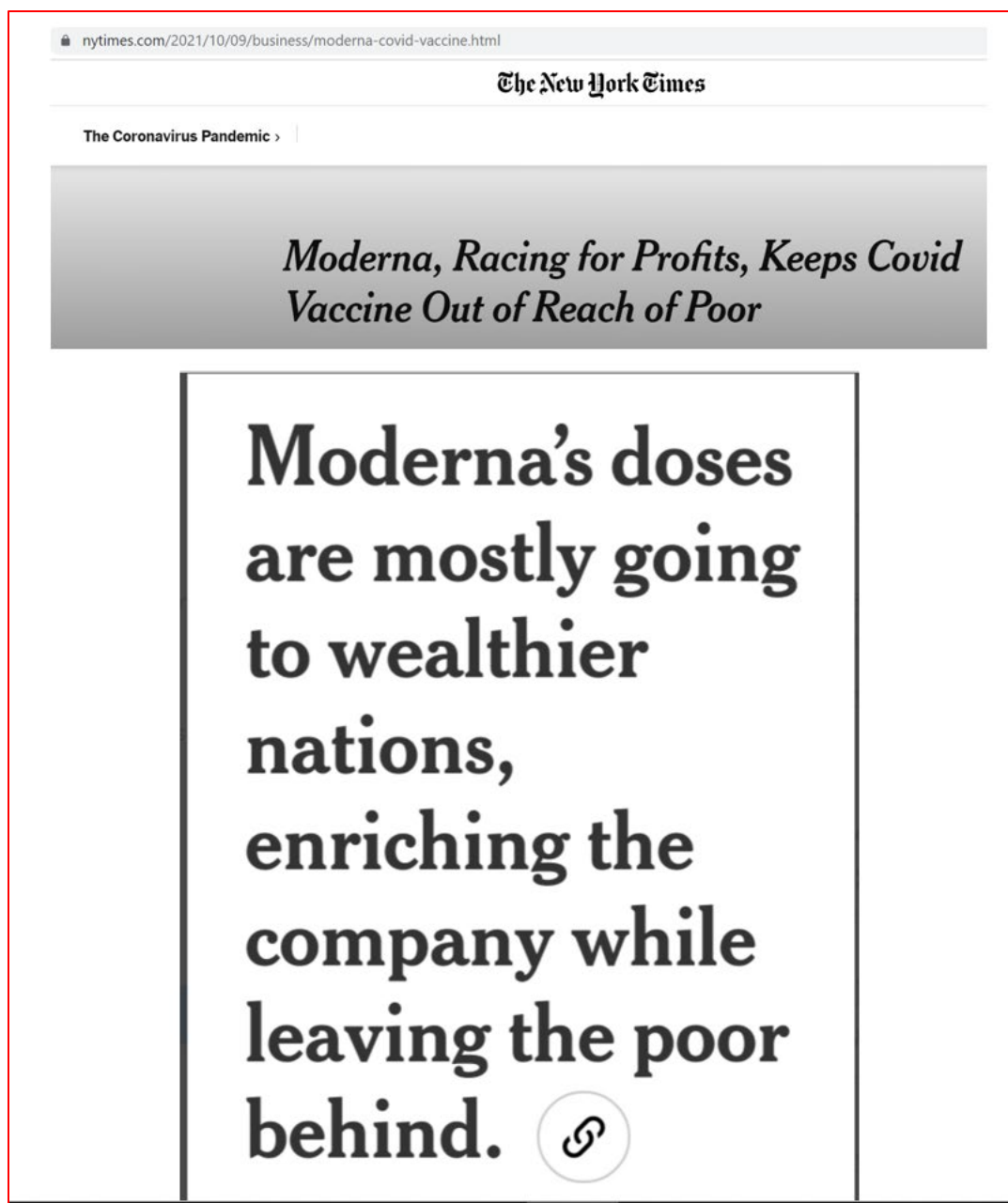


Figure 50: Alternates to vaccines may be necessary due to global inequity<sup>649</sup>. But, small molecules may not be the definitive alternative, rather it may be one cautious option. Use of small molecules have witnessed milestones<sup>650</sup>, boom (acyclovir<sup>651</sup>) and busts (deaths due fialuridine<sup>652</sup> during human clinical trials). Specificity, bio-availability and cytotoxicity could pose insurmountable barriers, not to mention what might be observed when we must account for phylogenetics, genomics and co-morbidities. Unless designed, we wouldn't know what may happen if these factors were stratified by socio-demographics.



## WHERE CAN WE GO FROM HERE? SHOULD WE LOOK FAR BACK TO THINK FAR AHEAD?

Pandemic-driven recession, shortages and decrease in productivity could erase small gains made by poor nations in their preparation to fight poverty<sup>653</sup>. The affluent response to the current pandemic may be beyond the reach of many nations, for many decades. This pandemic is not the last episode of health crisis. CoVID-19 has crippled most of the impoverished nations and regressed their attempts to grow human capital essential for economic recovery, social development and plight for freedom.

The less fortunate world inhabited by 80% of the global population needs a “platform” strategy to mitigate the risks from assault by microbes. The affluent platforms of vaccination, monoclonal antibodies and small molecule<sup>654</sup> therapeutics<sup>655</sup> may be “band-aids” for 80% of the world’s population catalyzed by unreliable (pseudo?) magnanimity from fickle philanthropists and flash-in-the-pan “made for TV” acts of PR and marketing, smugly disguised as charity, unity and the elusive pursuit of equity.

In this context of globalization<sup>656</sup> (and its discontent<sup>657</sup>) we may wish to look back to review the promise of a platform<sup>658</sup> approach which relies on natural biomimicry involving plants. Plant biologics may become affordable for less affluent nations. Preventative therapeutics through strategic convergence may be deliberately made *domestic-by-design* by incorporating components within elements of natural vegetation and/or household flowering plants (rose, marigold, tulip). Such plants, naturally, are woven into the fabric of our daily lives (humans, animals), the global economy as well as economic history<sup>659</sup>.

The potential of plant-based solutions was unleashed 30 years ago<sup>660</sup> but its promise<sup>661</sup> for global health still seems utopian due to fake sensationalism. Lack of progress due to *the march of unreason*<sup>662</sup> sponsored by diabolical groups<sup>663</sup> such as Greenpeace<sup>664</sup> and disenfranchised inhuman individuals such as Danilo Ramos<sup>665</sup> are one reason why millions of children are still suffering from preventable blindness (before they turn 5 years old) from xerophthalmia<sup>666</sup>, partially due to lack or reduced dietary intake of Vitamin A. Twenty five years ago scientists<sup>667</sup> delivered a feasible solution, embedded in the staple food, rice. Genetically supplementing *Oryza sativa* (rice) with phytoene synthase, an enzyme from daffodils (*Narcissus pseudonarcissus*), leads to the accumulation of phytoene, a precursor in the pathway of Vitamin A biosynthesis. Consumption of *golden rice*<sup>668</sup> was aimed at lifting many boats by enhancing a staple low-cost food (rice) with precursor for Vitamin A, a measure<sup>669</sup> to reduce preventable morbidities due to xerophthalmia. Irrational stupidity and suppression of science<sup>670</sup> are reasons why adoption<sup>671</sup> and implementation is sluggish<sup>672</sup> (while increasing access to fake rice products instead of Golden rice<sup>673</sup>).

Early scientific milestones in transgenic plant biologics includes expression of HBV (Hepatitis B) surface antigen<sup>674</sup>, demonstration of its immunogenic<sup>675</sup> potential and oral immunization<sup>676</sup> from edible produce (lettuce<sup>677</sup>, potatoes<sup>678</sup>) to prevent infection by Hepatitis B virus (HBV<sup>679</sup>). In the “whole virus” approach Norwalk virus<sup>680</sup> (Norovirus) and human papillomavirus-like particles<sup>681</sup> (HPV VLPs created with HPV type 11 L1 major capsid protein<sup>682</sup>) were introduced in tobacco and potato. In 2007, HIV<sup>683</sup> and influenza<sup>684</sup> virus-like particles (VLP) were added to the list of progress<sup>685</sup> followed by recent vaccine candidates (influenza virus<sup>686</sup>) including SARS-CoV-2<sup>687</sup> related recent advances<sup>688</sup>.

The past three decades of progress in plant biologics<sup>689</sup> (specifically for anti-virals) begs to ask what is preventing the mass adoption of transgenic plant based vaccine-type alternatives, especially when it may be a low-cost option for several poor nations?

## Imagine ...

The “note” (page 87) comes to life in the form of viral antigens and virus-like particles (VLPs) which may be used to elicit immune responses in humans, when administered orally (edible produce) or through sublingual/nasal modes.

Imagine, if, VLPs are introduced in roses (rose rosette virus, fimo<sup>690</sup>; rose yellow mosaic virus, poty<sup>691</sup>) or tulips (tulip-breaking virus<sup>692</sup>, potyviridae), then the following may be feasible.

Using RRV<sup>693</sup> or RYMV<sup>694</sup> as vectors, perhaps we could create roses specifically expressing VLPs for Ebola – Rosabola or the Rose Ebola. We could create a rose specifically expressing VLPs for SARS-CoV-2 and variants – the Rose CoVID-X. Roses expressing VLPs for flu and variants – the Rose FluX. Couldn't we generate Tulip for viruses by name and strain? If the VLPs are expressed in the sap or in the petals, then a teaspoon of crushed stem/leaf from rose/tulip will contain millions/billions of impotent virions (VLPs) – use a drop of this diluted sap “mush” under the tongue (sublingual). If VLPs are in petals, even better. Rose petals are edible but instead of eating just leave it under the tongue. Of course, edible versions are also possible. Naturally, the VLPs enter your body and voila! Can natural biomimicry help to make this list<sup>695</sup> even longer and better prepare the world with plants to combat other microbes?

By extending this hypothetical *modus operandi* to other plants as vehicles for VLPs, imagine the inclusion of tomatoes<sup>696</sup> (*Solanum lycopersicum*), basil (*Ocimum basilicum*), tulsi (*Ocimum tenuiflorum*) and neem<sup>697</sup> (*Azadirachta indica*). Plant-based biologics may reach communities under perpetual economic constraints where vaccines may not reach (in significant doses) to make a lasting difference.

With even more imagination, we may improve bioengineering design to reduce the number of virus-specific flowers and plants by adopting a combo<sup>698</sup> approach. What “if” immunogenic epitopes from viral antigens (SARS-CoV-2, Influenza, Ebola, Variola) were embedded/inserted (with spacers, in between) in plant virus vectors? Mix and match modular blocks of epitopes (think drag and drop) using a plant virus vector to sow yet another “platform” strategy for VLPs and/or other tools to create another hypothetical **Rose X** or **neem tree** which is not specific for Ebola or SARS or flu but the VLPs may confer “combo” immunity for all three by transforming the “epitopes on a string” suggestion into reality (?). Almost 30-years ago, an epitope derived from VP1 of foot-and-mouth disease virus (FMDV) was cloned into the CPMV (cowpea mosaic virus) genome encoding the small (S) coat protein. Chimeric S protein produced in plants infected with the insertion reacted with FMDV-specific antiserum<sup>699</sup>. Thus, plant virus vectors may be used as antigen presentation systems (not just one antigen but multiple epitopes).

These hypothetical and imaginary suggestions involving cross-species immunogens and tools for immunity are based on abstractions and lessons extracted from seminal research advances which defied conventional wisdom. Immune responses involving cross-kingdom RNA<sup>700</sup> interference was uncovered as one ammunition in our arms race with pathogens. The latter represents potentially new<sup>701</sup> mechanisms for disease control. These advances coupled with reverse genetic engineering and classical mutagenesis<sup>702</sup> are potential instruments to dissect the molecular basis of virulence, a key to controlling spread of disease. Convergence<sup>703</sup> of diverse ideas<sup>704</sup> from different domains<sup>705</sup> may be a compass for serendipitous minds prepared to insert square pegs in round holes and unafraid to discard roadmaps.



Temporarily, for the last segment of our *imagination* and hypothetical *what if* scenarios, let us explore the *least pragmatic suggestion* which may present cacophony of problems upon rational analysis and stir conundrums for molecular medicine verging on science fiction in the domain of public health.

Mosquitoes (genus *Anopheles*<sup>706</sup>) in molecular medicine and public health are often linked<sup>707</sup> with malaria<sup>708</sup> and the epic discoveries<sup>709</sup> in the 19<sup>th</sup> century by Ronald<sup>710</sup> Ross<sup>711</sup> which was inspired by Charles Louis Alphonse<sup>712</sup> Laveran<sup>713</sup>. After more than a century, a vaccine for malaria<sup>714</sup> has finally<sup>715</sup> emerged and clinical trials<sup>716</sup> with RTS,S/AS01<sup>717</sup> (against malarial parasite *Plasmodium falciparum* N54) were modestly (39%) successful yet approve<sup>718</sup> for immunization of children < 2 years old, in Malawi, sub-Saharan Africa. The cost for vaccine development<sup>719</sup> and infrastructure<sup>720</sup> for immunization (as well as “hidden” costs<sup>721</sup>) makes it difficult for nations under economic constraints to pursue meaningful strategic<sup>722</sup> vaccine programs or support mass immunization, even when/if vaccines were contributed.



Figure 51: Sir Ronald Ross, Mrs. Ross, Mr. Mahomed Bux and assistants at the laboratory in Calcutta, India (now referred to as “Kolkata”) where the life history of the malaria parasite (using birds) was elucidated circa 1898. This seminal work may have catalyzed the establishment of the Calcutta School of Hygiene and Tropical Medicine<sup>723</sup> and associated hospitals for tropical diseases<sup>724</sup> as well as medical research<sup>725</sup> (photograph<sup>726</sup> courtesy of the London School of Hygiene and Tropical Medicine, UK).

In other words, almost 80% of the global population may remain unvaccinated (more or less) due to high cost of infrastructure and (the lack of) human capital necessary for immunization efforts, even if traditional or non-traditional vaccine (or alternatives) were made available, at least in principle.

Genetic modifications<sup>727</sup> aimed at controlling<sup>728</sup> the proliferation of vector-borne<sup>729</sup> (for example, mosquitoes) parasitic<sup>730</sup> and viral<sup>731</sup> diseases is a strategic breakthrough of immense value for reducing the disease burden and improving public health in most parts of the less-affluent world.

Insect vectors which transmit microbes to human hosts, are, therefore, Nature's mechanism for cross-kingdom "infection". In a biomimicry approach, it may be possible to apply a "reverse" design. Could we use Nature's mechanism to "infect" people with vectors (mosquitoes) carrying antigens and virus-like particles (VLPs)? If we leave aside ethical, legal and societal issues from *pre-planned* mosquito bite, this natural biomimicry (*insects as immunization delivery platform*) may be the Holy Grail for low-cost immunization delivery for billions of people who cannot afford infrastructure costs associated with the "last mile" delivery which continues to pose insurmountable problems for vaccination programs.

Genetic engineering of viruses which infect *Anopheles*<sup>732</sup> and *Aedes*<sup>733</sup> mosquitoes are the targets for development of vectors to shuttle antigens and for creating virus-like particles for specific viruses, for example, Ebola, SARS-CoV-2, and others. Targets for genetic modification may include re-engineering tissue-specific regulation of the small interfering RNA (siRNA) pathway controlled by the double stranded (ds) RNA binding protein Loqs2<sup>734</sup> (and its genetic circuit) which appears to be specific to *Aedes aegypti* mosquito. Are there Loqs2 equivalents or similar pathways in other mosquitoes?

Differential regulation by nucleic acid binding proteins are key modulators. In humans, antiviral effectors encoded by IFN-stimulated genes (ISGs) inhibits SARS-CoV-2 replication. 2'-5'-oligoadenylate synthetase 1 (OAS1) recognizes short stretches of double-stranded RNA (dsRNA) in the 5'- untranslated region (UTR) of SARS-CoV-2 RNA and activated degradation by RNase L. But, *only* individuals with splice-acceptor single nucleotide *polymorphism* in OAS1 (G at Rs10774671)<sup>735</sup> expressed a C-terminal prenylated form of OAS1 (p46) which targets OAS1 to perinuclear structures rich in viral dsRNA. Thus, individuals with this polymorphism had reduced susceptibility to infection and less severe CoVID-19 (non-prenylated OAS1 was diffused and unable to initiate detectable block to SARS-CoV-2 replication).

Because *Aedes aegypti* is a vector for number of viral diseases (dengue, Zika, Chikungunya, yellow fever) and adept at using RNA interference to selectively degrade viral genomes, it may be worth exploring comparative molecular mechanisms of action and rate-limiting factors where [1] pathogens deliver small RNAs (sRNAs are often generated from transposable element regions<sup>736</sup>) into host cells to suppress host immunity versus [2] hosts (*Arabidopsis* cells) which secrete exosome-like extracellular vesicles to deliver sRNAs into pathogens (*Botrytis cinerea*) to silence genes for pathogenicity (Cai *et al*, 2018).

These RNA tools<sup>737</sup> and their association with transposable elements may be indicative of an evolutionary “shuttle service” (mobile “security” forces for hire) for cross-species and/or cross-kingdom exchanges to influence the state of immunity (immunity-on-demand). Other “shuttle services” from mobile “gene editing” elements on transposons<sup>738</sup> may be combined with “immunity-on-demand” tools creating a “drag and drop” biological “menu of choices” which may be mixed and matched, selectively, deployed-on-demand and/or activated/inhibited by allosteric<sup>739</sup> switching, responsive to gradient of spatio-temporal specificity modulated by time, temperature, tissue-specificity as well as external feedback or feed-forward (for example, ligands, small molecules, allosteric<sup>740</sup> enantiomers<sup>741</sup>).

These amorphous clues may be haphazard at a first glance but abstractions by astute experts may distill creative tool kits which may tune the engines of biomimicry to reverse engineer counter-intuitive solutions. Instead of billion-dollar vaccination infrastructures, imagine mobile mosquito *bite squads*<sup>742</sup> (small kiosks with *VLP-in-every-bite* mosquitoes) on cycle-rickshaws<sup>743</sup> or bullock-carts<sup>744</sup> wandering through villages to immunize billions of people (perhaps even animals). How about mosquitoes as global health public goods? “If it’s *inaccessible* to the poor<sup>745</sup>, it is neither radical nor revolutionary<sup>746</sup>.”

## NOTE · ECONOMIC HISTORY (indicated as ref 659 on pg. 83, 3rd para, last line)

### *Flowers and Trees as Anti-Virals: Roses for Ebola, Tulips for Variola, Neem Tree for Everything*

Circa 1593-1594, Carolus Clusius, (Charles de l'Ecluse, 1526-1609) a botanist, begun a botanical garden at the University of Leiden, to cultivate tulips<sup>747</sup> in Netherlands<sup>748</sup>. Originally a wildflower, tulips were cultivated in the Ottoman Empire (Turkey, *-stan* states), as early as 1,000 AD and later imported to Holland in the 17<sup>th</sup> century, where they became a formidable economic force. It was the first economic bubble to explode<sup>749</sup> circa 1637. Tulips were key to the Dutch economy<sup>750</sup> in the form of auctions (eBay). *Tulipmania* coincided with the formal establishment of the Amsterdam Stock Exchange in 1602. Tulips catalyzed the Dutch economy to achieve a high standard of living by the middle of the 17th century. The affluence spawned a Golden Age, for the Dutch, typified by the artist Rembrandt van Rijn (1606–1669).

The bursting of the bubble of the tumultuous tulip economy may have been evident from the price of *Semper Augustus*, a variegated tulip with distinct striations on its petals. Around 1636, the price of the bulb of *Semper Augustus* was about \$34,584 and as high as \$97,200 (by other forms of valuation). These colorful patterns were caused by infection due to potyviruses<sup>751</sup> which are plant viruses with RNA (positive-strand) genomes, which alters distribution of pigments in the petal due to virus replication. A specific potyvirus, the tulip-breaking virus (TBV), causes successive generations of the bulb to shrink until it can no longer flower. Hence, the incredible price<sup>752</sup> of the *Semper Augustus* bulb. Tulipmania was brought to its knees by potyviruses (circa 1636-1637) and almost destroyed<sup>753</sup> the Dutch economy. The decline of the Dutch economy, due to potyviruses, coincided with the bubonic plague, which ravaged The Netherlands<sup>754</sup>, a fifth of the population dying in Amsterdam in the period 1635-1636.



VIROLOGY 8, 99-115 (1959)

### **Classification of Elongated Plant Viruses on the Basis of Particle Morphology**

J. BRANDES AND C. WETTER

*Biologische Bundesanstalt für Land- und Forstwirtschaft; Institut für landwirtschaftliche Virusforschung, and Institut für Virusserologie, Braunschweig, Germany*

Accepted February 6, 1959

Figure 52: Colorful striations of *Semper Augustus* caused by infection due to tulip-breaking virus (TBV). Identification<sup>755</sup> of potyvirus (1959).

## ACKNOWLEDGEMENTS

The author is thankful to Brittany [Newell](#) (section 2), Larry [Gold](#) (section 5), Andrew [Fire](#) and Greg [Winter](#) (for section 6) as well as John [Rossi](#), Britt [Glaunsinger](#) and Sanjay [Sarma](#) (for general review). The critical review by Jack [Szostak](#) expressed skepticism about the sensitivity of the optical signal (from a nano quantum dot conjugated DNA aptamer bound to the target viral protein in a sample of saliva from humans or animals). [Jack's](#) skepticism focused on the *comparative sensitivity* between the proposed hypothetical optical signal versus signal from traditional qRT-PCR (reverse transcriptase-polymerase chain reaction) of viral RNA genome. (Jack Szostak is Chairman of the [Scientific Advisory Board of Moderna Inc.](#), Cambridge, MA). Comments from [Jason McLellan](#) and caution from [Katalin Karikó](#) (regarding the suggestions in the concluding section) is deeply appreciated.

*This note is a suggestion and a proposal from [Shoumen Datta](#) (SD) based on published scientific research. **It is not new and copyright free.** It may be used by anybody for any purpose without any need to cite or credit the author. If you have questions please email [shoumen@mit.edu](mailto:shoumen@mit.edu) and/or [sdatta8@mgh.harvard.edu](mailto:sdatta8@mgh.harvard.edu)*



## REFERENCES

- <sup>1</sup> Subirana, Brian, et al. “Hi Sigma, Do I Have the Coronavirus?: Call for a New Artificial Intelligence Approach to Support Health Care Professionals Dealing with the COVID-19 Pandemic.” April 2020. <http://arxiv.org/abs/2004.06510> and <https://arxiv.org/ftp/arxiv/papers/2004/2004.06510.pdf>
- <sup>2</sup> Massachusetts General Hospital (2020) New Tool Can Detect COVID-19 Outbreaks in US Counties. <https://www.massgeneral.org/news/press-release/New-tool-can-detect-covid-19-outbreaks-in-us-counties> and <https://analytics-modeling.shinyapps.io/outbreakdetection/>
- <sup>3</sup> Engels, D. W., et al. “Networked Physical World: Automated Identification Architecture.” *Proceedings. The Second IEEE Workshop on Internet Applications. WIAPP 2001*, IEEE Comput. Soc, 2001, pp. 76–77. doi:10.1109/WIAPP.2001.941872 <https://ieeexplore.ieee.org/document/941872>
- <sup>4</sup> Sarma, Sanjay, Brock, David and Ashton, Kevin (2000) *The Networked Physical World: Proposals for Engineering the Next Generation of Computing, Commerce & Automatic-Identification*. White Paper WH-001. October 1, 2000. MIT Auto-ID Center, Massachusetts Institute of Technology. [https://cocoa.ethz.ch/downloads/2014/06/None\\_MIT-AUTOID-WH-001.pdf](https://cocoa.ethz.ch/downloads/2014/06/None_MIT-AUTOID-WH-001.pdf)
- <sup>5</sup> Susan Symington, William Polk and Murugiah Souppaya (2020) *Internet of Things (IoT) Device Network-Layer Onboarding and Lifecycle Management*. NIST Cybersecurity White Paper (Draft). <https://nvlpubs.nist.gov/nistpubs/CSWP/NIST.CSWP.09082020-draft.pdf>
- <sup>6</sup> Chinese Numerology [https://en.wikipedia.org/wiki/Chinese\\_numerology](https://en.wikipedia.org/wiki/Chinese_numerology)
- <sup>7</sup> Shambhavi Shubham, Jan Hoinka, Soma Banerjee, Emma Swanson, Jacob A. Dillard, Nicholas J. Lennemann, Teresa M. Przytycka, Wendy Maury and Marit Nilsen-Hamilton (2018) A 2'FY-RNA Motif Defines an Aptamer for Ebolavirus Secreted Protein. *Nature Scientific Reports* 2018 Aug 17; 8(1):12373 doi: 10.1038/s41598-018-30590-8 [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6098113/pdf/41598\\_2018\\_Article\\_30590.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6098113/pdf/41598_2018_Article_30590.pdf)
- <sup>8</sup> Jakub Ptacek, Dong Zhang, Liming Qiu, Sven Kruspe, Lucia Motlova, Petr Kolenko, Zora Novakova, Shambhavi Shubham, Barbora Havlinova, Petra Baranova, Shi-Jie Chen, Xiaoqin Zou, Paloma Giangrande, Cyril Barinka. Structural basis of prostate-specific membrane antigen recognition by the A9g RNA aptamer. *Nucleic Acids Research*, gkaa494. <https://doi.org/10.1093/nar/gkaa494>
- <sup>9</sup> Datta, Shoumen (2020) *CITCOM – An Incomplete Review of Ideas and Facts about SARS-CoV-2* <https://dspace.mit.edu/handle/1721.1/128017> and <https://dspace.mit.edu/handle/1721.1/111021>
- <sup>10</sup> Fairbanks, Antony J. (2017) “The ENGases: Versatile Biocatalysts for the Production of Homogeneous N-Linked Glycopeptides and Glycoproteins.” *Chemical Society Reviews*, vol. 46, no. 16, 2017, pp. 5128–46. doi:10.1039/C6CS00897F. <https://pubs.rsc.org/en/content/articlepdf/2017/cs/c6cs00897f>
- <sup>11</sup> Khargonekar, Pramod P. (2020) Kalman Filtering, Sensor Fusion, and Eye Tracking [https://faculty.sites.uci.edu/khargonekar/files/2020/08/KFSensorFusion\\_PPK.pdf](https://faculty.sites.uci.edu/khargonekar/files/2020/08/KFSensorFusion_PPK.pdf)
- <sup>12</sup> NIST Differential Privacy Engineering (2018) <https://www.nist.gov/itl/applied-cybersecurity/privacy-engineering/collaboration-space/focus-areas/de-id/tools#dpchallenge>

- <sup>13</sup> Victoria Morgan, Lisseth Casso-Hartman, David Bahamon-Pinzon, Kelli McCourt, Robert G. Hjort, Sahar Bahramzadeh, Irene Velez-Torres, Eric McLamore, Carmen Gomes, Evangelyn C. Alocilja, Shoumen Palit Austin Datta and Diana C. Vanegas (2019) *Sensor-as-a-Service: Convergence of Sensor Analytic Point Solutions (SNAPS) and Pay-A-Penny-Per-Use (PAPPU) Paradigm as a Catalyst for Democratization of Healthcare in Underserved Communities*. *Diagnostics* 2020, 10 (1), 22  
<https://doi.org/10.3390/diagnostics10010022>  
“SNAPS TRILOGY” MIT Libraries <https://dspace.mit.edu/handle/1721.1/123983>
- <sup>14</sup> Winn, Zach (2020) Real-time data for a better response to disease outbreaks.  
<https://news.mit.edu/2020/kinsa-health-0821>
- <sup>15</sup> Lamb, James (2020) ADD Appendix – Description of Major Components (*personal communication*)  
<https://github.com/jameslamb>
- <sup>16</sup> Zamecnik PC, Stephenson ML.(1978) *Inhibition of Rous sarcoma virus replication and cell transformation by a specific oligodeoxynucleotide*. *Proc Natl Acad Sci (US)*. 1978 January; 75(1):280-4. doi: 10.1073/pnas.75.1.280 [www.ncbi.nlm.nih.gov/pmc/articles/PMC411230/pdf/pnas00013-0285.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC411230/pdf/pnas00013-0285.pdf)
- <sup>17</sup> Cohn, Danny M., et al. “Antisense Inhibition of Prekallikrein to Control Hereditary Angioedema.” *New England J of Med*. vol. 383, no. 13, September 2020, pp. 1242–1247. doi:10.1056/NEJMoa1915035
- <sup>18</sup> APPENDIX Figure 1: Description of Components <https://github.com/shoumendatta/ADD-DIGITAL>
- <sup>19</sup> McLamore, Eric (2019) Surveillance Smartphone App (*personal communication*)  
<https://emclamor.wixsite.com/mclamorelab>
- <sup>20</sup> Shmerling, Robert (2020) *Which test is best for COVID-19?* Harvard Medical School. 21 Sept 2020.  
<https://www.health.harvard.edu/blog/which-test-is-best-for-covid-19-2020081020734>
- <sup>21</sup> Classification: True vs. False and Positive vs. Negative  
<https://developers.google.com/machine-learning/crash-course/classification/true-false-positive-negative>
- <sup>22</sup> Candela M, Luconi V, Vecchio A. *Impact of the COVID-19 pandemic on the Internet latency: A large-scale study*. *Computer Networks*. 2020 December 9; 182:107495. doi: 10.1016/j.comnet.2020.107495. Epub 2020 August 20. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7439971/pdf/main.pdf>
- <sup>23</sup> World Economic Forum (2016) *4 billion people still don't have internet access*.  
<https://www.weforum.org/agenda/2016/05/4-billion-people-still-don-t-have-internet-access-here-s-how-to-connect-them/>
- <sup>24</sup> Likert Scale <https://www.sciencedirect.com/topics/psychology/likert-scale>
- <sup>25</sup> Roh, Yuji, et al. *A Survey on Data Collection for Machine Learning*. August 2019.  
<http://arxiv.org/abs/1811.03402> and <https://arxiv.org/pdf/1811.03402.pdf>
- <sup>26</sup> CDC - 2019-nCoV Real-Time RT-PCR Diagnostic Panel. [www.fda.gov/media/134920/download](http://www.fda.gov/media/134920/download)
- <sup>27</sup> Hutson, Matthew. “Artificial-Intelligence Tools Aim to Tame the Coronavirus Literature.” *Nature*, June 2020, pp. d41586-020-01733–37. doi:10.1038/d41586-020-01733-7
- <sup>28</sup> Kanter, James Max, et al. “Machine Learning 2.0 : Engineering Data Driven AI Products.” July 2018.  
<http://arxiv.org/abs/1807.00401> and <https://arxiv.org/pdf/1807.00401.pdf>
- <sup>29</sup> Collection of papers and books on FEATURES (200MB) <http://bit.ly/BOOKS-FEATURES>

- <sup>30</sup> Escalante, Hugo Jair. “Automated Machine Learning - Brief Review at the End of the Early Years.” August 2020. <http://arxiv.org/abs/2008.08516> and <https://arxiv.org/pdf/2008.08516.pdf>
- <sup>31</sup> Kanter, James Max, and Kalyan Veeramachaneni. “Deep Feature Synthesis: Towards Automating Data Science Endeavors.” *2015 IEEE International Conference on Data Science and Advanced Analytics (DSAA)*, IEEE, 2015, pp. 1–10. doi:10.1109/DSAA.2015.7344858. [http://axperia-ventures.com/wp-content/uploads/2015/10/DSAA\\_DSM\\_2015.pdf](http://axperia-ventures.com/wp-content/uploads/2015/10/DSAA_DSM_2015.pdf)
- <sup>32</sup> Gilad Katz, Eui Chul Richard Shin and Dawn Song (2016) “ExploreKit: Automatic Feature Generation and Selection.” *2016 IEEE 16th International Conference on Data Mining*, IEEE, 2016, pp. 979–84. doi:10.1109/ICDM.2016.0123 <https://people.eecs.berkeley.edu/~dawnsong/papers/icdm-2016.pdf>
- <sup>33</sup> Data Distribution Service <https://www.dds-foundation.org/what-is-dds-3/>
- <sup>34</sup> Berke EM. *Geographic Information Systems: recognizing the importance of place in primary care research & practice*. J Am Board Fam Med. 2010 Jan-Feb; 23(1):9-12. doi: 10.3122/jabfm.2010.01.090119 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3174470/pdf/nihms-323401.pdf>
- <sup>35</sup> Tebani, A., Gummesson, A., Zhong, W. *et al.* Integration of molecular profiles in a longitudinal wellness profiling cohort. *Nat Commun* 11, 4487 (2020). <https://doi.org/10.1038/s41467-020-18148-7> <https://www.nature.com/articles/s41467-020-18148-7.pdf>
- <sup>36</sup> Pennisi, Elizabeth. “Massive Project Reveals Complexity of Gene Regulation.” *Science*, vol. 369, no. 6509, 10 September 2020, pp. 1286–1287. doi:10.1126/science.369.6509.1286
- <sup>37</sup> Zahn, Laura M. “Custom Transcriptome.” *Science*, vol. 369, no. 6509, 10 September 2020, pp. 1316-17 doi:10.1126/science.abe4492
- <sup>38</sup> The GTEx Consortium. “The GTEx Consortium Atlas of Genetic Regulatory Effects across Human Tissues.” *Science*, vol. 369, no. 6509, 10 September 2020, pp. 1318–1330. doi:10.1126/science.aaz1776 <https://science.sciencemag.org/content/sci/369/6509/1318.full.pdf>
- <sup>39</sup> Hutchinson, L., Romero, D. Precision or imprecision medicine?. *Nat Rev Clin Oncol* 13, 713 (2016) <https://doi.org/10.1038/nrclinonc.2016.190> & <https://www.nature.com/articles/nrclinonc.2016.190.pdf>
- <sup>40</sup> Gold, Larry (2020) *Personal Communication* <https://amzn.to/2QGSz1i>
- <sup>41</sup> Dodig-Crnković, Tea, et al. “Facets of Individual-Specific Health Signatures Determined from Longitudinal Plasma Proteome Profiling.” *EBioMedicine*, vol. 57, July 2020, p. 102854. doi:10.1016/j.ebiom.2020.102854 [www.thelancet.com/action/showPdf?pii=S2352-3964%2820%2930229-2](http://www.thelancet.com/action/showPdf?pii=S2352-3964%2820%2930229-2)
- <sup>42</sup> <https://somascandiscovery.com/publications/>
- <sup>43</sup> APTAMER (Collected Papers) <https://bit.ly/APTAMER>
- <sup>44</sup> She, Richard, et al. “Comprehensive and Quantitative Mapping of RNA–Protein Interactions across a Transcribed Eukaryotic Genome.” *Proceedings of the National Academy of Sciences*, vol. 114, no. 14, April 2017, pp. 3619–3624. doi:10.1073/pnas.1618370114 <https://www.pnas.org/content/pnas/114/14/3619.full.pdf>

---

<sup>45</sup> Pál, Gábor, et al. “Comprehensive and Quantitative Mapping of Energy Landscapes for Protein-Protein Interactions by Rapid Combinatorial Scanning.” *Journal of Biological Chemistry*, vol. 281, no. 31, August 2006, pp. 22378–22385. doi:10.1074/jbc.M603826200.

<https://www.jbc.org/content/281/31/22378.full.pdf>

<sup>46</sup> Petricoin, Emanuel F., et al. “Use of Proteomic Patterns in Serum to Identify Ovarian Cancer.” *The Lancet*, vol. 359, no. 9306, Feb. 2002, pp. 572–77. doi:10.1016/S0140-6736(02)07746-2

[http://fenyolab.org/presentations/Proteomics\\_Informatics\\_2014/pdf/02\\_petricoin\\_lancet.pdf](http://fenyolab.org/presentations/Proteomics_Informatics_2014/pdf/02_petricoin_lancet.pdf)

<sup>47</sup> Canna, Scott W., and Edward M. Behrens. “Making Sense of the Cytokine Storm: A Conceptual Framework for Understanding, Diagnosing, and Treating Hemophagocytic Syndromes.” *Pediatric Clinics of North America*, vol. 59, no. 2, April 2012, pp. 329–44. doi:10.1016/j.pcl.2012.03.002

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3368378/pdf/nihms370318.pdf>

<sup>48</sup> Tisoncik, J. R., et al. “Into the Eye of the Cytokine Storm.” *Microbiology and Molecular Biology Reviews*, vol. 76, no. 1, March 2012, pp. 16–32. doi:10.1128/MMBR.05015-11

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3294426/pdf/zmr16.pdf>

<sup>49</sup> Spit Shines <https://science.sciencemag.org/content/369/6507/1041/tab-pdf> and

<https://www.nejm.org/doi/pdf/10.1056/NEJMc2016359>

<sup>50</sup> Gold, Larry. “SELEX: How It Happened and Where It Will Go.” *Journal of Molecular Evolution*, vol. 81, no. 5–6, December 2015, pp. 140–143. doi:10.1007/s00239-015-9705-9

[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4661202/pdf/239\\_2015\\_Article\\_9705.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4661202/pdf/239_2015_Article_9705.pdf)

<sup>51</sup> De La Fuente, Adriana, et al. “Aptamers against Mouse and Human Tumor-Infiltrating Myeloid Cells as Reagents for Targeted Chemotherapy.” *Science Translational Medicine*, vol. 12, no. 548, June 2020, p. eaav9760 doi:10.1126/scitranslmed.aav9760

<sup>52</sup> Hopfield, J. J. “Kinetic Proofreading: A New Mechanism for Reducing Errors in Biosynthetic Processes Requiring High Specificity.” *Proc National Academy of Sciences*, vol. 71, no. 10, Oct. 1974, pp. 4135–4139

doi:10.1073/pnas.71.10.4135 <https://www.pnas.org/content/pnas/71/10/4135.full.pdf>

<sup>53</sup> Oran DP, Topol EJ. *Prevalence of Asymptomatic SARS-CoV-2 Infection : A Narrative Review*. *Annals of Internal Medicine*. 2020 September 1;173(5):362-367. doi: 10.7326/M20-3012. Epub 2020 June 3.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7281624/pdf/aim-olf-M203012.pdf>

<sup>54</sup> Gheblawi M, Wang K, Viveiros A, Nguyen Q, Zhong JC, Turner AJ, Raizada MK, Grant MB, Oudit GY. *Angiotensin-Converting Enzyme 2: SARS-CoV-2 Receptor and Regulator of the Renin-Angiotensin System: Celebrating the 20th Anniversary of the Discovery of ACE2*. *Circulation Research*. 2020 May 8; 126(10):1456-1474. doi: 10.1161/CIRCRESAHA.120.317015. Epub 2020 April 8.

[www.ncbi.nlm.nih.gov/pmc/articles/PMC7188049/pdf/res-126-10.1161.circresaha.120.317015.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC7188049/pdf/res-126-10.1161.circresaha.120.317015.pdf)

<sup>55</sup> López-Otín, Carlos, and Judith S. Bond. “Proteases: Multifunctional Enzymes in Life and Disease.” *Journal of Biological Chemistry*, vol. 283, no. 45, November 2008, pp. 30433–30437

doi:10.1074/jbc.R800035200 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2576539/pdf/30433.pdf>

- <sup>56</sup> Hoagland, M. B., Stephenson, M. L., Scott, J. F., Hecht, L. I., and Zamecnik, P. C. (1958) *A Soluble Ribonucleic Acid Intermediate in Protein Synthesis*. *J. Biol. Chem.* **231** 241–257.  
[https://pdfs.semanticscholar.org/aad2/985a4bcb40430260e3570683cb2b3b79f7e7.pdf?\\_ga=2.242690306.1804402263.1599417152-1424810635.1595859001](https://pdfs.semanticscholar.org/aad2/985a4bcb40430260e3570683cb2b3b79f7e7.pdf?_ga=2.242690306.1804402263.1599417152-1424810635.1595859001) ■ <https://www.jbc.org/content/280/40/e37>
- <sup>57</sup> Brenner, S., Jacob, F. & Meselson, M. (1961). *An unstable intermediate carrying information from genes to ribosomes for protein synthesis*. *Nature* **190** 576–581
- <sup>58</sup> Gros, F., Hiatt, H., Gilbert, W., Kurland, C. G., Risebrough, R. W. & Watson, J. D. (1961). *Unstable ribonucleic acid revealed by pulse labelling of Escherichia coli*. *Nature* **190** 581–585
- <sup>59</sup> Jacob, F. and Monod, J. (1961). *Genetic regulatory mechanisms in the synthesis of proteins*. *Journal of Molecular Biology* **3** 318–356
- <sup>60</sup> Monod, J., Changeux, J. P. and Jacob, F. (1963). *Allosteric proteins and cellular control systems*. *Journal of Molecular Biology* **6** 306–329
- <sup>61</sup> Alberts, Bruce and Frey, Linda (1970) *T4 Bacteriophage Gene 32: A Structural Protein in the Replication and Recombination of DNA*. *Nature* **227** 1313–1318  
<https://brucealberts.ucsf.edu/wp-content/uploads/2016/07/Alberts-and-Frey-1970-YO.pdf>
- <sup>62</sup> Tuerk, Craig and Gold, Larry (1990) *Systematic evolution of ligands by exponential enrichment: RNA ligands to bacteriophage T4 DNA polymerase*. *Science* **249** 505–510
- <sup>63</sup> Ellington, Andy D. and Szostak, Jack W. (1990) *In vitro selection of RNA molecules that bind specific ligands*. *Nature* **346** 818–822
- <sup>64</sup> Gold, Larry (2015) “SELEX: How It Happened and Where It Will Go.” *Journal of Molecular Evolution*, vol. 81, no. 5–6, December 2015, pp. 140–143. doi:10.1007/s00239-015-9705-9  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4661202/pdf/239\\_2015\\_Article\\_9705.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4661202/pdf/239_2015_Article_9705.pdf)
- <sup>65</sup> Lakhin AV, Tarantul VZ, Gening LV. (2013) *Aptamers: problems, solutions and prospects*. *Acta Naturae*. 2013 October, 5(4):34-43.  
[www.ncbi.nlm.nih.gov/pmc/articles/PMC3890987/pdf/AN20758251-19-034.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3890987/pdf/AN20758251-19-034.pdf)
- <sup>66</sup> Ku TH, Zhang T, Luo H, Yen TM, Chen PW, Han Y, Lo YH. (2015) *Nucleic Acid Aptamers: An Emerging Tool for Biotechnology and Biomedical Sensing*. *Sensors (Basel)*. 2015 July 6; 15(7): 16281-313 doi: 10.3390/s150716281 [www.ncbi.nlm.nih.gov/pmc/articles/PMC4541879/pdf/sensors-15-16281.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4541879/pdf/sensors-15-16281.pdf)
- <sup>67</sup> Kato, Teru, et al (2000) “In Vitro Selection of DNA Aptamers Which Bind to Cholic Acid.” *Biochimica et Biophysica Acta (BBA) - Gene Structure and Expression*, vol. 1493, no. 1–2, September 2000, pp. 12–18 doi:10.1016/S0167-4781(00)00080-4
- <sup>68</sup> Kalra, Priya, et al. “Simple Methods and Rational Design for Enhancing Aptamer Sensitivity and Specificity.” *Frontiers in Molecular Biosciences*, vol. 5, May 2018, p. 41. doi:10.3389/fmolb.2018.00041  
<https://www.frontiersin.org/articles/10.3389/fmolb.2018.00041/pdf>
- <sup>69</sup> Strauss S, Nickels PC, Strauss MT, Jimenez Sabinina V, Ellenberg J, Carter JD, Gupta S, Janjic N, Jungmann R. (2018) *Modified aptamers enable quantitative sub-10-nm cellular DNA-PAINT imaging*. *Nature Methods*. 2018 September; 15(9):685-688. doi: 10.1038/s41592-018-0105-0 Epub 2018 Aug 20.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6345375/pdf/emss-78630.pdf>



- <sup>70</sup> Kato Teru, Yano K, Ikebukuro K, Karube I. (2000) *Interaction of three-way DNA junctions with steroids*. *Nucleic Acids Res.* 2000 May 1; 28(9):1963-1968. doi: 10.1093/nar/28.9.1963  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC103303/pdf/gkd319.pdf>
- <sup>71</sup> Sun, B.B., Maranville, J.C., Peters, J.E. *et al.* (2018) *Genomic atlas of the human plasma proteome*. *Nature* 558 73–79 (2018). <https://doi.org/10.1038/s41586-018-0175-2>
- <sup>72</sup> Emilsson, Valur, *et al* (2018) *Co-Regulatory Networks of Human Serum Proteins Link Genetics to Disease*. *Science*, vol. 361, no. 6404, Aug. 2018, pp. 769–773. doi:10.1126/science.aaq1327
- <sup>73</sup> Joshi A and Mayr M. (2018) *In Aptamers They Trust: The Caveats of the SOMAscan Biomarker Discovery Platform from SomaLogic*. *Circulation*. 2018 November 27; 138(22):2482-2485. doi: 10.1161/CIRCULATIONAHA.118.036823. Epub 2018 November 26.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6277005/pdf/emss-80116.pdf>
- <sup>74</sup> Zhang Y, Lai BS, Juhas M. *Recent Advances in Aptamer Discovery and Applications*. *Molecules*. 2019 March 7; 24(5):941. doi: 10.3390/molecules24050941  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6429292/pdf/molecules-24-00941.pdf>
- <sup>75</sup> Song, Kyung-Mi; Lee, Seonghwan; Ban, Changill. (2012) *Aptamers and Their Biological Applications*. *Sensors* 12, no. 1: 612-631. <https://www.mdpi.com/1424-8220/12/1/612/pdf>
- <sup>76</sup> Wu, Yao, *et al.* “Perspective on the Future Role of Aptamers in Analytical Chemistry.” *Analytical Chemistry*, vol. 91, no. 24, December 2019, pp. 15335–15344. doi:10.1021/acs.analchem.9b03853
- <sup>77</sup> De La Fuente, Adriana, *et al.* “Aptamers against Mouse and Human Tumor-Infiltrating Myeloid Cells as Reagents for Targeted Chemotherapy.” *Science Translational Medicine*, vol. 12, no. 548, June 2020, p. eaav9760. doi:10.1126/scitranslmed.aav9760.
- <sup>78</sup> Schmitz, F.R.W., Valério, A., de Oliveira, D. *et al.* An overview and future prospects on aptamers for food safety. *Appl Micro Biotech* 104, 6929–6939 (2020) <https://doi.org/10.1007/s00253-020-10747-0>  
<https://link.springer.com/content/pdf/10.1007/s00253-020-10747-0.pdf>
- <sup>79</sup> Sumedha D Jayasena, *Aptamers: An Emerging Class of Molecules That Rival Antibodies in Diagnostics*, *Clinical Chemistry*, Volume 45, Issue 9, 1 September 1999, Pages 1628-1650.  
<https://doi.org/10.1093/clinchem/45.9.1628>
- <sup>80</sup> Hicke BJ, Stephens AW, Gould T, *et al.* (2006) Tumor targeting by an aptamer. *J Nucl Med*. 2006; 47(4):668-678. <http://jnm.snmjournals.org/content/47/4/668.full.pdf+html>
- <sup>81</sup> Trausch, Jeremiah J., *et al.* “Replacing Antibodies with Modified DNA Aptamers in Vaccine Potency Assays.” *Vaccine*, vol. 35, no. 41, October 2017, pp. 5495–502. doi:10.1016/j.vaccine.2017.04.003
- <sup>82</sup> Javier DJ, Nitin N, Levy M, Ellington A, Richards-Kortum R. *Aptamer-targeted gold nanoparticles as molecular-specific contrast agents for reflectance imaging*. *Bioconjug Chem*. 2008 June; 19(6):1309-12. doi: 10.1021/bc8001248. Epub 2008 May 31.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2651625/pdf/bc8001248.pdf>
- <sup>83</sup> Hianik T, Porfireva A, Grman I, Evtugyn G. EQCM biosensors based on DNA aptamers and antibodies for rapid detection of prions. *Protein Pept Lett*. 2009;16(4):363-367. doi:10.2174/092986609787848090

- <sup>84</sup> Lee, J., So, H., Jeon, E. *et al.* Aptamers as molecular recognition elements for electrical nanobiosensors. *Anal Bioanal Chem* **390**, 1023–1032 (2008). <https://doi.org/10.1007/s00216-007-1643-y>  
<https://link.springer.com/content/pdf/10.1007/s00216-007-1643-y.pdf>
- <sup>85</sup> Arshavsky-Graham, Sofia, et al. “Aptamers vs. Antibodies as Capture Probes in Optical Porous Silicon Biosensors.” *The Analyst*, vol. 145, no. 14, 2020, pp. 4991–5003. doi:10.1039/D0AN00178C
- <sup>86</sup> Zou X, Wu J, Gu J, Shen L, Mao L. Application of Aptamers in Virus Detection and Antiviral Therapy. *Front Microbiol.* 2019 July 3;10:1462. doi: 10.3389/fmicb.2019.01462.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6618307/pdf/fmicb-10-01462.pdf>
- <sup>87</sup> Ahn DG, Jeon IJ, Kim JD, et al. RNA aptamer-based sensitive detection of SARS coronavirus nucleocapsid protein. *Analyst.* 2009;134(9):1896-1901. doi:10.1039/b906788d  
<https://pubs.rsc.org/en/content/articlepdf/2009/an/b906788d>
- <sup>88</sup> Wyllie, Anne L., et al. Saliva or Nasopharyngeal Swab Specimens for Detection of SARS-CoV-2. *New England J of Medicine*, Aug 2020, p. NEJMc2016359 doi:10.1056/NEJMc2016359  
<https://www.nejm.org/doi/pdf/10.1056/NEJMc2016359>  
<https://science.sciencemag.org/content/369/6507/1041/tab-pdf>
- <sup>89</sup> Shubham, Shambhavi, "Selection of functional RNA aptamers against Ebola glycoproteins" (2017). Graduate Theses and Dissertations. Iowa State University. <https://lib.dr.iastate.edu/etd/16528>
- <sup>90</sup> Eichel, H. J., et al. “Acid and Alkaline Ribonucleases of Human Parotid, Submaxillary, and Whole Saliva.” *Arch of Biochemistry and Biophysics*, vol. 107, no. 2, August. 1964, pp. 197–208  
doi:10.1016/0003-9861(64)90322-4
- <sup>91</sup> Cho SJ, Woo HM, Kim KS, Oh JW, Jeong YJ. Novel system for detecting SARS coronavirus nucleocapsid protein using an ssDNA aptamer. *J Biosci Bioeng.* 2011 Dec; 112(6):535-40. doi: 10.1016/j.jbiosc.2011.08.014. [www.ncbi.nlm.nih.gov/pmc/articles/PMC7106535/pdf/main.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC7106535/pdf/main.pdf)
- <sup>92</sup> Chen Z, Wu Q, Chen J, Ni X, Dai J. DNA Aptamer Based Method for Detection of SARS-CoV-2 Nucleocapsid Protein. *Virol Sin.* 2020 June; 35(3):351-354. doi: 10.1007/s12250-020-00236-z. Epub 2020 May 25. [www.ncbi.nlm.nih.gov/pmc/articles/PMC7246297/pdf/12250\\_2020\\_Article\\_236.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC7246297/pdf/12250_2020_Article_236.pdf)
- <sup>93</sup> Nucleocapsid Phosphoprotein: Severe Acute Respiratory Syndrome Coronav - Protein - NCBI. [https://www.ncbi.nlm.nih.gov/protein/YP\\_009724397.2](https://www.ncbi.nlm.nih.gov/protein/YP_009724397.2)
- <sup>94</sup> Nucleocapsid Protein: Severe Acute Respiratory Syndrome-Related Corona - Protein - NCBI. [https://www.ncbi.nlm.nih.gov/protein/NP\\_828858.1](https://www.ncbi.nlm.nih.gov/protein/NP_828858.1)
- <sup>95</sup> Zhang, Liyun, et al. “Discovery of Sandwich Type COVID-19 Nucleocapsid Protein DNA Aptamers.” *Chem Communications*, vol. 56, no. 70, 2020, pp. 10235–10238. doi:10.1039/D0CC03993D. <https://pubs-rsc-org.libproxy.mit.edu/en/content/articlepdf/2020/cc/d0cc03993d>
- <sup>96</sup> Li, Hui-Yan, et al. Advances in Detection of Infectious Agents by Aptamer-Based Technologies. *Emerging Microbes & Infections*, vol. 9, no. 1, January 2020, pp. 1671–1681.  
doi:10.1080/22221751.2020.1792352.  
<https://www.tandfonline.com/doi/pdf/10.1080/22221751.2020.1792352?needAccess=true>
- <sup>97</sup> Lucia Wang and Maureen McKeague (2020) Aptamers in the pursuit of COVID-19 management. Aptamers (2020), Vol 4, in press. <http://japtamers.co.uk/wp-content/uploads/2020/07/Wang.pdf>

- <sup>98</sup> <https://www.aptamergroup.co.uk/aptamers-in-the-battle-against-covid19/>
- <sup>99</sup> Cleri, Fabrizio, Lensink, Marc F., Blossey, Ralf (2020): *DNA Aptamers Block the Receptor Binding Domain at the Spike Protein of SARS-CoV-2*. ChemRxiv. Preprint.  
<https://doi.org/10.26434/chemrxiv.12696173.v1> <https://chemrxiv.org/ndownloader/files/24042815>
- <sup>100</sup> NSF Award Abstract #2028531(RAPID) Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2)-Prevention: Multiple-Site Binding with Fusing Aptamers to mitigate Coronavirus Disease 2019.  
[https://www.nsf.gov/awardsearch/showAward?AWD\\_ID=2028531&HistoricalAwards=false](https://www.nsf.gov/awardsearch/showAward?AWD_ID=2028531&HistoricalAwards=false)
- <sup>101</sup> Song, Yanling, et al. “Discovery of Aptamers Targeting the Receptor-Binding Domain of the SARS-CoV-2 Spike Glycoprotein.” *Analytical Chemistry*, vol. 92, no. 14, July 2020, pp. 9895–900.  
doi:10.1021/acs.analchem.0c01394. <https://pubs.acs.org/doi/pdf/10.1021/acs.analchem.0c01394>
- <sup>102</sup> Bojkova, D., Klann, K., Koch, B. *et al.* Proteomics of SARS-CoV-2-infected host cells reveals therapy targets. *Nature* 583, 469–472 (2020). <https://doi.org/10.1038/s41586-020-2332-7>  
<https://www.nature.com/articles/s41586-020-2332-7.pdf>
- <sup>103</sup> Wu, Canrong, et al. “Analysis of Therapeutic Targets for SARS-CoV-2 and Discovery of Potential Drugs by Computational Methods.” *Acta Pharmaceutica Sinica B*, vol. 10, no. 5, May 2020, pp. 766–788.  
doi:10.1016/j.apsb.2020.02.008. <https://www.sciencedirect.com/science/article/pii/S2211383520302999>
- <sup>104</sup> Thoms, Matthias, et al. “Structural Basis for Translational Shutdown and Immune Evasion by the Nsp1 Protein of SARS-CoV-2.” *Science*, vol. 369, no. 6508, September 2020, pp. 1249–55.  
doi:10.1126/science.abc8665. <https://science.sciencemag.org/content/369/6508/1249.full.pdf>
- <sup>105</sup> Rong, Y., et al. “Post Hoc Support Vector Machine Learning for Impedimetric Biosensors Based on Weak Protein–Ligand Interactions.” *The Analyst*, vol. 143, no. 9, 2018, pp. 2066–2075.  
doi:10.1039/C8AN00065D <https://pubs.rsc.org/ko/content/getauthorversionpdf/C8AN00065D>
- <sup>106</sup> Choi, Jong Hyun, et al. “Aptamer-Capped Nanocrystal Quantum Dots: A New Method for Label-Free Protein Detection.” *J of the American Chemical Society*, vol. 128, no. 49, December 2006, pp. 15584–85.  
doi:10.1021/ja066506k.
- <sup>107</sup> Smith, Andrew M., et al. “Quantum Dot Nanocrystals for In Vivo Molecular and Cellular Imaging.” *Photochemistry & Photobiology*, vol. 80, no. 3, 2004, p. 377. doi:10.1562/2004-06-21-IR-209.1
- <sup>108</sup> Bagalkot, Vaishali, et al. “Quantum Dot–Aptamer Conjugates for Synchronous Cancer Imaging, Therapy, and Sensing of Drug Delivery Based on Bi-Fluorescence Resonance Energy Transfer.” *Nano Letters*, vol. 7, no. 10, October 2007, pp. 3065–3070. doi:10.1021/nl071546n.
- <sup>109</sup> Li, Zhiming, et al. “Aptamer-Conjugated Dendrimer-Modified Quantum Dots for Glioblastoma Cells Imaging.” *J of Physics: Conference Series*, vol. 188, September 2009, p. 012032.  
doi:10.1088/1742-6596/188/1/012032.
- <sup>110</sup> Roberts, Kenneth and Bumm Lloyd, A. (2010) *Quantum Dot Nano-crystals Coupled to DNA Aptamers: Sensors for Biological Weapons Detection* <https://apps.dtic.mil/dtic/tr/fulltext/u2/a567156.pdf>
- <sup>111</sup> Zhou, DJ (2012) *Quantum dot-nucleic acid/aptamer bioconjugate based fluorimetric biosensors*. Biochemical Society Transactions 40 635 – 639 ISSN 0300-5127 <https://doi.org/10.1042/BST20120059>
- <sup>112</sup> <https://www.microsoft.com/en-us/hololens/hardware>
- <sup>113</sup> <https://www.denso-wave.com/en/technology/vol1.html>

---

<sup>114</sup> <https://en.wikipedia.org/wiki/Kinect>

<sup>115</sup> Carlo Tomasi and Takeo Kanade (1992) *Shape and Motion from Image Streams under Orthography: Factorization Method*. International Journal of Computer Vision 9:2, 137-154

[https://people.eecs.berkeley.edu/~yang/courses/cs294-6/papers/TomasiC\\_Shape%20and%20motion%20from%20image%20streams%20under%20orthography.pdf](https://people.eecs.berkeley.edu/~yang/courses/cs294-6/papers/TomasiC_Shape%20and%20motion%20from%20image%20streams%20under%20orthography.pdf)

<sup>116</sup> <https://www.microsoft.com/en-us/research/blog/in-search-for-future-of-cloud-storage-researchers-look-to-holographic-storage-solutions/>

<sup>117</sup> <https://emclamor.wixsite.com/mclamorelab>

<sup>118</sup> Philip Levis, Sam Madden, David Gay, Joseph Polastre, Robert Szewczyk, Alec Woo, Eric Brewer and David Culler (2003) *The Emergence of Networking Abstractions and Techniques in TinyOS*

[www.usenix.org/legacy/events/nsdi04/tech/full\\_papers/levisEmerge/levisEmerge\\_html/tinyos-nsdi03.html](http://www.usenix.org/legacy/events/nsdi04/tech/full_papers/levisEmerge/levisEmerge_html/tinyos-nsdi03.html)

<sup>119</sup> McLamore, E.S., S.P.A. Datta, V. Morgan, N. Cavallaro, G. Kiker, D.M. Jenkins, Y. Rong, C. Gomes, J. Claussen, D. Vanegas, E.C. Alocilja (2019) *SNAPS: Sensor Analytics Point Solutions for Detection and Decision Support*. Sensors, vol. 19, no. 22, November 2019, p. 4935

[www.mdpi.com/1424-8220/19/22/4935/pdf](http://www.mdpi.com/1424-8220/19/22/4935/pdf) and <https://dspace.mit.edu/handle/1721.1/123983>

<sup>120</sup> Walker, Bruce (2020) MIT Biology course number 7.00 (COVID-19, SARS-CoV-2 and the Pandemic)

<https://biology.mit.edu/undergraduate/current-students/subject-offerings/covid-19-sars-cov-2-and-the-pandemic/> and [https://biology.mit.edu/wp-content/uploads/2020/09/7.00\\_Syllabus\\_9.8.20.pdf](https://biology.mit.edu/wp-content/uploads/2020/09/7.00_Syllabus_9.8.20.pdf)

<sup>121</sup> Jack-in-the-Box <https://en.wikipedia.org/wiki/Jack-in-the-box>

<sup>122</sup> Walls, Alexandra C., et al. "Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein." *Cell*, vol. 181, no. 2, April 2020, pp. 281-292.e6. doi:10.1016/j.cell.2020.02.058

[https://www.cell.com/cell/pdf/S0092-8674\(20\)30262-2.pdf](https://www.cell.com/cell/pdf/S0092-8674(20)30262-2.pdf)

<sup>123</sup> Ellen Shrock, Eric Fujimura, Tomasz Kula, Richard T. Timms, I-Hsiu Lee, Yumei Leng, Matthew L.

Robinson, Brandon M. Sie, Mamie Z. Li, Yuezhou Chen, Jennifer Logue, Adam Zuiani, Denise

Mcculloch, Felipe J. N. Lelis, Stephanie Henson, Daniel E. Monaco, Meghan Travers, Shaghayegh

Habibi, William A. Clarke, Patrizio Caturegli, Oliver Laeyendecker, Alicja Piechocka-Trocha, Jon

Li, Ashok Khatri, Helen Y. Chu, MGH COVID-19 Collection & Processing Team, Alexandra-Chloé

Villani, Kyle Kays, Marcia B. Goldberg, Nir Hacohen, Michael R. Filbin, Xu G. Yu, Bruce D.

Walker, Duane R. Wesemann, H. Benjamin Larman, James A. Lederer, and Stephen J. Elledge (2020)

*Viral epitope profiling of covid-19 patients reveals cross-reactivity and correlates of severity*. Published online 29 sep 2020. Science doi: 10.1126/science.abd4250

<https://science.sciencemag.org/content/early/2020/09/28/science.abd4250/tab-pdf>

<sup>124</sup> McKee DL, Sternberg A, Stange U, Laufer S, Naujokat C. *Candidate drugs against SARS-CoV-2 and*

*COVID-19*. Pharmacol Res. 2020 July; 157:104859. doi: 10.1016/j.phrs.2020.104859. Epub 2020 April 29.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7189851/pdf/main.pdf>

- 
- <sup>125</sup> Mark Fischetti, Veronica Falconieri Hays, Britt Glaunsinger and Jen Christiansen (2020) *Inside the Coronavirus*. Scientific American 323, 1, 32-37 (July 1, 2020) doi:[10.1038/scientificamerican0720-32](https://doi.org/10.1038/scientificamerican0720-32)  
*A Visual Guide to the SARS-CoV-2 Coronavirus: What scientists know about the inner workings of the pathogen that has infected the world*. July 1, 2020. Scientific American.  
<https://www.scientificamerican.com/article/a-visual-guide-to-the-sars-cov-2-coronavirus/> &  
<https://www.scientificamerican.com/interactive/inside-the-coronavirus/>
- <sup>126</sup> Glaunsinger, Britt (2020) MIT Biology course 7.00 (COVID-19, SARS-CoV-2 and the Pandemic)  
<https://biology.mit.edu/undergraduate/current-students/subject-offerings/covid-19-sars-cov-2-and-the-pandemic/> and [https://biology.mit.edu/wp-content/uploads/2020/09/7.00\\_Syllabus\\_9.8.20.pdf](https://biology.mit.edu/wp-content/uploads/2020/09/7.00_Syllabus_9.8.20.pdf)
- <sup>127</sup> Jia HP, Look DC, Shi L, Hickey M, Pewe L, Netland J, Farzan M, Wohlford-Lenane C, Perlman S, McCray PB Jr. *ACE2 receptor expression and severe acute respiratory syndrome coronavirus infection depend on differentiation of human airway epithelia*. J Virol. 2005 December; 79(23):14614-14621. doi: 10.1128/JVI.79.23.14614-14621.2005  
[www.ncbi.nlm.nih.gov/pmc/articles/PMC1287568/pdf/1240-05.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1287568/pdf/1240-05.pdf)
- <sup>128</sup> Saheb Sharif-Askari, Narjes, et al. “Airways Expression of SARS-CoV-2 Receptor, ACE2, and TMPRSS2 Is Lower in Children Than Adults and Increases with Smoking and COPD.” *Molecular Therapy - Methods & Clin Development*, vol. 18, Sept. 2020, pp. 1–6. doi:10.1016/j.omtm.2020.05.013  
[https://www.cell.com/molecular-therapy-family/methods/pdf/S2329-0501\(20\)30100-5.pdf](https://www.cell.com/molecular-therapy-family/methods/pdf/S2329-0501(20)30100-5.pdf)
- <sup>129</sup> Hilgenfeld R. From SARS to MERS: crystallographic studies on coronaviral proteases enable antiviral drug design. *FEBS J*. 2014; 281(18):4085-4096. doi:10.1111/febs.12936  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7163996/pdf/FEBS-281-4085.pdf>
- <sup>130</sup> Goodsell, D. S. “Coronavirus Proteases.” *RCSB Protein Data Bank*, February 2020. doi:10.2210/rcsb\_pdb/mom\_2020\_2 <https://pdb101.rcsb.org/motm/242>
- <sup>131</sup> Barretto, Naina, et al. “The Papain-Like Protease of Severe Acute Respiratory Syndrome Coronavirus Has Deubiquitinating Activity.” *Journal of Virology*, vol. 79, no. 24, December 2005, pp. 15189–15198. doi:10.1128/JVI.79.24.15189-15198.2005 <https://jvi.asm.org/content/jvi/79/24/15189.full.pdf>
- <sup>132</sup> Boras, Britton, et al. *Discovery of a Novel Inhibitor of Coronavirus 3CL Protease as a Clinical Candidate for the Potential Treatment of COVID-19*. preprint, Pharmacology and Toxicology, 13 September 2020. doi:10.1101/2020.09.12.293498  
<https://www.biorxiv.org/content/10.1101/2020.09.12.293498v2.full.pdf>
- <sup>133</sup> Xue, Xiaoyu, et al. “Structures of Two Coronavirus Main Proteases: Implications for Substrate Binding and Antiviral Drug Design.” *Journal of Virology*, vol. 82, no. 5, March 2008, pp. 2515–2527. doi:10.1128/JVI.02114-07 <https://jvi.asm.org/content/jvi/82/5/2515.full.pdf>
- <sup>134</sup> Tahir ul Qamar, Muhammad, et al. “Structural Basis of SARS-CoV-2 3CLpro and Anti-COVID-19 Drug Discovery from Medicinal Plants.” *Journal of Pharmaceutical Analysis*, vol. 10, no. 4, Aug. 2020, pp. 313–319. doi:10.1016/j.jpha.2020.03.009  
<https://www.sciencedirect.com/science/article/pii/S2095177920301271>



- <sup>135</sup> Rathnayake, Athri D., et al. “3C-like Protease Inhibitors Block Coronavirus Replication in Vitro and Improve Survival in MERS-CoV Infected Mice.” *Sci Trans Med*, vol. 12, no. 557, Aug. 2020, p. eabc5332. doi:10.1126/scitranslmed.abc5332 <https://stm.sciencemag.org/content/12/557/eabc5332/tab-pdf>
- <sup>136</sup> Zhang, Linlin, et al. “Crystal Structure of SARS-CoV-2 Main Protease Provides a Basis for Design of Improved  $\alpha$ -Ketoamide Inhibitors.” *Science*, March 2020, p. eabb3405. doi:10.1126/science.abb3405 <https://science.sciencemag.org/content/368/6489/409/tab-pdf>
- <sup>137</sup> Báez-Santos YM, Barraza SJ, Wilson MW, Agius MP, Mielech AM, Davis NM, Baker SC, Larsen SD, Mesecar AD. (2014) *X-ray structural and biological evaluation of a series of potent and highly selective inhibitors of human coronavirus papain-like proteases*. *J Med Chem*. 2014 March 27; 57(6):2393-412. doi: 10.1021/jm401712t. Epub 2014 March 14. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3983375/pdf/jm401712t.pdf>
- <sup>138</sup> Rut, Wioletta, et al. *Activity Profiling and Structures of Inhibitor-Bound SARS-CoV-2-PLpro Protease Provides a Framework for Anti-COVID-19 Drug Design*. preprint, *Biochemistry*, 29 April 2020. doi:10.1101/2020.04.29.068890 <https://www.biorxiv.org/content/biorxiv/early/2020/04/29/2020.04.29.068890.full.pdf>
- <sup>139</sup> Shin, D., Mukherjee, R., Grewe, D. et al. Papain-like protease regulates SARS-CoV-2 viral spread and innate immunity. *Nature* (2020). <https://doi.org/10.1038/s41586-020-2601-5> [https://www.nature.com/articles/s41586-020-2601-5\\_reference.pdf](https://www.nature.com/articles/s41586-020-2601-5_reference.pdf)
- <sup>140</sup> Thoms, Matthias, et al. “Structural Basis for Translational Shutdown and Immune Evasion by the Nsp1 Protein of SARS-CoV-2.” *Science*, vol. 369, no. 6508, September 2020, pp. 1249–55. doi:10.1126/science.abc8665 <https://science.sciencemag.org/content/369/6508/1249.full.pdf>
- <sup>141</sup> Kamitani W, Huang C, Narayanan K, Lokugamage KG, Makino S. (2009) *A two-pronged strategy to suppress host protein synthesis by SARS coronavirus Nsp1 protein*. *Nat Struct Mol Biol*. 2009 November; 16(11):1134-40. doi: 10.1038/nsmb.1680. Epub 2009 October 18. [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2784181/pdf/41594\\_2009\\_Article\\_BFnsmb1680.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2784181/pdf/41594_2009_Article_BFnsmb1680.pdf)
- <sup>142</sup> Lapointe, Christopher P., et al. *Dynamic Competition between SARS-CoV-2 NSP1 and MRNA on the Human Ribosome Inhibits Translation Initiation*. preprint, *Biophysics*, 21 August 2020. doi:10.1101/2020.08.20.259770 <https://www.biorxiv.org/content/10.1101/2020.08.20.259770v1.full.pdf>
- <sup>143</sup> Schubert, K., Karousis, E.D., Jomaa, A. et al. SARS-CoV-2 Nsp1 binds the ribosomal mRNA channel to inhibit translation. *Nat Struct Mol Biol* (2020). <https://doi.org/10.1038/s41594-020-0511-8> <https://www.nature.com/articles/s41594-020-0511-8.pdf>
- <sup>144</sup> Lee, J.S., Shin, E. The type I interferon response in COVID-19: implications for treatment. *Nature Review of Immunology* 20, 585–586 (2020). <https://doi.org/10.1038/s41577-020-00429-3> <https://www.nature.com/articles/s41577-020-00429-3.pdf>
- <sup>145</sup> Zhang, Qian, et al. “Inborn Errors of Type I IFN Immunity in Patients with Life-Threatening COVID-19.” *Science*, September 24, 2020, p. eabd4570. doi:10.1126/science.abd4570 <https://science.sciencemag.org/content/early/2020/09/23/science.abd4570/tab-pdf>

- <sup>146</sup> Hadjadj, Jérôme, et al. “Impaired Type I Interferon Activity and Inflammatory Responses in Severe COVID-19 Patients.” *Science*, vol. 369, no. 6504, August 7, 2020, pp. 718–724.  
doi:10.1126/science.abc6027 <https://science.sciencemag.org/content/369/6504/718/tab-pdf>
- <sup>147</sup> Xia, H., Cao, Z., Xie, X., Zhang, X., Yun-Chung Chen, J., Wang, H., Menachery, V.D., Rajsbaum, R., Shi, P.-Y. (2020) *Evasion of type-I interferon by SARS-CoV-2*. Cell Reports (2020).  
<https://doi.org/10.1016/j.celrep.2020.108234>  
<https://www.cell.com/action/showPdf?pii=S2211-1247%2820%2931223-7>
- <sup>148</sup> Ella Hartenian, Divya Nandakumar, Azra Lari, Michael Ly, Jessica M Tucker and Britt A Glaunsinger. “The Molecular Virology of Coronaviruses.” *J of Biol Chem*, vol. 295, no. 37, September 2020, pp. 12910–12934. <https://www.jbc.org/cgi/doi/10.1074/jbc.REV120.013930>  
<https://www.jbc.org/content/early/2020/07/13/jbc.REV120.013930.full.pdf>
- <sup>149</sup> Eckerle LD, Becker MM, Halpin RA, Li K, Venter E, Lu X, Scherbakova S, Graham RL, Baric RS, Stockwell TB, Spiro DJ, Denison MR. (2010) *Infidelity of SARS-CoV Nsp14-exonuclease mutant virus replication is revealed by complete genome sequencing*. PLoS Pathog. 2010 May 6; 6(5):e1000896. doi: 10.1371/journal.ppat.1000896 [www.ncbi.nlm.nih.gov/pmc/articles/PMC2865531/pdf/ppat.1000896.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2865531/pdf/ppat.1000896.pdf)
- <sup>150</sup> Streisinger G, Okada Y, Emrich J, et al. (1966) Frameshift mutations and the genetic code. This paper is dedicated to Professor Theodosius Dobzhansky on the occasion of his 66th birthday. *Cold Spring Harb Symp Quant Biol*. 1966; 31:77-84. doi:10.1101/sqb.1966.031.01.014  
<http://symposium.cshlp.org/content/31/77.long>
- <sup>151</sup> Reddy, E. P., et al. A point mutation is responsible for the acquisition of transforming properties by the T24 human bladder carcinoma oncogene. *Nature* 300, 149–152 (1982)
- <sup>152</sup> Ou X, Zheng W, Shan Y, Mu Z, Dominguez SR, Holmes KV, Qian Z. (2016) *Identification of the Fusion Peptide-Containing Region in Betacoronavirus Spike Glycoproteins*. J Virol. 2016 May 27; 90(12):5586-5600. doi: 10.1128/JVI.00015-16  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4886789/pdf/zjv5586.pdf>
- <sup>153</sup> Xu, Yanhui, et al. “Crystal Structure of Severe Acute Respiratory Syndrome Coronavirus Spike Protein Fusion Core.” *Journal of Biological Chemistry*, vol. 279, no. 47, November 2004, pp. 49414-419. doi:10.1074/jbc.M408782200 <https://www.jbc.org/content/279/47/49414.full.pdf>
- <sup>154</sup> Madu IG, Roth SL, Belouzard S, Whittaker GR. (2009) *Characterization of a highly conserved domain within the severe acute respiratory syndrome coronavirus spike protein S2 domain with characteristics of a viral fusion peptide*. J Virol. 2009 August; 83(15):7411-21. doi: 10.1128/JVI.00079-09. Epub 2009 May 13  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2708636/pdf/0079-09.pdf>
- <sup>155</sup> Tang T, Bidon M, Jaimes JA, Whittaker GR, Daniel S. (2020) *Coronavirus membrane fusion mechanism offers a potential target for antiviral development*. Antiviral Res. 2020 June; 178:104792. doi: 10.1016/j.antiviral.2020.104792. Epub 2020 April 6.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7194977/pdf/main.pdf>
- <sup>156</sup> Pattnaik GP, Chakraborty H. *Entry Inhibitors: Efficient Means to Block Viral Infection*. J Membrane Biology. 2020 August 30: 1–20. doi: 10.1007/s00232-020-00136-z  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7456447/pdf/232\\_2020\\_Article\\_136.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7456447/pdf/232_2020_Article_136.pdf)

- <sup>157</sup> Bosch BJ, Martina BE, Van Der Zee R, Lepault J, Haijema BJ, Versluis C, Heck AJ, De Groot R, Osterhaus AD, Rottier PJ. (2004) *Severe acute respiratory syndrome coronavirus (SARS-CoV) infection inhibition using spike protein heptad repeat-derived peptides*. Proc Natl Acad Sci U S A. 2004 June 1; 101(22):8455-60. doi: 10.1073/pnas.0400576101. Epub 2004 May 18.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC420415/pdf/1018455.pdf>
- <sup>158</sup> Leila Kashefi-Kheyraadi, Junmoo Kim, Sudesna Chakravarty, Sunyoung Park, Hogyong Gwak, Seung-Il Kim, Mohsen Mohammadniaei, Min-Ho Lee, Kyung-A Hyun and Hyo-Il Jung (2020) Detachable microfluidic device implemented with electrochemical aptasensor (DeMEA) for sequential analysis of cancerous exosomes. *Biosensors & Bioelectronics* ( IF 10.257 ) Published September 17, 2020. DOI: 10.1016/j.bios.2020.112622
- <sup>159</sup> Contreras-Naranjo JC, Wu HJ, Ugaz VM. (2017) *Microfluidics for exosome isolation and analysis: enabling liquid biopsy for personalized medicine*. Lab Chip. 2017 October 25; 17(21):3558-3577. doi: 10.1039/c7lc00592j [www.ncbi.nlm.nih.gov/pmc/articles/PMC5656537/pdf/nihms903616.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5656537/pdf/nihms903616.pdf)
- <sup>160</sup> Economics of Technology (Collected Papers) <http://bit.ly/Economics-of-Technology>
- <sup>161</sup> Brat, Gabriel A., et al. "International Electronic Health Record-Derived COVID-19 Clinical Course Profiles: The 4CE Consortium." *Npj Digital Medicine*, vol. 3, no. 1, December. 2020, p. 109. doi:10.1038/s41746-020-00308-0 <https://www.nature.com/articles/s41746-020-00308-0.pdf>
- <sup>162</sup> Tromberg BJ, Schwetz TA, Pérez-Stable EJ, et al. Rapid Scaling Up of Covid-19 Diagnostic Testing in the United States - The NIH RADx Initiative. *N Engl J Med*. September 10, 2020; 383(11):1071-1077. doi:10.1056/NEJMSr2022263 [www.nejm.org/doi/pdf/10.1056/NEJMSr2022263?articleTools=true](http://www.nejm.org/doi/pdf/10.1056/NEJMSr2022263?articleTools=true)
- <sup>163</sup> Yan, Li-Meng, et al. *Unusual Features of the SARS-CoV-2 Genome Suggesting Sophisticated Laboratory Modification Rather Than Natural Evolution and Delineation of Its Probable Synthetic Route*. Sept. 2020. doi:10.5281/ZENODO.4028830  
[https://zenodo.org/record/4028830/files/The\\_Yan\\_Report.pdf?download=1](https://zenodo.org/record/4028830/files/The_Yan_Report.pdf?download=1)
- <sup>164</sup> Mariana Guadalupe Matus García (2018) *Analysis of fecal biomarkers to impact clinical care and public health*. Thesis: Ph. D., Massachusetts Institute of Technology, Computational and Systems Biology Program, 2018. MIT Library <https://dspace.mit.edu/handle/1721.1/119603>
- <sup>165</sup> Polo, David, et al. "Making Waves: Wastewater-Based Epidemiology for COVID-19 – Approaches and Challenges for Surveillance and Prediction." *Water Research*, vol. 186, November 2020, p. 116404. doi:10.1016/j.watres.2020.116404 [www.ncbi.nlm.nih.gov/pmc/articles/PMC7480445/pdf/main.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC7480445/pdf/main.pdf)
- <sup>166</sup> Duvallat C, Hayes BD, Erickson TB, Chai PR, Matus M. (2020) *Mapping Community Opioid Exposure Through Wastewater-Based Epidemiology as a Means to Engage Pharmacies in Harm Reduction Efforts*. Prevention of Chronic Diseases 2020; 17:200053. DOI: <http://dx.doi.org/10.5888/pcd17.200053>
- <sup>167</sup> Leonard, Maureen, et al. (2020) "Multi-Omics Analysis Reveals the Influence of Genetic and Environmental Risk Factors on Developing Gut Microbiota in Infants at Risk of Celiac Disease." *Microbiome*, vol. 8, no. 1, December 2020, p. 130. doi:10.1186/s40168-020-00906-w
- <sup>168</sup> Lee, J., Hyeon, D.Y. and Hwang, D. (2020) Single-cell multiomics: technologies and data analysis methods. *Exp Mol Med* (2020). <https://doi.org/10.1038/s12276-020-0420-2> & [www.nature.com/articles/s12276-020-0420-2.pdf](http://www.nature.com/articles/s12276-020-0420-2.pdf)

- 
- <sup>169</sup> Foy BH, Carlson JCT, Reinertsen E, et al. Association of Red Blood Cell Distribution Width (RDW) With Mortality Risk in Hospitalized Adults With SARS-CoV-2 Infection. *JAMA Network Open* 2020; 3(9):e2022058. doi:10.1001/jamanetworkopen.2020.22058  
<https://jamanetwork.com/journals/jamanetworkopen/fullarticle/2770945>
- <sup>170</sup> Mackuľak, Tomáš, et al. “Music Festivals and Drugs: Wastewater Analysis.” *Science of The Total Environment*, vol. 659, April 2019, pp. 326–334. doi:10.1016/j.scitotenv.2018.12.275
- <sup>171</sup> Castiglioni, S. and Vandam, L. (2016) *A global overview of wastewater-based epidemiology*. pp. 45–54 in “Assessing illicit drugs in wastewater: advances in wastewater-based drug epidemiology” EMCDDA Insights 22 (EU). [www.emcdda.europa.eu/system/files/publications/2273/TDXD16022ENC\\_4.pdf](http://www.emcdda.europa.eu/system/files/publications/2273/TDXD16022ENC_4.pdf)
- <sup>172</sup> EMCDDA (European Monitoring Centre for Drugs and Drug Addiction) (2020) *PERSPECTIVES ON DRUGS: Wastewater analysis and drugs: results from a European multi-city study*. EU Publications Office [www.emcdda.europa.eu/system/files/publications/2757/POD\\_Wastewater%20analysis\\_update2020.pdf](http://www.emcdda.europa.eu/system/files/publications/2757/POD_Wastewater%20analysis_update2020.pdf)
- <sup>173</sup> Gina Pocock, Leanne Coetzee, Janet Mans, Maureen Taylor and Bettina Genthe (2020) *PROOF OF CONCEPT STUDY: Application of wastewater-based surveillance to monitor SARS-CoV-2 prevalence in South African communities*. Report to the Water Research Commission WRC Report no. TT 832/20 ISBN 978-0-6392-0187-0 (September 2020) Water Research Commission, South Africa [www.wrc.org.za http://wrcwebsite.azurewebsites.net/wp-content/uploads/mdocs/TT%20832-20%20final%20web.pdf](http://wrcwebsite.azurewebsites.net/wp-content/uploads/mdocs/TT%20832-20%20final%20web.pdf)
- <sup>174</sup> Delaney SK, Hultner ML, Jacob HJ, Ledbetter DH, McCarthy JJ, Ball M, Beckman KB, Belmont JW, Bloss CS, Christman MF, Cosgrove A, Damiani SA, Danis T, Delledonne M, Dougherty MJ, Dudley JT, Faucett WA, Friedman JR, Haase DH, Hays TS, Heilsberg S, Huber J, Kaminsky L, Ledbetter N, Lee WH, Levin E, Libiger O, Linderman M, Love RL, Magnus DC, Martland A, McClure SL, Megill SE, Messier H, Nussbaum RL, Palaniappan L, Patay BA, Popovich BW, Quackenbush J, Savant MJ, Su MM, Terry SF, Tucker S, Wong WT, Green RC. (2016) *Toward clinical genomics in everyday medicine: perspectives and recommendations*. *Expert Rev Mol Diagn.* 2016; 16(5):521-532. doi: 10.1586/14737159.2016.1146593. Epub 2016 February 24. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4841021/pdf/iero-16-521.pdf>
- <sup>175</sup> <http://www.opengroup.org/iot/odf/>
- <sup>176</sup> Perron, Paul. “Relire Le Fou d’Elsa de Louis Aragon.” *Études Littéraires*, vol. 28, no. 1, April 2005, pp. 69–82. doi:10.7202/501111ar.
- <sup>177</sup> Stephens DS, McElrath MJ. COVID-19 and the Path to Immunity. *JAMA*. Published online September 11, 2020. doi:10.1001/jama.2020.16656
- <sup>178</sup> Korber, B., et al. Spike Mutation Pipeline Reveals the Emergence of a More Transmissible Form of SARS-CoV-2. preprint, *Evolutionary Biology*, 30 April 2020. doi:10.1101/2020.04.29.069054  
<https://www.biorxiv.org/content/10.1101/2020.04.29.069054v2.full.pdf>

---

<sup>179</sup> Jerald Sadoff, Mathieu Le Gars, Georgi Shukarev, Dirk Heerwegh, Carla Truyers, Anne Marit de Groot, Jeroen Stoop, Sarah Tete, Wim Van Damme, Isabel Leroux-Roels, Pieter-Jan Berghmans, Murray Kimmel, Pierre Van Damme, Jan de Hoon, Williams Smith, Kathryn E. Stephenson, Dan H. Barouch, Stephen C. De Rosa, Kristen W. Cohen, M. Juliana McElrath, Emmanuel Cormier, Gert Scheper, Jenny Hendriks, Frank Struyf, Macaya Douoguih, Johan Van Hoof, and Hanneke Schuitemaker (2020) *Safety and Immunogenicity of the Ad26.COV2.S COVID-19 Vaccine Candidate: Interim Results of a Phase 1/2a, Double-Blind, Randomized, Placebo-Controlled Trial*. preprint, Infectious Diseases (except HIV/AIDS), 25 September 2020. doi:10.1101/2020.09.23.20199604

<https://www.medrxiv.org/content/10.1101/2020.09.23.20199604v1.full.pdf>

<sup>180</sup> Logunov, D. Y. et al (2020) *Safety and immunogenicity of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine in two formulations: two open, non-randomised phase 1/2 studies from Russia*. *Lancet* 2020; 396: 887–897. Published Online September 4, 2020.

<https://www.thelancet.com/action/showPdf?pii=S0140-6736%2820%2931866-3>

<sup>181</sup> Chadi M. Saad-Roy, Caroline E. Wagner, Rachel E. Baker, Sinead E. Morris, Jeremy Farrar, Andrea I. Graham, Simon A. Levin, Michael J. Mina, C. Jessica E. Metcalf, and Bryan T. Grenfell (2020) Immune life history, vaccination, and the dynamics of SARS-CoV-2 over the next 5 years. Published 21 Sep 2020 doi:10.1126/science.abd7343

<https://science.sciencemag.org/content/early/2020/09/18/science.abd7343>  
<https://science.sciencemag.org/content/early/2020/09/18/science.abd7343/tab-pdf>

<sup>182</sup> Neilan, A. M. et al (2020) Clinical Impact, Costs, and Cost-Effectiveness of Expanded SARS-CoV-2 Testing in Massachusetts. *Clinical Infectious Diseases* <https://doi.org/10.1093/cid/ciaa1418>

<sup>183</sup> The Global HIV/AIDS Epidemic

<https://www.kff.org/global-health-policy/fact-sheet/the-global-hiv-aids-epidemic/#footnote-475993-1>

<sup>184</sup> Corey L, Mascola JR, Fauci AS, Collins FS. A strategic approach to COVID-19 vaccine R&D. *Science*. 2020 May 29; 368(6494):948-950. doi: 10.1126/science.abc5312. Epub 2020 May 11.

<https://science.sciencemag.org/content/368/6494/948/tab-pdf>

<sup>185</sup> Anderson E. J. et al and the mRNA-1273 Study Group\* (2020) *Safety and immunogenicity of SARS-CoV-2 mRNA1273 vaccine in older adults*. Published 29 Sep 2020 NEJM DOI: 10.1056/NEJMoa2028436

<https://www.nejm.org/doi/pdf/10.1056/NEJMoa2028436?articleTools=true>

<sup>186</sup> McLellan JS, Chen M, Joyce MG, Sastry M, Stewart-Jones GB, Yang Y, Zhang B, Chen L, Srivatsan S, Zheng A, Zhou T, Graepel KW, Kumar A, Moin S, Boyington JC, Chuang GY, Soto C, Baxa U, Bakker AQ, Spits H, Beaumont T, Zheng Z, Xia N, Ko SY, Todd JP, Rao S, Graham BS, Kwong PD. *Structure-based design of a fusion glycoprotein vaccine for respiratory syncytial virus*. *Science*. 2013 November 1; 342(6158):592-8. doi: 10.1126/science.1243283. Erratum in: *Science*. 2013 November 22;342(6161):931

<sup>187</sup> Hsieh CL, Goldsmith JA, Schaub JM, DiVenere AM, Kuo HC, Javanmardi K, Le KC, Wrapp D, Lee AG, Liu Y, Chou CW, Byrne PO, Hjorth CK, Johnson NV, Ludes-Meyers J, Nguyen AW, Park J, Wang N, Amengor D, Maynard JA, Finkelstein IJ, McLellan JS. *Structure-based Design of Prefusion-stabilized SARS-CoV-2 Spikes*. bioRxiv. 2020 May 30:2020.05.30.125484. doi: 10.1101/2020.05.30.125484. Update: *Science*. 2020 July 23.



<sup>188</sup> Anderson EJ, Roupael NG, Widge AT, et al. Safety and immunogenicity of SARS-CoV-2 mRNA1273 vaccine in older adults. *N Engl J Med*. DOI: 10.1056/NEJMoa2028436

[https://www.nejm.org/doi/suppl/10.1056/NEJMoa2028436/suppl\\_file/nejmoa2028436\\_protocol.pdf](https://www.nejm.org/doi/suppl/10.1056/NEJMoa2028436/suppl_file/nejmoa2028436_protocol.pdf)

<sup>189</sup> Our World in Data

<https://ourworldindata.org/grapher/daily-covid-cases-7-day?time=2020-03-31..latest>

<sup>190</sup> Endalew AD, Faburay B, Wilson WC, Richt JA. *Schmallenberg Disease-A Newly Emerged Culicoides-borne Viral Disease of Ruminants*. *Viruses*. 2019 November 15; 11(11):1065. doi: 10.3390/v11111065

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6893508/pdf/viruses-11-01065.pdf>

<sup>191</sup> Collins ÁB, Doherty ML, Barrett DJ, Mee JF. *Schmallenberg virus: a systematic international literature review (2011-19) from an Irish perspective*. *Irish Vet J*. 2019 Oct 9; 72:9. doi: 10.1186/s13620-019-0147-3.

[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6785879/pdf/13620\\_2019\\_Article\\_147.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6785879/pdf/13620_2019_Article_147.pdf)

<sup>192</sup> Lechevalier H. Dmitri Iosifovich Ivanovski (1864-1920). *Bacteriol Rev*. 1972 June; 36(2):135-45.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC408320/pdf/bactrev00200-0002.pdf>

<sup>193</sup> Lustig, Alice and Levine, Arnold J. (1992) One hundred years of virology. *J Virol*. 1992 August; 66(8):4629-4631. doi: 10.1128/JVI.66.8.4629-4631.1992.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC241285/pdf/jvirol00040-0015.pdf>

<sup>194</sup> Fauci, Anthony (2020) MIT Biology course number 7.00 (COVID-19, SARS-CoV-2 and the Pandemic) [https://biology.mit.edu/wp-content/uploads/2020/09/7.00\\_Syllabus\\_9.8.20.pdf](https://biology.mit.edu/wp-content/uploads/2020/09/7.00_Syllabus_9.8.20.pdf) and

<https://biology.mit.edu/undergraduate/current-students/subject-offerings/covid-19-sars-cov-2-and-the-pandemic/>

<sup>195</sup> Baltimore, David (2020) MIT Biology course number 7.00 (COVID-19, SARS-CoV-2 and the Pandemic) <https://biology.mit.edu/undergraduate/current-students/subject-offerings/covid-19-sars-cov-2-and-the-pandemic/>

and [https://biology.mit.edu/wp-content/uploads/2020/09/7.00\\_Syllabus\\_9.8.20.pdf](https://biology.mit.edu/wp-content/uploads/2020/09/7.00_Syllabus_9.8.20.pdf)

<sup>196</sup> Baltimore, David. <https://www.nobelprize.org/prizes/medicine/1975/baltimore/facts/>

<sup>197</sup> Daly, James L., et al. “Neuropilin-1 Is a Host Factor for SARS-CoV-2 Infection.” *Science*, Oct 20, 2020. p. eabd3072. doi:10.1126/science.abd3072.

<sup>198</sup> Cantuti-Castelvetri, Ludovico, et al. “Neuropilin-1 Facilitates SARS-CoV-2 Cell Entry and Infectivity.” *Science*, October 20, 2020, p. eabd2985. doi:10.1126/science.abd2985.

<sup>199</sup> Tufekci, Zeynep. “This Overlooked Variable Is the Key to the Pandemic.” *The Atlantic*, 30 Sept. 2020. <https://www.theatlantic.com/health/archive/2020/09/k-overlooked-variable-driving-pandemic/616548/>

<sup>200</sup> Kai Kupferschmidt (2020) Why do some COVID-19 patients infect many others, whereas most don't spread the virus at all? *Science*. May 19, 2020. doi:10.1126/science.abc8931

<sup>201</sup> Thurner, Stefan, et al. *Why Are Most COVID-19 Infection Curves Linear?* preprint, *Epidemiology*, 24 May 2020. doi:10.1101/2020.05.22.20110403.

<https://www.medrxiv.org/content/10.1101/2020.05.22.20110403v1.full.pdf>

<sup>202</sup> Vilfredo Pareto, *Cours d'conomie Politique* Profess a l'Universit de Lausanne, Vol. I, 1896; Vol. II, 1897. <https://pdfs.semanticscholar.org/ef82/dfef7b0ef7a88727636f5ad680a464e33e345.pdf>

- <sup>203</sup> Zhang, Qian, *et al.* Inborn Errors of Type I IFN Immunity in Patients with Life-Threatening COVID-19. *Science*, vol. 370, no. 6515, October 23, 2020, p. eabd4570. doi:10.1126/science.abd4570.
- <sup>204</sup> Noah G. Schwartz, Anne C. Moorman, Anna Makaretz, Karen T. Chang, Victoria T. Chu, Christine M. Szablewski, Anna R. Yousaf, Marie M. Brown, Ailis Clyne, Amanda DellaGrotta, Jan Drobeniuc, Jacqueline Korpics, Adam Muir, Cherie Drenzek, Utpala Bandy, Hannah L. Kirking, Jacqueline E. Tate, Aron J. Hall, Tatiana M. Lanzieri and Rebekah J. Stewart (2020) *Adolescent with COVID-19 as the Source of an Outbreak at a 3-Week Family Gathering — Four States, June–July 2020*. CDC MMWR vol. 69, October 5, 2020. <https://www.cdc.gov/mmwr/volumes/69/wr/pdfs/mm6940e2-H.pdf>
- <sup>205</sup> Peccia, J., Zulli, A., Brackney, D.E. *et al.* Measurement of SARS-CoV-2 RNA in wastewater tracks community infection dynamics. *Nature Biotechnology* **38**, 1164–1167 (2020). <https://doi.org/10.1038/s41587-020-0684-z> and <https://www.nature.com/articles/s41587-020-0684-z.pdf>
- <sup>206</sup> MIT begins testing wastewater to help detect Covid-19 on campus. <https://news.mit.edu/2020/testing-wastewater-covid-19-1002>
- <sup>207</sup> Gusella JF, Wexler NS, Conneally PM, Naylor SL, Anderson MA, Tanzi RE, Watkins PC, Ottina K, Wallace MR, Sakaguchi AY, *et al.* (1983) A polymorphic DNA marker genetically linked to Huntington's disease. *Nature*. 1983 November 17-23; 306(5940): 234-8. doi: 10.1038/306234a0 <https://www.nature.com/articles/306234a0>
- <sup>208</sup> Mazini PS, Alves HV, Reis PG, Lopes AP, Sell AM, Santos-Rosa M, Visentainer JE, Rodrigues-Santos P. (2016) *Gene Association with Leprosy: A Review of Published Data*. *Frontiers in Immunology*. 2016 January 12; 6:658. doi: 10.3389/fimmu.2015.00658 [www.ncbi.nlm.nih.gov/pmc/articles/PMC4709443/pdf/fimmu-06-00658.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4709443/pdf/fimmu-06-00658.pdf)
- <sup>209</sup> Dallmann-Sauer M, Fava VM, Gzara C, Orlova M, Van Thuc N, Thai VH, *et al.* (2020) The complex pattern of genetic associations of leprosy with HLA class I and class II alleles can be reduced to four amino acid positions. *PLoS Pathog* 16(8): e1008818. <https://doi.org/10.1371/journal.ppat.1008818> <https://journals.plos.org/plospathogens/article/file?id=10.1371/journal.ppat.1008818&type=printable>
- <sup>210</sup> Zeberg, H. and Pääbo, S. (2020) The major genetic risk factor for severe COVID-19 is inherited from Neanderthals. *Nature* (30 September 2020). <https://doi.org/10.1038/s41586-020-2818-3> [https://www.nature.com/articles/s41586-020-2818-3\\_reference.pdf](https://www.nature.com/articles/s41586-020-2818-3_reference.pdf)
- <sup>211</sup> Kenney AD, Dowdle JA, Bozzacco L, McMichael TM, St Gelais C, Panfil AR, Sun Y, Schlesinger LS, Anderson MZ, Green PL, López CB, Rosenberg BR, Wu L, Yount JS. *Human Genetic Determinants of Viral Diseases*. *Annu Rev Genet*. 2017 Nov 27; 51:241-263. doi: 10.1146/annurev-genet-120116-023425. Epub 2017 August 30. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6038703/pdf/nihms-978947.pdf>
- <sup>212</sup> Rubin R. *Investigating Whether Blood Type Is Linked to COVID-19 Risk*. *JAMA*. 2020 September 16. doi: 10.1001/jama.2020.16516. <https://jamanetwork.com/journals/jama/fullarticle/2770889>
- <sup>213</sup> Kaiser, Jocelyn. “How Sick Will the Coronavirus Make You? The Answer May Be in Your Genes.” *Science*, March 27, 2020. doi:10.1126/science.abb9192

- <sup>214</sup> Liu, Y.-M., Xie, J., Chen, M.-M., Zhang, X., Cheng, X., Li, H., Zhou, F., Qin, J.-J., Lei, F., Chen, Z., Lin, L., Yang, C., Mao, W., Chen, G., Lu, H., Xia, X., Wang, D., Liao, X., Yang, J., Huang, X., Zhang, B.-H., Yuan, Y., Cai, J., Zhang, X.-J., Wang, Y., Zhang, X., She, Z.-G., Li, H., *Kidney function indicators predict adverse outcomes of COVID-19*. *Med* (2020).  
[https://www.cell.com/med/pdf/S2666-6340\(20\)30017-9.pdf](https://www.cell.com/med/pdf/S2666-6340(20)30017-9.pdf)
- <sup>215</sup> Janardhan, Vallabh, et al. “COVID-19 as a Blood Clotting Disorder Masquerading as a Respiratory Illness: A Cerebrovascular Perspective and Therapeutic Implications for Stroke Thrombectomy.” *J of Neuroimaging*, vol. 30, no. 5, September 2020, pp. 555–561.  
<https://onlinelibrary.wiley.com/doi/epdf/10.1111/jon.12770>
- <sup>216</sup> Bastard, Paul, et al. Autoantibodies against Type I IFNs in Patients with Life-Threatening COVID-19. *Science*, vol. 370, no. 6515, October 23, 2020, p. eabd4585. doi:10.1126/science.abd4585.
- <sup>217</sup> Torrente-Rodriguez, Rebeca *et al.*, *SARS-CoV-2 RapidPlex: A Graphene-Based Multiplexed Telemedicine Platform for Rapid and Low-Cost COVID-19 Diagnosis and Monitoring*. *Matter* (Oct 1, 20)  
<https://doi.org/10.1016/j.matt.2020.09.027> ; [www.cell.com/matter/pdfExtended/S2590-2385\(20\)30553-1](http://www.cell.com/matter/pdfExtended/S2590-2385(20)30553-1)
- <sup>218</sup> Sanjay E. Sarma, Stephen A. Weis, Daniel W. Engels (2002) *RFID Systems, Security and Privacy Implications*. White Paper. MIT Auto ID Center. Published November 1, 2002.  
[https://cocoa.ethz.ch/downloads/2014/06/None\\_MIT-AUTOID-WH-014.pdf](https://cocoa.ethz.ch/downloads/2014/06/None_MIT-AUTOID-WH-014.pdf)
- <sup>219</sup> Mark Roberti (2004) *The 5-cent Challenge*. *RFID Journal*. Published August 30, 2004.  
<https://www.rfidjournal.com/the-5-cent-challenge>
- <sup>220</sup> Zhou, Lian, et al. *Detection of SARS-CoV-2 in Exhaled Breath from COVID-19 Patients Ready for Hospital Discharge*. preprint, *Public and Global Health*, 2 June 2020. doi:10.1101/2020.05.31.20115196.  
<https://www.medrxiv.org/content/10.1101/2020.05.31.20115196v1.full.pdf>
- <sup>221</sup> Ma, Jianxin, et al. *Exhaled Breath Is a Significant Source of SARS-CoV-2 Emission*. preprint, *Public and Global Health*, 2 June 2020. doi:10.1101/2020.05.31.20115154.  
<https://www.medrxiv.org/content/10.1101/2020.05.31.20115154v1.full.pdf>
- <sup>222</sup> Emam, Shadi, et al. *Verification of Gas Sensors to Detect Alzheimer’s Disease Biomarkers with Diabetic Rats*. *ALZ*, 2020. *alz.confex.com*, <https://alz.confex.com/alz/20amsterdam/meetingapp.cgi/Paper/43611>  
<https://www.wired.com/story/could-breathalyzers-make-covid-testing-quicker-and-easier>
- <sup>223</sup> Kybert, Nicholas, et al. “Exploring Ovarian Cancer Screening Using a Combined Sensor Approach: A Pilot Study.” *AIP Advances*, vol. 10, no. 3, March 2020, p. 035213. doi:10.1063/1.5144532  
<https://aip.scitation.org/doi/pdf/10.1063/1.5144532>
- <sup>224</sup> Shan, Benjie, et al. “Multiplexed Nanomaterial-Based Sensor Array for Detection of COVID-19 in Exhaled Breath.” *ACS Nano*, vol. 14, no. 9, Sept. 2020, pp. 12125–32. *ACS Publications*, doi:10.1021/acsnano.0c05657. <https://pubs.acs.org/doi/pdf/10.1021/acsnano.0c05657>
- <sup>225</sup> Kahn N, Lavie O, Paz M, Segev Y, Haick H. (2015) *Dynamic Nanoparticle-Based Flexible Sensors: Diagnosis of Ovarian Carcinoma from Exhaled Breath*. *Nano Lett.* 2015 October 14;15(10):7023-8. doi: 10.1021/acs.nanolett.5b03052. Epub 2015 Sep 11.  
<https://pubs.acs.org/doi/pdf/10.1021/acs.nanolett.5b03052>

- <sup>226</sup> Chandrapalan, S. *et al* (2020) *Breath diagnostics in the era of SARS-CoV-2 - clinical and research arena*. Journal of Breath Research 14 042002  
<https://iopscience.iop.org/article/10.1088/1752-7163/ab924a/pdf>
- <sup>227</sup> Oliveira AD, Prats C, Espasa M, Zarzuela Serrat F, Montañola Sales C, Silgado A, Codina DL, Arruda ME, I Prat JG, Albuquerque J. (2017) *The Malaria System MicroApp: A New, Mobile Device-Based Tool for Malaria Diagnosis*. JMIR Research Protocols. 2017 April 25; 6(4):e70. doi: 10.2196/resprot.6758.
- <sup>228</sup> Pirstill CW, Coté GL. *Malaria Diagnosis Using a Mobile Phone Polarized Microscope*. Nature Science Reports 2015 Aug 25; 5:13368. doi: 10.1038/srep13368.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4548194/pdf/srep13368.pdf>
- <sup>229</sup> Hwang SI, Franconi NG, Rothfuss MA, Bocan KN, Bian L, White DL, Burkert SC, Euler RW, Sopher BJ, Vinay ML, Sejdic E, Star A. (2019) *Tetrahydrocannabinol Detection Using Semiconductor-Enriched Single-Walled Carbon Nanotube Chemiresistors*. ACS Sensors 2019 August 23; 4(8): pp. 2084-2093. doi: 10.1021/acssensors.9b00762. Epub 2019 August 1.
- <sup>230</sup> SARS-CoV-2 (COVID-19) Detection Using the Breath Analyzer TeraSystem  
<https://clinicaltrials.gov/ct2/show/NCT04497610>
- <sup>231</sup> Parinaz Fozouni, Sungmin Son, María Díaz de León Derby, Gavin J. Knott, Carley N. Gray, Michael V. D'Ambrosio, Chunyu Zhao, Neil A. Switz, G. Renuka Kumar, Stephanie I. Stephens, Daniela Boehm, Chia-Lin Tsou, Jeffrey Shu, Abdul Bhuiya, Max Armstrong, Andrew Harris, Jeannette M. Osterloh, Anke Meyer-Franke, Charles Langelier, Katherine S. Pollard, Emily D. Crawford, Andreas S. Puschnik, Maira Phelps, Amy Kistler, Joseph L. DeRisi, Jennifer A. Doudna, Daniel A. Fletcher and Melanie Ott (2020) *Direct detection of SARS-CoV-2 using CRISPR-Cas13a and a mobile phone*. Published 30 Sep 2020  
<https://doi.org/10.1101/2020.09.28.20201947>  
<https://www.medrxiv.org/content/10.1101/2020.09.28.20201947v1.full.pdf>
- <sup>232</sup> Rockefeller Foundation. *National Covid-19 testing and tracing action plan*. July 16, 2020.  
[https://www-rockefellerfoundation-org.libproxy.mit.edu/wp-content/uploads/2020/07/TheRockefellerFoundation\\_ExecutiveSummary\\_7\\_20.pdf](https://www-rockefellerfoundation-org.libproxy.mit.edu/wp-content/uploads/2020/07/TheRockefellerFoundation_ExecutiveSummary_7_20.pdf)
- <sup>233</sup> “The Nobel Prize in Chemistry 2020.” <https://www.nobelprize.org/uploads/2020/10/press-chemistryprize2020.pdf> & <https://www.nobelprize.org/prizes/chemistry/2020/press-release/> (10-7-2020)
- <sup>234</sup> Service, R. New Test Detects Coronavirus in 5 Minutes. *Science*, 10-8-20. doi:10.1126/science.abf1752  
<https://www.sciencemag.org/news/2020/10/new-test-detects-coronavirus-just-5-minutes>
- <sup>235</sup> Service, Robert. In ‘Milestone,’ FDA OKs Simple, Accurate Coronavirus Test That Could Cost \$5. *Science*, August 27-28, 2020. doi:10.1126/science.abe5319
- <sup>236</sup> The Nobel Prize in Physiology or Medicine 2020. October 5, 2020.  
<https://www.nobelprize.org/prizes/medicine/2020/press-release/>
- <sup>237</sup> Schweitzer, Albert (1922) *The Decay and the Restoration of Civilization* (Translated by C. T. Champion. Published by A. C. Black, London, 1923)  
<https://ia800904.us.archive.org/9/items/p1decayrestorati00schwuoft/p1decayrestorati00schwuoft.pdf>
- <sup>238</sup> David M Cutler and Larry H Summers. *COVID-19 Pandemic and the \$16 Trillion Virus*. JAMA. Pub Oct 12, 2020. doi:10.1001/jama.2020.19759 <https://jamanetwork.com/journals/jama/fullarticle/2771764>

- 
- <sup>239</sup> “The World’s \$80 Trillion Economy - in One Chart.” *World Economic Forum*,  
<https://www.weforum.org/agenda/2018/10/the-80-trillion-world-economy-in-one-chart/>
- <sup>240</sup> <https://www.worldometers.info/gdp/gdp-by-country>
- <sup>241</sup> Lulla, Valeria, et al. *Antisense Oligonucleotides Target a Nearly Invariant Structural Element from the SARS-CoV-2 Genome and Drive RNA Degradation*. preprint, *Molecular Biology*, 19 September 2020.  
doi:10.1101/2020.09.18.304139 <https://www.biorxiv.org/content/10.1101/2020.09.18.304139v1.full.pdf>
- <sup>242</sup> David Chiriboga, Juan Garay, Paulo Buss, Rocío Sáenz Madrigal, Laetitia Charmaine Rispel (2020)  
“Health Inequity during the COVID-19 Pandemic: A Cry for Ethical Global Leadership.” *The Lancet*,  
vol. 395, no. 10238, May 2020, pp. 1690–1691. doi:10.1016/S0140-6736(20)31145-4  
<https://www.thelancet.com/action/showPdf?pii=S0140-6736%2820%2931145-4>
- <sup>243</sup> Lorenzo Casalino, Zied Gaieb, Jory A. Goldsmith, Christy K. Hjorth, Abigail C. Dommer, Aoife M.  
Harbison, Carl A. Fogarty, Emilia P. Barros, Bryn C. Taylor, Jason S. McLellan, Elisa Fadda, and Rommie  
E. Amaro (2020) “Beyond Shielding: The Roles of Glycans in the SARS-CoV-2 Spike Protein.” *ACS  
Central Science*, September 2020, p. acscentsci.0c01056. doi:10.1021/acscentsci.0c01056.  
<https://pubs.acs.org/doi/pdf/10.1021/acscentsci.0c01056>
- <sup>244</sup> Hodcroft, Emma B., et al. *Emergence and Spread of a SARS-CoV-2 Variant through Europe in the  
Summer of 2020*. preprint, *Epidemiology*, 28 October 2020. doi:10.1101/2020.10.25.20219063.
- <sup>245</sup> Shaman, J. and Galanti, Marta (2020) Will SARS-CoV-2 become endemic? *Science* 14 Oct 2020.  
DOI: 10.1126/science.abe5960  
<https://science.sciencemag.org/content/early/2020/10/13/science.abe5960/tab-pdf>
- <sup>246</sup> Cao L, Goreshnik I, Coventry B, Case JB, Miller L, Kozodoy L, Chen RE, Carter L, Walls L, Park YJ,  
Stewart L, Diamond M, Veessler D, Baker D. *De novo design of picomolar SARS-CoV-2 miniprotein  
inhibitors*. bioRxiv [Preprint]. 2020 Aug 3. DOI: 10.1101/2020.08.03.234914. *Science* 23 October 2020:  
Vol. 370, Issue 6515, pp. 426-431 DOI: 10.1126/science.abd9909.  
<https://www.biorxiv.org/content/10.1101/2020.08.03.234914v1.full.pdf>
- <sup>247</sup> Coase, R. H. (1937) “The Nature of the Firm.” *Economica*, vol. 4, no. 16, Nov. 1937, pp. 386–405.  
doi:10.1111/j.1468-0335.1937.tb00002.x  
<https://www.law.uchicago.edu/files/file/coase-nature.pdf> <http://bit.ly/COASE5PAPERS>
- <sup>248</sup> Stephanie K. Pell and Christopher Soghoian (2014) Your Secret Stingray’s No Secret Anymore: The  
Vanishing Government Monopoly Over Cell Phone Surveillance and Its Impact On National Security  
and Consumer Privacy. *Harvard Journal of Law and Technology* Volume 28, Number 1 Fall 2014
- <sup>249</sup> Kevin S. Bankston and Ashkan Soltani (2014) Tiny Constables and the Cost of Surveillance: Making  
Cents Out of United States v. Jones, 123 *Yale Law Journal* ONLINE 335 (2014)  
<http://yalelawjournal.org/forum/tiny-constables-and-the-cost-of-surveillance-making-cents-out-of-united-states-v-jones>
- <sup>250</sup> Roosa Tikkanen and Melinda K. Abrams (2020) U.S. Health Care from a Global Perspective, 2019:  
Higher Spending, Worse Outcomes? January 30, 2020  
<https://www.commonwealthfund.org/publications/issue-briefs/2020/jan/us-health-care-global-perspective-2019>



- 
- <sup>251</sup> Landgrebe, D. (1998) *Information Extraction Principals and Methods for Multispectral and Hyperspectral Image Data, Information processing for Remote Sensing*. World Scientific Publishing Co. <https://engineering.purdue.edu/~landgreb/whitepaper.pdf>
- <sup>252</sup> Moghimi, Ali, et al. "A Novel Machine Learning Approach to Estimate Grapevine Leaf Nitrogen Concentration Using Aerial Multispectral Imagery." *Remote Sensing*, vol. 12, no. 21, October 2020, p. 3515. doi:10.3390/rs12213515. <https://www.mdpi.com/2072-4292/12/21/3515/pdf>
- <sup>253</sup> Huang, Y., Ren, Z., Li, D. *et al.* Phenotypic techniques and applications in fruit trees: a review. *Plant Methods* **16**, 107 (2020). <https://doi.org/10.1186/s13007-020-00649-7>
- <sup>254</sup> Lorenzen, Bent & Arne, Jensen (1988) "Reflectance of Blue, Green, Red and near Infrared Radiation from Wetland Vegetation Used in a Model Discriminating Live and Dead above Ground Biomass." *New Phytologist*, vol. 108, no. 3, March 1988, pp. 345–55. doi:10.1111/j.1469-8137.1988.tb04173.x.
- <sup>255</sup> Soo Chung, Lane E. Breshears and Jeong-Yeol Yoon, "Smartphone Near Infrared Monitoring of Plant Stress," *Computers and Electronics in Ag*, 2018, 154: 93-98. <https://doi.org/10.1016/j.compag.2018.08.046>
- <sup>256</sup> Marturano F, Ciparisse JF, Chierici A, d'Errico F, Di Giovanni D, Fumian F, Rossi R, Martellucci L, Gaudio P, Malizia A. *Enhancing Radiation Detection by Drones through Numerical Fluid Dynamics Simulations*. *Sensors* (Basel). 2020 March 23; 20(6):1770. doi: 10.3390/s20061770. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7147154/pdf/sensors-20-01770.pdf>
- <sup>257</sup> William Joseph Butera (2002) *Programming a Paintable Computer*. Submitted to the Program in Media Arts & Sciences, School of Architecture and Planning, in partial fulfillment of the requirements for the degree of Doctor of Philosophy in Media Arts & Sciences, Massachusetts Institute of Technology February 2002 <http://cba.mit.edu/docs/theses/02.02.butera.pdf>  
[http://cba.mit.edu/events/01.11.retreat/Butera/Butera\\_files/v3\\_document.htm](http://cba.mit.edu/events/01.11.retreat/Butera/Butera_files/v3_document.htm)
- <sup>258</sup> Wicaksono, I., Tucker, C.I., Sun, T. *et al.* A tailored, electronic textile conformable suit for large-scale spatiotemporal physiological sensing in vivo. *npj Flex Electron* **4**, 5 (2020). <https://doi.org/10.1038/s41528-020-0068-y>
- <sup>259</sup> Mark Weiser, Rich Gold and John Seeley Brown (1999) The origins of ubiquitous computing research at PARC in the late 1980s. *IBM Systems Journal*, vol 38, no. 4, pp. 693-696, 1999. <http://www.cs.cmu.edu/~jasonh/courses/ubicomp-sp2007/papers/03-weiser-origins.pdf>
- <sup>260</sup> Mark Weiser (1991) *The Computer for the 21st Century*. *Scientific American* 265, No. 3, pp. 94 –104 (September 1991). <https://www.lri.fr/~mbl/Stanford/CS477/papers/Weiser-SciAm.pdf>
- <sup>261</sup> William D. Nordhaus (2001) *Progress of Computing*. Cowles Foundation Discussion Paper No. 1324. Cowles Foundation For Research in Economics, Yale University, New Haven, Connecticut, USA. <http://cowles.econ.yale.edu/> <https://cowles.yale.edu/sites/default/files/files/pub/d13/d1324.pdf> and <https://aiimpacts.org/trends-in-the-cost-of-computing/>
- <sup>262</sup> David H Wolpert. (2019) The stochastic thermodynamics of computation, *Journal of Physics A: Mathematical and Theoretical* (2019). DOI: 10.1088/1751-8121/ab0850
- <sup>263</sup> Charles, J. (2012) <https://jcharles00.wordpress.com/2012/05/>
- <sup>264</sup> What is IPv6? <https://www.internetsociety.org/deploy360/ipv6/>

- 
- <sup>265</sup> Hinden, Bob and Krishnan, Suresh (2018) IPv6 is an Internet Standard.  
<https://www.ietf.org/blog/ipv6-internet-standard/>
- <sup>266</sup> Raffi C. Krikorian (2004) *Internet 0*. Submitted to the Program in Media Arts & Sciences, School of Architecture and Planning in partial fulfillment of the requirements for the degree of Master of Science in Media Arts & Sciences at the Massachusetts Institute Of Technology, August 2004  
<https://dspace.mit.edu/bitstream/handle/1721.1/28866/60412683-MIT.pdf?sequence=2&isAllowed=y>
- <sup>267</sup> Ishii, H. and Ullmer, B. (1997) *Tangible Bits: Towards Seamless Interfaces between People, Bits, and Atoms*. Proceedings of CHI'97 (ACM, March 1997) <http://tangible.media.mit.edu/person/hiroshi-ishii/>
- <sup>268</sup> Hiroshi Ishi <https://tangible.media.mit.edu/person/hiroshi-ishii/>
- <sup>269</sup> Gershenfeld, Neil A. *When Things Start to Think*. 1st ed, Henry Holt, 1999.
- <sup>270</sup> Nelson Minar, Matthew Gray, Oliver Roup, Raffi Krikorian and Patti Maes “Hive: Distributed Agents for Networking Things.” *IEEE Concurrency*, vol. 8, no. 2, Apr. 2000, pp. 24-33. doi:10.1109/4434.846191
- <sup>271</sup> Gershenfeld, Neil. <http://ng.cba.mit.edu/>
- <sup>272</sup> Origins of IoT <https://autoid.mit.edu/>
- <sup>273</sup> Greengard, Samuel. *The Internet of Things*. MIT Press, 2015.
- <sup>274</sup> <https://autoid.mit.edu/people-2>
- <sup>275</sup> <https://openlearning.mit.edu/about/our-team/sanjay-sarma>
- <sup>276</sup> Simon, Herbert, A. The Steam Engine and the Computer: What Makes Technology Revolutionary. *EDUCOM Bulletin*, vol. 22, no.1, pp. 2-5 Spring 1987  
<https://er.educause.edu/-/media/files/article-downloads/erm0132.pdf>  
<http://digitalcollections.library.cmu.edu/awweb/awarchive?type=file&item=34057>
- <sup>277</sup> Sarma, S., Brock, D. and Engels, D. (2001) “Radio Frequency Identification and the Electronic Product Code,” *IEEE Micro*, vol. 21, no. 6, December 2001, pp. 50–54. doi:10.1109/40.977758.
- <sup>278</sup> MIT Project Oxygen (2001) Computer Science and Artificial Intelligence Laboratory, MIT.  
<http://www.ai.mit.edu/projects/oxygen/oxygen-book2001/oxygenbook2001.pdf>
- <sup>279</sup> Katz, Randall (2002) Sketch - <http://oxygen.csail.mit.edu/videosketching.html> and Sketch Video Demonstration <http://oxygen.csail.mit.edu/videos/sketching.mpeg>
- <sup>280</sup> Project Oxygen. Massachusetts Institute of Technology <http://oxygen.csail.mit.edu/Overview.html>
- <sup>281</sup> Jorgenson, Dale Weldeau, et al., editors. *Measuring and Sustaining the New Economy: Report of a Workshop*. National Academy Press, 2002. <https://www.nap.edu/download/10282>
- <sup>282</sup> <https://www.pcgamesn.com/intel/memory-nand-ssd-price-drop-2019-oversupply>
- <sup>283</sup> <https://www.iotone.com/term/decreasing-cost-of-storage/t172>
- <sup>284</sup> Golubchik, Leana, Khuller, Samir, Mukherjee, Koyel and Yao, Yuan. (2013). *To send or not to send: Reducing the cost of data transmission*. IEEE Infocom. 2472-2478. Doi: 10.1109/INFCOM.2013.6567053
- <sup>285</sup> Asimov, Isaac (1953) [http://www.self.gutenberg.org/articles/Sally\\_\(short\\_story\)](http://www.self.gutenberg.org/articles/Sally_(short_story))
- <sup>286</sup> Asimov, Isaac. *The Complete Robot*. HarperCollins, 1995.  
[https://en.wikipedia.org/wiki/Sally\\_\(short\\_story\)](https://en.wikipedia.org/wiki/Sally_(short_story))

- <sup>287</sup> Sanjay Sarma, David Brock and Kevin Ashton (1999) “*The Networked Physical World - Proposals for Engineering the Next Generation of Computing, Commerce, and Automatic-Identification*,” MIT Auto-ID Center White Paper. MIT-AUTOID-WH001, 1999. <https://autoid.mit.edu/publications-0>  
<https://pdfs.semanticscholar.org/88b4/a255082d91b3c88261976c85a24f2f92c5c3.pdf>
- <sup>288</sup> Ghasempour, Alireza. *Internet of Things in Smart Grid: Architecture, Applications, Services, Key Technologies, and Challenges*. Inventions, vol. 4, no. 1, Mar 2019, p. 22. doi:10.3390/inventions4010022
- <sup>289</sup> Christopher Greer, Martin Burns, David Wollman and Edward Griffor (2019) *Cyber-Physical Systems and Internet of Things*. <https://doi.org/10.6028/NIST.SP.1900-202>  
<https://nvlpubs.nist.gov/nistpubs/SpecialPublications/NIST.SP.1900-202.pdf>
- <sup>290</sup> Edward A. Lee and Sanjit A. Seshia. *Introduction to Embedded Systems, A Cyber-Physical Systems Approach*, Second Edition, MIT Press, 2017. ISBN 978-0-262-53381-2  
[https://ptolemy.berkeley.edu/books/leeseshia/releases/LeeSeshia\\_DigitalV2\\_2.pdf](https://ptolemy.berkeley.edu/books/leeseshia/releases/LeeSeshia_DigitalV2_2.pdf)
- <sup>291</sup> Rajkumar R, Lee I, Sha L, Stankovic J (2010) *Cyber-Physical Systems: The Next Computing Revolution*. Proceedings of the 47th Design Automation Conference (IEE, Anaheim, CA), pp 731– 736.  
<https://ieeexplore.ieee.org/document/5523280>  
<https://www.cs.virginia.edu/~stankovic/psfiles/Rajkumar-DAC2010-Final.pdf>
- <sup>292</sup> Robert E. Horn (2015) *Information Design: Emergence of a New Profession*  
<https://steinhardtapps.es.its.nyu.edu/create/courses/2015/reading/Horn.pdf>
- <sup>293</sup> Jacobson, Robert E., editor. *Information Design*. MIT Press, 1999.  
<https://epdf.pub/queue/information-design.html>
- <sup>294</sup> Jimmy Soni and Rob Goodman (2018) *A Mind at Play: How Claude Shannon Invented the Information Age*. Simon & Schuster (July 17, 2018) ISBN13: 9781476766690  
<https://spectrum.ieee.org/geek-life/history/a-man-in-a-hurry-claude-shannons-new-york-years>
- <sup>295</sup> Datta, Shoumen (2020) *HIP: History & Evolution of the Internet of Things - Can PEAS Improve Performance?* Unpublished essay. MIT Library <https://dspace.mit.edu/handle/1721.1/111021>
- <sup>296</sup> IoT. MIT Auto-ID Labs. Massachusetts Institute of Technology. <https://autoid.mit.edu/>
- <sup>297</sup> Pham NM, Karlen W, Beck HP, Delamarche E. *Malaria and the 'last' parasite: how can technology help?* *Malaria Journal* 2018 July 11; 17(1):260. doi: 10.1186/s12936-018-2408-0  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6042346/pdf/12936\\_2018\\_Article\\_2408.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6042346/pdf/12936_2018_Article_2408.pdf)
- <sup>298</sup> Pierce, David (2016) *Project Ara Lives: Google's Modular Phone Is Ready for You Now*.  
<https://www.wired.com/2016/05/project-ara-lives-googles-modular-phone-is-ready/>
- <sup>299</sup> NK Labs, Cambridge, Massachusetts <https://www.nklabs.com/ara-knaian>
- <sup>300</sup> Bhardwaj, Vinay, and Vinod Gaur. “Raman Spectroscopy as a Blood Glucose Monitoring Tool.” *European Pharmaceutical Review*, vol. 22, no. 3, 2017, pp. 26–29.
- <sup>301</sup> Pandey R, Paidi SK, Valdez TA, Zhang C, Spegazzini N, Dasari RR, Barman I. *Noninvasive Monitoring of Blood Glucose with Raman Spectroscopy*. *Acc Chem Res*. 2017 February 21; 50(2):264-272. doi: 10.1021/acs.accounts.6b00472. Epub 2017 January 10.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5896772/pdf/nihms955832.pdf>

- <sup>302</sup> Kakkar, T., Richards, B., Jha, A., Saha, S., Ajjan, R., Grant, P. and Jose, G. (2015) Glucosense: Photonic Chip Based Non-Invasive Glucose Monitor. *Diabetes Technology Therapeutics*, vol. 17, pp. A82-A83
- <sup>303</sup> Jose, Gin. GLUCOSENSE. University of Leeds, UK. [www.leeds.ac.uk/site/custom\\_scripts/profile-single.php?profileTypeID=3&categoryID=2000&profileID=116](http://www.leeds.ac.uk/site/custom_scripts/profile-single.php?profileTypeID=3&categoryID=2000&profileID=116)
- <sup>304</sup> Jung, D.G.; Jung, D.; Kong, S.H. (2017) A Lab-on-a-Chip-Based Non-Invasive Optical Sensor for Measuring Glucose in Saliva. *Sensors* **2017**, *17*, 2607. <https://www.mdpi.com/1424-8220/17/11/2607/pdf>
- <sup>305</sup> Zhang R, Liu S, Jin H, Luo Y, Zheng Z, Gao F, Zheng Y. (2019) *Noninvasive Electromagnetic Wave Sensing of Glucose*. *Sensors* (Basel). 2019 March 7; 19(5):1151. doi: 10.3390/s19051151 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6427587/pdf/sensors-19-01151.pdf>
- <sup>306</sup> Narayan K MV, Zhang P, Kanaya AM, et al. *Diabetes: The Pandemic and Potential Solutions*. In: Jamison DT, Breman JG, Measham AR, et al., editors. *Disease Control Priorities in Developing Countries*. 2nd ed. Washington (DC): International Bank for Reconstruction and Development, The World Bank. 2006. Chapter 30. Oxford University Press, NY. <https://www.ncbi.nlm.nih.gov/books/NBK11777/> [https://www.ncbi.nlm.nih.gov/books/NBK11777/pdf/Bookshelf\\_NBK11777.pdf](https://www.ncbi.nlm.nih.gov/books/NBK11777/pdf/Bookshelf_NBK11777.pdf)
- <sup>307</sup> Villena Gonzales W, Mobashsher AT, Abbosh A. *The Progress of Glucose Monitoring-A Review of Invasive to Minimally and Non-Invasive Techniques, Devices and Sensors*. *Sensors* (Basel). 2019 Feb 15; 19(4):800. doi: 10.3390/s19040800 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6412701/pdf/sensors-19-00800.pdf>
- <sup>308</sup> Kang, Jeon Woong, et al. (2020) "Direct Observation of Glucose Fingerprint Using in Vivo Raman Spectroscopy." *Science Advances*, vol. 6, no. 4, Jan. 2020, p. eaay5206. doi:10.1126/sciadv.aay5206
- <sup>309</sup> Zhu, Lian, et al. (2013) "Direct Electrochemistry of Cholesterol Oxidase Immobilized on Gold Nanoparticles-Decorated Multiwalled Carbon Nanotubes and Cholesterol Sensing." *Talanta*, vol. 106, March 2013, pp. 192–99. doi:10.1016/j.talanta.2012.12.036
- <sup>310</sup> N. Agrawal, B. Zhang, C. Saha, C. Kumar, X. Pu and S. Kumar. *Ultra-Sensitive Cholesterol Sensor Using Gold and Zinc-Oxide Nanoparticles Immobilized Core Mismatch MPM/SPS Probe*. *Journal of Lightwave Technology*, vol. 38, no. 8, pp. 2523-2529, 15 April 2020. doi: 10.1109/JLT.2020.2974818
- <sup>311</sup> Oncescu V, Mancuso M, Erickson D. *Cholesterol testing on a smartphone*. *Lab Chip*. 2014 Feb 21; 14(4):759-63. doi: 10.1039/c3lc51194d. Epub 2013 December 13.
- <sup>312</sup> de Bold AJ, Borenstein HB, Veress AT, Sonnenberg H. (1981) *A rapid and potent natriuretic response to intravenous injection of atrial myocardial extract in rats*. *Life Sci* 1981; vol. 28: pp. 89–94
- <sup>313</sup> Sudoh, T., Kangawa, K., Minamino, N. *et al.* (1988) A new natriuretic peptide in porcine brain. *Nature* **332**, 78–81 (1988). <https://doi.org/10.1038/332078a0>
- <sup>314</sup> Sudoh T, Minamino N, Kangawa K, Matsuo H. *C-type natriuretic peptide (CNP): A new member of natriuretic peptide family identified in porcine brain*. *Biochem Biophys Res Commun* 1990; 168: 863–870
- <sup>315</sup> Pollok, Nicole E., et al. *Electrochemical Detection of NT-ProBNP Using a Metalloimmunoassay on a Paper Electrode Platform*. *ACS Sensors*, vol. 5, n. 3, Mar 2020, p 853-60. doi: 10.1021/acssensors.0c00167
- <sup>316</sup> Grabowska I, Sharma N, Vasilescu A, Iancu M, Badea G, Boukherroub R, Ogale S, Szunerits S. *Electrochemical Aptamer-Based Biosensors for the Detection of Cardiac Biomarkers*. *ACS Omega*. 2018 September 30; 3(9): pp. 12010-12018. doi: 10.1021/acsomega.8b01558. Epub 2018 September 26.

- <sup>317</sup> Gaetano Ruocco, Marco Pellegrini, Carmelo De Gori, Beatrice Franci, Ranuccio Nuti, Alberto Palazzuoli. (2016) The prognostic combined role of B-type natriuretic peptide, blood urea nitrogen and congestion signs persistence in patients with acute heart failure. *Journal of Cardiovascular Medicine*, vol. 17, no. 11, pp. 818-827 (2016) doi: 10.2459/JCM.0000000000000350
- <sup>318</sup> Fonarow GC, Peacock WF, Phillips CO, Givertz MM, Lopatin M; ADHERE Scientific Advisory Committee and Investigators. Admission B-type natriuretic peptide levels and in-hospital mortality in acute decompensated heart failure. *Journal of the American College of Cardiology*. 2007 May 15; 49(19): pp. 1943-50. doi: 10.1016/j.jacc.2007.02.037. Epub 2007 April 30.
- <sup>319</sup> Lin, Chung-Yin, et al. "Detection of Oxytocin, Atrial Natriuretic Peptide, and Brain Natriuretic Peptide Using Novel Imprinted Polymers Produced with Amphiphilic Monomers." *Journal of Peptide Science*, vol. 25, no. 3, March 2019. doi:10.1002/psc.3150
- <sup>320</sup> Ven Manda, Tommy D. Bennett and Zhongping Yang (2003) *Implantable biosensor devices for monitoring cardiac marker molecules* <https://patents.google.com/patent/US7632234B2/en> <https://patentimages.storage.googleapis.com/20/27/7b/4ba70677881f9c/US7632234.pdf>
- <sup>321</sup> Gutterman, David (2013) *Silent Myocardial Ischemia* <https://www.escardio.org/static-file/Escardio/education/live-events/courses/education-resource/Fri-11-SMI-Gutterman.pdf>
- <sup>322</sup> Deanfield JE, Maseri A, Selwyn AP, Ribeiro P, Chierchia S, Krikler S, Morgan M. (1983) *Myocardial ischaemia during daily life in patients with stable angina: its relation to symptoms and heart rate changes*. *Lancet*. 1983 Oct 1; 2(8353): pp. 753-758. doi: 10.1016/s0140-6736(83)92295-x.
- <sup>323</sup> Eric S. McLamore, Evangelyn Alocilja, Carmen Gomes, Sundaram Gunasekaran, Daniel Jenkins, Shoumen.P.A. Datta, Yanbin Li, Yu (Jessie) Mao, Sam R. Nugen, José I. Reyes-De-Corcuera, Paul Takhistov, Olga Tsyusko, Jarad P. Cochran, Tzuen-Rong (Jeremy) Tzeng, Jeong-Yeol Yoon, Chenxu Yu and Anhong Zhou (2021) *FEAST of Biosensors: Food, Environmental and Agricultural Sensing Technologies (FEAST) in North America*. *Biosensors and Bioelectronics*, 2021, 113011. ISSN 0956-5663 <https://doi.org/10.1016/j.bios.2021.113011> MIT Library <https://dspace.mit.edu/handle/1721.1/123983> <http://www.sciencedirect.com/science/article/pii/S0956566321000476>
- <sup>324</sup> Schoettker P, Degott J, Hofmann G, Proença M, Bonnier G, Lemkaddem A, Lemay M, Schorer R, Christen U, Knebel JF, Wuerzner A, Burnier M, Wuerzner G. *Blood pressure measurements with the OptiBP smartphone app validated against reference auscultatory measurements*. *Nature Science Reports* 2020 October 20; 10(1):17827. doi: 10.1038/s41598-020-74955-4 [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7576142/pdf/41598\\_2020\\_Article\\_74955.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7576142/pdf/41598_2020_Article_74955.pdf)
- <sup>325</sup> *Noninvasive Device Could End Daily Finger Pricking for People with Diabetes*. <https://medicalxpress.com/news/2015-07-noninvasive-device-daily-finger-people.html>
- <sup>326</sup> *Glucosense Monitoring*. University of Leeds, UK <https://www.youtube.com/watch?v=j--utQE9Pz8>
- <sup>327</sup> Sato H, Chiba H, Tashiro H, Ozaki Y.(2001) *Excitation wavelength-dependent changes in Raman spectra of whole blood and hemoglobin: comparison of the spectra with 514.5-, 720-, and 1064-nm excitation*. *J Biomed Opt*. 2001 July; 6(3): pp. 366-370. doi: 10.1117/1.1380668 <https://www.spiedigitallibrary.org/journalArticle/Download?fullDOI=10.1117%2F1.1380668>



- <sup>328</sup> Hasan MK, Haque MM, Adib R, Tumpa JF, Begum A, Love RR, Kim YL, Sheikh IA. (2018) *SmartHeLP: Smartphone-based Hemoglobin Level Prediction Using an Artificial Neural Network*. AMIA Annual Symposium Proceedings. 2018 December 5 : pp. 535-544.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6371334/pdf/2976625.pdf>
- <sup>329</sup> Mannino, R.G., Myers, D.R., Tyburski, E.A. *et al.* (2018) *Smartphone app for non-invasive detection of anemia using only patient-sourced photos*. *Nature Communications* 9, 4924 (2018).  
<https://doi.org/10.1038/s41467-018-07262-2>
- <sup>330</sup> Yan, Q., Zhi, N., Yang, L. *et al.* A highly sensitive uric acid electrochemical biosensor based on a nano-cube cuprous oxide/ferrocene/uricase modified glassy carbon electrode. *Sci Rep* 10, 10607 (2020).  
<https://doi.org/10.1038/s41598-020-67394-8>
- <sup>331</sup> Yang, Y., Song, Y., Bo, X. *et al.* A laser-engraved wearable sensor for sensitive detection of uric acid and tyrosine in sweat. *Nat Biotechnol* 38, 217–224 (2020). <https://doi.org/10.1038/s41587-019-0321-x>
- <sup>332</sup> Bastawrous A, Giardini ME, Bolster NM, Peto T, Shah N, Livingstone IA, Weiss HA, Hu S, Rono H, Kuper H, Burton M. (2016) *Clinical Validation of a Smartphone-Based Adapter for Optic Disc Imaging in Kenya*. *JAMA Ophthalmology* 2016 Feb; 134(2): pp.151-158. doi: 10.1001/jamaophthalmol.2015.4625  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5321504/pdf/emss-71475.pdf>
- <sup>333</sup> Di Santo P, Harnett DT, Simard T, Ramirez FD, Pourdjabbar A, Yousef A, Moreland R, Bernick J, Wells G, Dick A, Le May M, Labinaz M, So D, Motazedian P, Jung RG, Chandrasekhar J, Mehran R, Chong AY, Hibbert B. *Photoplethysmography using a smartphone application for assessment of ulnar artery patency: a randomized clinical trial*. *CMAJ*. 2018 April 3; 190(13): E380-E388.  
doi: 10.1503/cmaj.170432 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5880645/pdf/190e380.pdf>
- <sup>334</sup> Malamas, Peter (2011) *A 3-D Heart Model for Arrhythmia Simulation and Visualization*  
<https://sunfest.seas.upenn.edu/wp-content/uploads/2018/07/MALAMASPETER.pdf>
- <sup>335</sup> Saxon Leslie A. *Ubiquitous wireless ECG recording: a powerful tool physicians should embrace*. *J Cardiovasc Electrophysiol*. 2013 April; 24(4): pp. 480-483. doi: 10.1111/jce.12097. Epub 2013 Feb 19.
- <sup>336</sup> Fadel Adib, Hongzi Mao, Zachary Kabelac, Dina Katabi and Robert C. Miller. “Smart Homes That Monitor Breathing and Heart Rate.” *Proceedings of the 33rd Annual ACM Conference on Human Factors in Computing Systems*, Association for Computing Machinery, 2015, pp. 837–846. *ACM Digital Library*, doi:10.1145/2702123.2702200. <http://www.mit.edu/~fadel/papers/vitalradio-paper.pdf>  
<http://witrack.csail.mit.edu/vitalradio/>
- <sup>337</sup> Fadel Adib and Dina Katabi (2013) *See Through Walls with Wi-Fi*. SIGCOMM’13, August 12–16, 2013. Hong Kong, China. <https://people.csail.mit.edu/fadel/papers/wivi-paper.pdf>
- <sup>338</sup> Tomlinson S, Behrmann S, Cranford J, Louie M, Hashikawa A. (2018) *Accuracy of Smartphone-Based Pulse Oximetry Compared with Hospital-Grade Pulse Oximetry in Healthy Children*. *Telemed J E Health*. 2018 July; 24(7): pp. 527-535. doi: 10.1089/tmj.2017.0166. Epub 2017 December 7.
- <sup>339</sup> Li J, Lillehoj PB. *Microfluidic Magneto Immunosensor for Rapid, High Sensitivity Measurements of SARS-CoV-2 Nucleocapsid Protein in Serum*. *ACS Sensors* 2021 Feb 25. doi: 10.1021/acssensors.0c02561

- <sup>340</sup> Hansen CH, Yang D, Koussa MA, Wong WP. *Nanoswitch-linked immunosorbent assay (NLISA) for fast, sensitive, and specific protein detection*. Proc Natl Acad Sci. 2017 Sep 26; 114(39):10367-10372. doi: 10.1073/pnas.1708148114 [www.ncbi.nlm.nih.gov/pmc/articles/PMC5625919/pdf/pnas.201708148.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5625919/pdf/pnas.201708148.pdf)
- <sup>341</sup> Banerjee, Abhijit and Duflo, Esther (2019) *Good Economics for Hard Times*. PublicAffairs, a division of Perseus Books, a subsidiary of Hachette Book Groups (Nov 12, 2019) ISBN-13: 978-1610399500
- <sup>342</sup> Howes, Anton. *Arts and Minds: How the Royal Society of Arts Changed a Nation*. Princeton University Press, 2020. ISBN-13: 978-0691182643
- <sup>343</sup> Schoof M, Faust B, Saunders RA, Sangwan S, Rezelj V, Hoppe N, Boone M, Billesbølle CB, Puchades C, Azumaya CM, Kratochvil HT, Zimanyi M, Deshpande I, Liang J, Dickinson S, Nguyen HC, Chio CM, Merz GE, Thompson MC, Diwanji D, Schaefer K, Anand AA, Dobzinski N, Zha BS, Simoneau CR, Leon K, White KM, Chio US, Gupta M, Jin M, Li F, Liu Y, Zhang K, Bulkley D, Sun M, Smith AM, Rizo AN, Moss F, Brilot AF, Pourmal S, Trenker R, Pospiech T, Gupta S, Barsi-Rhyne B, Belyy V, Barile-Hill AW, Nock S, Liu Y, Krogan NJ, Ralston CY, Swaney DL, García-Sastre A, Ott M, Vignuzzi M; QCRG Structural Biology Consortium, Walter P, and Manglik A. *An ultrapotent synthetic nanobody neutralizes SARS-CoV-2 by stabilizing inactive Spike*. Science. 2020 December 18; 370(6523):1473-1479. doi: 10.1126/science.abe3255. <https://science.sciencemag.org/content/370/6523/1473/tab-pdf> <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7430568/pdf/nihpp-2020.08.08.238469.pdf>
- <sup>344</sup> Xiang Y, Nambulli S, Xiao Z, Liu H, Sang Z, Duprex WP, Schneidman-Duhovny D, Zhang C, Shi Y. *Versatile and multivalent nanobodies efficiently neutralize SARS-CoV-2*. Science. 2020 December 18; 370(6523):1479-1484. doi: 10.1126/science.abe4747. Epub 2020 November 5. <https://science.sciencemag.org/content/370/6523/1479/tab-pdf>
- <sup>345</sup> Huang AT, Garcia-Carreras B, Hitchings MDT, Yang B, Katzelnick LC, Rattigan SM, Borgert BA, Moreno CA, Solomon BD, Rodriguez-Barraquer I, Lessler J, Salje H, Burke D, Wesolowski A, Cummings DAT. *A systematic review of antibody mediated immunity to coronaviruses: antibody kinetics, correlates of protection, and association of antibody responses with severity of disease*. medRxiv [Preprint]. 2020 April 17: doi: 10.1101/2020.04.14.20065771. Update in: *Nature Communications* 2020 Sept 17; 11(1):4704. [www.ncbi.nlm.nih.gov/pmc/articles/PMC7217088/pdf/nihpp-2020.04.14.20065771.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC7217088/pdf/nihpp-2020.04.14.20065771.pdf)
- <sup>346</sup> Ibarondo FJ, Fulcher JA, Goodman-Meza D, Elliott J, Hofmann C, Hausner MA, Ferbas KG, Tobin NH, Aldrovandi GM, Yang OO. *Rapid Decay of Anti-SARS-CoV-2 Antibodies in Persons with Mild Covid-19*. N Engl J Med. 2020 September 10; 383(11):1085-1087. doi: 10.1056/NEJMc2025179. Epub 2020 July 21. Erratum in: N Engl J Med. 2020 July 23 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7397184/pdf/NEJMc2025179.pdf>
- <sup>347</sup> A human coronavirus evolves antigenically to escape antibody immunity Rachel Eguia<sup>1</sup>, Katharine H. D. Crawford<sup>1,2,3</sup>, Terry Stevens-Ayers<sup>4</sup>, Laurel Kelnhofner-Millevolte<sup>3</sup>, Alexander L. Greninger<sup>4,5</sup>, Janet A. Englund<sup>6,7</sup>, Michael J. Boeckh<sup>4</sup>, Jesse D. Bloom<sup>1,2</sup>,# bioRxiv preprint doi: <https://doi.org/10.1101/2020.12.17.423313>; this version posted December 18, 2020. <https://www.biorxiv.org/content/10.1101/2020.12.17.423313v1.full.pdf>

- <sup>348</sup> Boni MF. Vaccination and antigenic drift in influenza. *Vaccine*. 2008 Jul 18;26 Suppl 3(Suppl 3):C8-14. doi: 10.1016/j.vaccine.2008.04.011. PMID: 18773534; PMCID: PMC2603026. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2603026/pdf/nihms-60062.pdf>
- <sup>349</sup> Budd J, Miller BS, Manning EM, Lampos V, Zhuang M, Edelstein M, Rees G, Emery VC, Stevens MM, Keegan N, Short MJ, Pillay D, Manley E, Cox IJ, Heymann D, Johnson AM, McKendry RA. *Digital technologies in the public-health response to COVID-19*. *Nature Med* 2020 Aug 7; 26(8):1183-1192. doi: 10.1038/s41591-020-1011-4. Epub 2020 Aug 7. [www.nature.com/articles/s41591-020-1011-4.pdf](http://www.nature.com/articles/s41591-020-1011-4.pdf)
- <sup>350</sup> Butler SM. Four COVID-19 Lessons for Achieving Health Equity. *JAMA Health Forum*. Nov 5, 2020. doi:10.1001/jamahealthforum.2020 <https://jamanetwork.com/channels/health-forum/fullarticle/2772835>
- <sup>351</sup> <https://www.dict.cc/german-english/Lippenbekenntnis.html>
- <sup>352</sup> Y. Rong, A.V. Padrona, K. J. Hagerty, N. Nelson, S. Chic, N. O. Keyhani, J. Katz, S.P.A. Datta, C. Gomes, and E.S. McLamore (2018) Post Hoc Support Vector Machine Learning for Impedimetric Biosensors Based on Weak Protein–Ligand Interactions. *The Analyst*, vol. 143, no. 9, pp. 2066–2075 doi:10.1039/C8AN00065D <https://pubs.rsc.org/en/content/getauthorversionpdf/C8AN00065D>
- <sup>353</sup> Cassie A. Giacobassi, Daniela A. Oliveira, Cicero C. Pola, Dong Xiang, Yifan Tang, Shoumen Palit Austin Datta, Eric S. McLamore and Carmen Gomes (2020) *Sense-Analyze-Respond-Actuate (SARA) systems span nanoscale and macroscale actuation for detection of Escherichia coli in water* (in press)
- <sup>354</sup> Nassauer, Sarah. “WSJ News Exclusive | Walmart Scraps Plan to Have Robots Scan Shelves.” *Wall St J*, 2 Nov 2020. [www.wsj.com/articles/walmart-shelves-plan-to-have-robots-scan-shelves-11604345341](http://www.wsj.com/articles/walmart-shelves-plan-to-have-robots-scan-shelves-11604345341)
- <sup>355</sup> Bejan, Teresa (2017) *Mere Civility: Disagreement and the Limits of Toleration*. Harvard University Press, 2017. ISBN-13 : 978-0674545496
- <sup>356</sup> Sen, Amartya (1999) *Development as Freedom*. Alfred A. Knopf, New York, 1999. ISBN-13 : 978-0385720274
- <sup>357</sup> Sen, Amartya (2009) *The Idea of Justice*. Belknap Press of Harvard University Press; 1st edition (September 30, 2009) ISBN-13 : 978-0674036130
- <sup>358</sup> Datta, Shoumen. APPENDIX Figure 1: Description of Major Software Components <https://github.com/shoumendatta/ADD-DIGITAL> and James Lamb <https://github.com/jameslamb>
- <sup>359</sup> Message Queueing using an open lightweight broker, such as, Message Queueing Telemetry Transport, MQTT (<https://mqtt.org/>) or RabbitMQ (<https://www.rabbitmq.com/>) or heavy-duty Apache Kafka (<https://kafka.apache.org/>). Self-managed or run behind a managed IoT service from cloud providers: AWS IoT Core (<https://aws.amazon.com/iot-core/>) or Azure IoT Hub (<https://azure.microsoft.com/en-us/services/iot-hub/>) or related services provided by other vendors (<https://www.zdnet.com/article/the-top-cloud-providers-of-2020-aws-microsoft-azure-google-cloud-hybrid-saas/>).
- <sup>360</sup> Operational Data Store choices include InfluxDB (<https://www.influxdata.com/>), Apache Cassandra (<https://cassandra.apache.org/>) or Prometheus (<https://prometheus.io/>). Managed cloud database from Amazon <https://docs.aws.amazon.com/amazondynamodb/latest/developerguide/Introduction.html>.
- <sup>361</sup> Amazon S3 Glacier <https://aws.amazon.com/glacier/>

- <sup>362</sup> Enterprise Resource Planning (ERP) – Master Data Management: Architecture and Technology <https://www.element61.be/en/resource/master-data-management-mdm-architecture-technology>
- <sup>363</sup> Vesna Vučković (2008) *Image and its Matrix, Matrix and its Image*. Преглед НЦД (NCD Review) 12 (2008) 17–31 <http://elib.mi.sanu.ac.rs/files/journals/ncd/12/ncd12017.pdf>
- <sup>364</sup> H3: Uber’s Hexagonal Hierarchical Spatial Index <https://eng.uber.com/h3/>
- <sup>365</sup> Liu J, Ranka S, Kahveci T. *Classification and feature selection algorithms for multi-class CGH data*. *Bioinformatics*. 2008 July 1; 24(13): i86-95. doi: 10.1093/bioinformatics/btn145 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2718623/pdf/btn145.pdf>
- <sup>366</sup> Bommert, Andrea, et al. “Benchmark for Filter Methods for Feature Selection in High-Dimensional Classification Data.” *Computational Statistics & Data Analysis*, vol. 143, March 2020, p. 106839. doi:10.1016/j.csda.2019.106839.
- <sup>367</sup> Chen, R., Dewi, C., Huang, S. *et al.* Selecting critical features for data classification based on machine learning methods. *Journal of Big Data* 7, 52 (2020). <https://doi.org/10.1186/s40537-020-00327-4> <https://journalofbigdata.springeropen.com/track/pdf/10.1186/s40537-020-00327-4>
- <sup>368</sup> Object stores: Amazon <https://aws.amazon.com/s3/>; Google Cloud <https://cloud.google.com/storage>; Microsoft Azure Blob <https://azure.microsoft.com/en-us/services/storage/blobs/>; Apache Cassandra <https://medium.com/walmartglobaltech/building-object-store-storing-images-in-cassandra-walmart-scale-a6b9c02af593>
- <sup>369</sup> Query-Over-Files Engines: Presto (<https://prestodb.io/>), Apache Drill (<https://drill.apache.org/>) or Apache Spark SparkSQL (<https://spark.apache.org/sql/>). If using application-specific custom code that directly reads files, orchestrated with batch-scheduling engine: Apache Airflow (<https://airflow.apache.org/>) or Prefect (<https://www.prefect.io/>)
- <sup>370</sup> Lechner, M., Hasani, R., Amini, A. *et al.* Neural circuit policies enabling auditable autonomy. *Nature Machine Intelligence* 2, 642–652 (13 October 2020). <https://doi.org/10.1038/s42256-020-00237-3>
- <sup>371</sup> Analytical Data Store: Traditional relational database PostgreSQL (<https://www.postgresql.org/>) or hosted relational database products provided by cloud providers (<https://aws.amazon.com/rds/>). If intelligent caching of repeated queries is needed, data warehouse technologies (managed cloud service) include: Snowflake (<https://www.snowflake.com/>), Amazon Redshift (<https://aws.amazon.com/redshift>) or Google BigQuery (<https://cloud.google.com/bigquery>). For on-premises option: Teradata (<https://www.teradata.com/>).
- <sup>372</sup> Machine Learning (see cartoon on the last page of this article) model training tools: Apache Spark (<https://spark.apache.org/>), Dask (<https://dask.org/>) or Ray (<https://rise.cs.berkeley.edu/projects/ray/>). If application specificity does not require high degree of customization - use “autoML” tools - DataRobot (<https://www.datarobot.com/>), h2o (<https://docs.h2o.ai/h2o/latest-stable/h2o-docs/automl.html>), Amazon SageMaker Autopilot (<https://aws.amazon.com/blogs/aws/amazon-sagemaker-autopilot-fully-managed-automatic-machine-learning/>) or Google Cloud AutoML (<https://cloud.google.com/automl>).
- <sup>373</sup> Tauro, C.J., Ganesan, N., Mishra, S.R., & Bhagwat, A. (2012). Object Serialization: A Study of Techniques of Implementing Binary Serialization in C++, Java & .NET. *Intl J of Computer Applications*, 45, 25-29. <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.685.1077&rep=rep1&type=pdf>

- 
- <sup>374</sup> Common Object Request Broker Architecture (CORBA) <https://www.corba.org/>
- <sup>375</sup> Container: Docker <https://www.docker.com/resources/what-container>
- <sup>376</sup> Developer's language of choice may include (but the "menu" is not limited to):  
Java jar ([https://en.wikipedia.org/wiki/JAR\\_\(file\\_format\)](https://en.wikipedia.org/wiki/JAR_(file_format))),  
Python pickle file (<https://docs.python.org/3/library/pickle.html>),  
R rds file (<https://stat.ethz.ch/R-manual/R-devel/library/base/html/readRDS.html>) or a precompiled executable which can read in input data from "stdin" (standard input is a stream from which a program reads its input data) or from a file, created with C/C++ or language-agnostic description of a model:  
Predictive Model Markup Language (PMML) ([https://en.wikipedia.org/wiki/Predictive\\_Model\\_Markup\\_Language](https://en.wikipedia.org/wiki/Predictive_Model_Markup_Language)) or  
Portable Format for Analytics (PFA) (<http://dmg.org/pfa/>)
- <sup>377</sup> Cao Z, Chen P, Ma Z, Li S, Gao X, Wu RX, Pan L, Shi Y. *Near-Field Communication Sensors*. Sensors (Basel). 2019 September 12; 19(18):3947. doi: 10.3390/s19183947  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6767079/pdf/sensors-19-03947.pdf>
- <sup>378</sup> Zhang W, Davis BD, Chen SS, Sincuir Martinez JM, Plummer JT, Vail E. *Emergence of a Novel SARS-CoV-2 Variant in Southern California*. JAMA. 2021 February 11. doi: 10.1001/jama.2021.1612.  
<https://www.medrxiv.org/content/10.1101/2021.01.18.21249786v1.full.pdf>  
<https://jamanetwork.com/journals/jama/fullarticle/2776543>
- <sup>379</sup> White KM, Rosales R, Yildiz S, Kehrer T, Miorin L, Moreno E, Jangra S, Uccellini MB, Rathnasinghe R, Coughlan L, Martinez-Romero C, Batra J, Rojc A, Bouhaddou M, Fabius JM, Obernier K, Dejoze M, Guillén MJ, Losada A, Avilés P, Schotsaert M, Zwaka T, Vignuzzi M, Shokat KM, Krogan NJ, García-Sastre A. (2021) Plitidepsin has potent preclinical efficacy against SARS-CoV-2 by targeting the host protein eEF1A. *Science*. 2021 February 26; 371(6532):926-931. doi: 10.1126/science.abf4058.  
<https://science.sciencemag.org/content/371/6532/926/tab-pdf>
- <sup>380</sup> Wu F, Thomas PA, Kravets VG, Arola HO, Soikkeli M, Iljin K, Kim G, Kim M, Shin HS, Andreeva DV, Neumann C, Küllmer M, Turchanin A, De Fazio D, Balci O, Babenko V, Luo B, Goykhman I, Hofmann S, Ferrari AC, Novoselov KS, Grigorenko AN. *Layered material platform for surface plasmon resonance biosensing*. Science Reports. 2019 December 30; 9(1):20286. doi: 10.1038/s41598-019-56105-7.  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6937298/pdf/41598\\_2019\\_Article\\_56105.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6937298/pdf/41598_2019_Article_56105.pdf)
- <sup>381</sup> Ciesielski GL, Hytönen VP, Kaguni LS. *Bi-layer Interferometry: A Novel Method to Elucidate Protein-Protein and Protein-DNA Interactions in the Mitochondrial DNA Replisome*. Methods in Molecular Biology 2016; 1351:223-31. doi: 10.1007/978-1-4939-3040-1\_17.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4703100/pdf/nihms747232.pdf>
- <sup>382</sup> Lasia, Andrzej. *Electrochemical Impedance Spectroscopy and Its Applications*. Springer, 2014. ISBN-13: 978-1-4614-8932-0 <https://file.zhibsci.com/202005/978-1-4614-8933-7-cn20200527170447.pdf>
- <sup>383</sup> Sauer, Bob (1999) Review of Chemical Equilibrium. MIT Biology Course 7.51  
<https://ocw.mit.edu/courses/biology/7-51-graduate-biochemistry-fall-2001/lecture-notes/fa01lec07.pdf>



- <sup>384</sup> Hopper JTS, Ambrose S, Grant OC, Krumm SA, Allison TM, Degiacomi MT, Tully MD, Pritchard LK, Ozorowski G, Ward AB, Crispin M, Doores KJ, Woods RJ, Benesch JLP, Robinson CV, Struwe WB. *The Tetrameric Plant Lectin BanLec Neutralizes HIV through Bidentate Binding to Specific Viral Glycans*. Structure. 2017 May 2; 25(5):773-782.e5. doi: 10.1016/j.str.2017.03.015. Epub 2017 April 20. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5556678/pdf/nihms889536.pdf>
- <sup>385</sup> Hsu, KL., Mahal, L. A lectin microarray approach for the rapid analysis of bacterial glycans. *Nature Protocols* 1, 543–549 (2006). <https://doi.org/10.1038/nprot.2006.76>
- <sup>386</sup> Parashar A. *Aptamers in Therapeutics*. J of Clinical Diagnostic Research 2016 Jun; 10(6):BE01-6. doi: 10.7860/JCDR/2016/18712.7922. Epub 2016 June 1. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4963637/pdf/jcdr-10-BE01.pdf>
- <sup>387</sup> Bradner, James. (2020) Small Molecule Therapeutics. MIT Course 7.00 “CoVID-19, SARS-CoV-2 and the Pandemic”. <https://biology.mit.edu/undergraduate/current-students/subject-offerings/covid-19-sars-cov-2-and-the-pandemic/>
- <sup>388</sup> Zaks, Tal (2020) A Phase 3, Randomized, Stratified, Observer-Blind, Placebo-Controlled Study to Evaluate the Efficacy, Safety, and Immunogenicity of mRNA-1273 SARS-CoV-2 Vaccine in Adults Aged 18 Years and Older. Protocol Number mRNA-1273-P301. ModernaTX (20 August 2020) <https://www.modernatx.com/sites/default/files/mRNA-1273-P301-Protocol.pdf>
- <sup>389</sup> Ramilowski JA, Yip CW, Agrawal S, Chang JC, Ciani Y, Kulakovskiy IV, Mendez M, Ooi JLC, Ouyang JF, Parkinson N, Petri A, Roos L, Severin J, Yasuzawa K, Abugessaisa I, Akalin A, Antonov IV, Arner E, Bonetti A, Bono H, Borsari B, Brombacher F, Cameron CJ, Cannistraci CV, Cardenas R, Cardon M, Chang H, Dostie J, Ducoli L, Favorov A, Fort A, Garrido D, Gil N, Gimenez J, Guler R, Handoko L, Harshbarger J, Hasegawa A, Hasegawa Y, Hashimoto K, Hayatsu N, Heutink P, Hirose T, Imada EL, Itoh M, Kaczkowski B, Kanhere A, Kawabata E, Kawaji H, Kawashima T, Kelly ST, Kojima M, Kondo N, Koseki H, Kouno T, Kratz A, Kurowska-Stolarska M, Kwon ATJ, Leek J, Lennartsson A, Lizio M, López-Redondo F, Luginbühl J, Maeda S, Makeev VJ, Marchionni L, Medvedeva YA, Minoda A, Müller F, Muñoz-Aguirre M, Murata M, Nishiyori H, Nitta KR, Noguchi S, Noro Y, Nurtdinov R, Okazaki Y, Orlando V, Paquette D, Parr CJC, Rackham OJL, Rizzu P, Sánchez Martinez DF, Sandelin A, Sanjana P, Semple CAM, Shibayama Y, Sivaraman DM, Suzuki T, Szumowski SC, Tagami M, Taylor MS, Terao C, Thodberg M, Thongjuea S, Tripathi V, Ulitsky I, Verardo R, Vorontsov IE, Yamamoto C, Young RS, Baillie JK, Forrest ARR, Guigó R, Hoffman MM, Hon CC, Kasukawa T, Kauppinen S, Kere J, Lenhard B, Schneider C, Suzuki H, Yagi K, de Hoon MJL, Shin JW, Carninci P. *Functional annotation of human long noncoding RNAs via molecular phenotyping*. Genome Research 2020 July; 30(7):1060-1072. doi: 10.1101/gr.254219.119. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7397864/pdf/1060.pdf>
- <sup>390</sup> Immune Responses and Immunity to SARS-CoV-2. European Centre for Disease Prevention and Control (30 June 2020) <https://www.ecdc.europa.eu/en/covid-19/latest-evidence/immune-responses>
- <sup>391</sup> Eric J. Topol and Zeynep Tufekci (2020) The Remarkable Value of Thinking Broadly: A COVID-19 Trifecta.” *Medscape* (October 28, 2020) <http://www.medscape.com/viewarticle/938808>

- <sup>392</sup> Barrangou R, Fremaux C, Deveau H, Richards M, Boyaval P, Moineau S, Romero DA, Horvath P. *CRISPR provides acquired resistance against viruses in prokaryotes*. *Science*. 2007 March 23; 315(5819): pp. 1709-1712. doi: 10.1126/science. <https://science.sciencemag.org/content/315/5819/1709.long>
- <sup>393</sup> Doudna JA, Charpentier E. *Genome editing. The new frontier of genome engineering with CRISPR-Cas9*. *Science*. 2014 November 28; 346(6213):1258096. doi:10.1126/science.1258096 <https://science.sciencemag.org/content/346/6213/1258096.long>
- <sup>394</sup> Lander, Eric S. *et al* Initial sequencing and analysis of the human genome. *Nature*. 2001 February 15; 409(6822): pp. 860-921. doi: 10.1038/35057062. <https://www.nature.com/articles/35057062.pdf>
- <sup>395</sup> Venter, J. Craig *et al* The sequence of the human genome. *Science*. 2001 February 16; 291(5507): pp.1304-1351. doi: 10.1126/science. <https://science.sciencemag.org/content/sci/291/5507/1304.full.pdf>
- <sup>396</sup> Brown TA. *Genomes*. 2nd edition. Oxford: Wiley-Liss; 2002. Chapter 1, The Human Genome. <https://www.ncbi.nlm.nih.gov/books/NBK21134/>
- <sup>397</sup> National Academies of Sciences, Engineering, and Medicine; Health and Medicine Division; Board on Health Sciences Policy; Roundtable on Genomics and Precision Health; Forum on Drug Discovery, Development, and Translation. *Enabling Precision Medicine: The Role of Genetics in Clinical Drug Development: Proceedings of a Workshop*. Washington (DC): National Academies Press (US); 2017 July 10. 4, Integrating Genetics into the Drug Development Pathway for Complex Diseases. <https://www.ncbi.nlm.nih.gov/books/NBK458881/>
- <sup>398</sup> Rahem SM, Epsi NJ, Coffman FD, Mitrofanova A. *Genome-wide analysis of therapeutic response uncovers molecular pathways governing tamoxifen resistance in ER+ breast cancer*. *EBioMedicine*. 2020 November; 61:103047. doi: 10.1016/j.ebiom.2020.103047. Epub 2020 October 21. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7585053/pdf/main.pdf>
- <sup>399</sup> Grifoni A, Weiskopf D, Ramirez SI, Mateus J, Dan JM, Moderbacher CR, Rawlings SA, Sutherland A, Premkumar L, Jadi RS, Marrama D, de Silva AM, Frazier A, Carlin AF, Greenbaum JA, Peters B, Krammer F, Smith DM, Crotty S, Sette A. *Targets of T Cell Responses to SARS-CoV-2 Coronavirus in Humans with COVID-19 Disease and Unexposed Individuals*. *Cell*. 2020 June 25; 181(7):1489-1501.e15. doi: 10.1016/j.cell.2020.05.015. Epub 2020 May 20. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7237901/pdf/main.pdf>
- <sup>400</sup> García LF (2020) Immune Response, Inflammation, and the Clinical Spectrum of COVID-19. *Front. Immunol.* 11:1441. doi: 10.3389/fimmu.2020.01441 <https://www.frontiersin.org/articles/10.3389/fimmu.2020.01441/pdf>
- <sup>401</sup> World Health Organization. What we know about the COVID-19 immune response (02 Aug 2020) [www.who.int/docs/default-source/coronaviruse/risk-comms-updates/update-34-immunity-2nd.pdf](http://www.who.int/docs/default-source/coronaviruse/risk-comms-updates/update-34-immunity-2nd.pdf)
- <sup>402</sup> NIAID Strategic Plan Details COVID-19 Research Priorities (April 22, 2020) *Immune Response to COVID-19: Comprehensive analyses of innate and adaptive immune responses during acute COVID-19 infection and convalescence*. <https://www.niaid.nih.gov/research/immune-response-covid-19>
- <sup>403</sup> Braun J, Loyal L, Frentsch M, Wendisch D, Georg P, Kurth F, et al. *Presence of SARS-CoV-2 reactive T cells in COVID-19 patients and healthy donors*. medRxiv. 2020:2020.04.17.20061440. <https://www.medrxiv.org/content/10.1101/2020.04.17.20061440v1.full.pdf>

- <sup>404</sup> Karki, R., Sharma, B.R., Tuladhar, S., Williams, E.P., Zalduondo, L., Samir, P., Zheng, M., Sundaram, B., Banoth, B., Malireddi, R.K.S., Schreiner, P., Neale, G., Vogel, P., Webby, R., Jonsson, C.B., Kanneganti, T.-D. (2020) *Synergism of TNF- $\alpha$  and IFN- $\gamma$  triggers inflammatory cell death, tissue damage, and mortality in SARS-CoV-2 infection and cytokine shock syndromes*. Cell (13 November 2020) <https://doi.org/10.1016/j.cell.2020.11.025>. [https://www.cell.com/cell/pdf/S0092-8674\(20\)31542-7.pdf](https://www.cell.com/cell/pdf/S0092-8674(20)31542-7.pdf) <https://www.biorxiv.org/content/10.1101/2020.10.29.361048v3.full.pdf>
- <sup>405</sup> Arunachalam PS, Wimmers F, Mok CKP, Perera RAPM, Scott M, Hagan T, Sigal N, Feng Y, Bristow L, Tak-Yin Tsang O, Wagh D, Coller J, Pellegrini KL, Kazmin D, Alaaeddine G, Leung WS, Chan JMC, Chik TSH, Choi CYC, Huerta C, Paine McCullough M, Lv H, Anderson E, Edupuganti S, Upadhyay AA, Bosinger SE, Maecker HT, Khatri P, Roupael N, Peiris M, Pulendran B. *Systems biological assessment of immunity to mild versus severe COVID-19 infection in humans*. Science. 2020 September 4; 369(6508): pp. 1210-1220. doi: 10.1126/science.abc6261. Epub 2020 August 11/ [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7665312/pdf/369\\_1210.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7665312/pdf/369_1210.pdf)
- <sup>406</sup> Hadjadj J, Yatim N, Barnabei L, Corneau A, Boussier J, Smith N, Péré H, Charbit B, Bondet V, Chenevier-Gobeaux C, Breillat P, Carlier N, Gauzit R, Morbieu C, Pène F, Marin N, Roche N, Szwebel TA, Merkling SH, Treluyer JM, Veyer D, Mouthon L, Blanc C, Tharaux PL, Rozenberg F, Fischer A, Duffy D, Rieux-Laucat F, Kernéis S, Terrier B. *Impaired type I interferon activity and inflammatory responses in severe COVID-19 patients*. Science. 2020 Aug 7; 369(6504): pp. 718-724. doi: 10.1126/science.abc6027. Epub 2020 July 13. <https://science.sciencemag.org/content/369/6504/718> [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7402632/pdf/369\\_718.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7402632/pdf/369_718.pdf)
- <sup>407</sup> Shrock E, Fujimura E, Kula T, Timms RT, Lee IH, Leng Y, Robinson ML, Sie BM, Li MZ, Chen Y, Logue J, Zuiani A, McCulloch D, Lelis FJN, Henson S, Monaco DR, Travers M, Habibi S, Clarke WA, Caturegli P, Laeyendecker O, Piechocka-Trocha A, Li JZ, Khatri A, Chu HY; MGH COVID-19 Collection & Processing Team, Villani AC, Kays K, Goldberg MB, Hacohen N, Filbin MR, Yu XG, Walker BD, Wesemann DR, Larman HB, Lederer JA, Elledge SJ. *Viral epitope profiling of COVID-19 patients reveals cross-reactivity and correlates of severity*. Science. 2020 November 27; 370(6520):eabd4250. doi: 10.1126/science.abd4250. Epub 2020 September 29. <https://science.sciencemag.org/content/370/6520/eabd4250/tab-pdf>
- <sup>408</sup> Struhl K. *Naturally occurring poly(dA-dT) sequences are upstream promoter elements for constitutive transcription in yeast*. Proc Natl Acad Sci. 1985 Dec; 82(24): pp. 8419-23. doi: 10.1073/pnas.82.24.8419 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC390927/pdf/pnas00364-0155.pdf>
- <sup>409</sup> Hasegawa Y, Struhl K. *Promoter-specific dynamics of TATA-binding protein association with the human genome*. Genome Res. 2019 Dec; 29(12): pp. 1939-1950. doi: 10.1101/gr.254466.119. Epub 2019 November 15. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6886507/pdf/1939.pdf>
- <sup>410</sup> Thompson C, Grayson N, Paton R, Lourenço J, Penman B, Lee LN, et al. *Neutralising antibodies to SARS coronavirus 2 in Scottish blood donors - a pilot study of the value of serology to determine population exposure*. medRxiv 2020. <https://www.medrxiv.org/content/10.1101/2020.04.13.20060467v2.full.pdf>

<sup>411</sup> Schluter J, Peled JU, Taylor BP, Markey KA, Smith M, Taur Y, Niehus R, Staffas A, Dai A, Fontana E, Amoretti LA, Wright RJ, Morjaria S, Fenelus M, Pessin MS, Chao NJ, Lew M, Bohannon L, Bush A, Sung AD, Hohl TM, Perales MA, van den Brink MRM, Xavier JB. *The gut microbiota is associated with immune cell dynamics in humans*. Nature. 2020 November 25. doi: 10.1038/s41586-020-2971-8.

<https://www.nature.com/articles/s41586-020-2971-8> and pre-prints:

<https://www.biorxiv.org/content/10.1101/618256v3.full.pdf>

<https://www.biorxiv.org/content/10.1101/618256v1.full.pdf>

<sup>412</sup> Tonegawa S, Steinberg C, Dube S, Bernardini A. *Evidence for somatic generation of antibody diversity*. Proc Natl Acad Sci U S A. 1974 Oct; 71(10): pp. 4027-4031. doi: 10.1073/pnas.71.10.4027.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC434321/pdf/pnas00073-0237.pdf>

[http://tonegawalab.mit.edu/wp-content/uploads/2014/06/087\\_BiosciRept\\_Tonegawa\\_1988.pdf](http://tonegawalab.mit.edu/wp-content/uploads/2014/06/087_BiosciRept_Tonegawa_1988.pdf)

<sup>413</sup> Janeway CA Jr, Travers P, Walport M, et al. *Immunobiology: The Immune System in Health and Disease*. 5th edition, 2001. The generation of diversity in immunoglobulins.

<https://www.ncbi.nlm.nih.gov/books/NBK27140/>

<sup>414</sup> Gordon DE, Jang GM, Bouhaddou M, Xu J, Obernier K, White KM, O'Meara MJ, Rezelj VV, Guo JZ, Swaney DL, Tummino TA, Hüttenhain R, Kaake RM, Richards AL, Tutuncuoglu B, Foussard H, Batra J, Haas K, Modak M, Kim M, Haas P, Polacco BJ, Braberg H, Fabius JM, Eckhardt M, Soucheray M, Bennett MJ, Cakir M, McGregor MJ, Li Q, Meyer B, Roesch F, Vallet T, Mac Kain A, Miorin L, Moreno E, Naing ZZC, Zhou Y, Peng S, Shi Y, Zhang Z, Shen W, Kirby IT, Melnyk JE, Chorba JS, Lou K, Dai SA, Barrio-Hernandez I, Memon D, Hernandez-Armenta C, Lyu J, Mathy CJP, Perica T, Pilla KB, Ganesan SJ, Saltzberg DJ, Rakesh R, Liu X, Rosenthal SB, Calviello L, Venkataramanan S, Liboy-Lugo J, Lin Y, Huang XP, Liu Y, Wankowicz SA, Bohn M, Safari M, Ugur FS, Koh C, Savar NS, Tran QD, Shengjuler D, Fletcher SJ, O'Neal MC, Cai Y, Chang JCJ, Broadhurst DJ, Klippsten S, Sharp PP, Wenzell NA, Kuzuoglu-Ozturk D, Wang HY, Trenker R, Young JM, Cavero DA, Hiatt J, Roth TL, Rathore U, Subramanian A, Noack J, Hubert M, Stroud RM, Frankel AD, Rosenberg OS, Verba KA, Agard DA, Ott M, Emerman M, Jura N, von Zastrow M, Verdin E, Ashworth A, Schwartz O, d'Enfert C, Mukherjee S, Jacobson M, Malik HS, Fujimori DG, Ideker T, Craik CS, Floor SN, Fraser JS, Gross JD, Sali A, Roth BL, Ruggero D, Taunton J, Kortemme T, Beltrao P, Vignuzzi M, García-Sastre A, Shokat KM, Shoichet BK, Krogan NJ. *A SARS-CoV-2 protein interaction map reveals targets for drug repurposing*. Nature. 2020 July; 583(7816): pp. 459-468. doi: 10.1038/s41586-020-2286-9. Epub 2020 April 30.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7431030/pdf/nihms-1587111.pdf>

<sup>415</sup> Zhang JR, Hardham JM, Barbour AG, Norris SJ. *Antigenic variation in Lyme disease borreliae by promiscuous recombination of VMP-like sequence cassettes*. Cell. 1997 April 18; 89(2): 275-85.

doi: 10.1016/s0092-8674(00)80206-8. Erratum in: Cell 1999 February 5; 96(3): pp. 447

<https://www.cell.com/action/showPdf?pii=S0092-8674%2800%2980206-8>

<sup>416</sup> McClintock B (1950) The origin and behavior of mutable loci in maize. Proc Natl Acad Sci USA 36(6): 344–355. <https://www.pnas.org/content/pnas/36/6/344.full.pdf>

<sup>417</sup> “The Nobel Prize in Physiology or Medicine 1983.” *NobelPrize.Org*

<https://www.nobelprize.org/prizes/medicine/1983/mcclintock/facts/>

---

<sup>418</sup> “Happy Birthday, Barbara McClintock.” *Wired*.

<https://www.wired.com/2012/06/happy-birthday-barbara-mcclintock/>

<sup>419</sup> Bean, W., Cox, N. and Kendal, A. *Recombination of human influenza A viruses in nature*.

*Nature* **284**, 638–640 (1980) <https://doi.org/10.1038/284638a0>

<sup>420</sup> Pays E, Van Assel S, Laurent M, Darville M, Vervoort T, Van Meirvenne N, Steinert M. *Gene conversion as a mechanism for antigenic variation in trypanosomes*. *Cell*. 1983 Sep; 34(2): pp.371-381  
doi: 10.1016/0092-8674(83)90371-9. [https://www.cell.com/cell/pdf/0092-8674\(83\)90371-9.pdf](https://www.cell.com/cell/pdf/0092-8674(83)90371-9.pdf)

<sup>421</sup> Scherf A, Hernandez-Rivas R, Buffet P, Bottius E, Benatar C, Pouvelle B, Gysin J, Lanzer M. *Antigenic variation in malaria: in situ switching, relaxed and mutually exclusive transcription of var genes during intra-erythrocytic development in Plasmodium falciparum*. *EMBO J*. 1998 Sep 15; 17(18): pp. 5418-5426.  
doi: 10.1093/emboj/17.18.5418

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1170867/pdf/005418.pdf>

<sup>422</sup> Eisen HN and Siskind GW. (1964) *Variations in Affinities of Antibodies During Immune Response*. *Biochemistry*. 1964 July; 3:996-1008. doi: 10.1021/bi00895a027

<sup>423</sup> Barnes CO, West AP Jr, Huey-Tubman KE, Hoffmann MAG, Sharaf NG, Hoffman PR, Koranda N, Gristick HB, Gaebler C, Muecksch F, Lorenzi JCC, Finkin S, Hägglöf T, Hurley A, Millard KG, Weisblum Y, Schmidt F, Hatziioannou T, Bieniasz PD, Caskey M, Robbiani DF, Nussenzweig MC, Bjorkman PJ. *Structures of Human Antibodies Bound to SARS-CoV-2 Spike Reveal Common Epitopes and Recurrent Features of Antibodies*. *Cell*. 2020 Aug 20; 182(4):828-842.e16. doi: 10.1016/j.cell.2020.06.025.  
Epub 2020 June 24. <https://www.cell.com/action/showPdf?pii=S0092-8674%2820%2930757-1>  
and pre-print <https://www.biorxiv.org/content/10.1101/2020.05.28.121533v1.full.pdf>

<sup>424</sup> Barnes CO, Jette CA, Abernathy ME, Dam KA, Esswein SR, Gristick HB, Malyutin AG, Sharaf NG, Huey-Tubman KE, Lee YE, Robbiani DF, Nussenzweig MC, West AP Jr, Bjorkman PJ. *SARS-CoV-2 neutralizing antibody structures inform therapeutic strategies*. *Nature*. 2020 October 12.  
doi: 10.1038/s41586-020-2852-1. <https://www.nature.com/articles/s41586-020-2852-1.pdf>

<sup>425</sup> Starr TN, Greaney AJ, Addetia A, Hannon WW, Choudhary MC, Dingens AS, Li JZ, Bloom JD. *Prospective mapping of viral mutations that escape antibodies used to treat COVID-19*. *bioRxiv* [Preprint]. 2020 December 1. doi: 10.1101/2020.11.30.405472. Update: *Science* Epub 2021 Jan 25. Published in *Science* 19 February 2021: Vol. 371, Issue 6531, pp. 850-854 DOI: 10.1126/science.abf9302  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7724661/pdf/nihpp-2020.11.30.405472.pdf>

<sup>426</sup> Rappazzo CG, Tse LV, Kaku CI, Wrapp D, Sakharkar M, Huang D, Deveau LM, Yockachonis TJ, Herbert AS, Battles MB, O'Brien CM, Brown ME, Geoghegan JC, Belk J, Peng L, Yang L, Hou Y, Scobey TD, Burton DR, Nemazee D, Dye JM, Voss JE, Gunn BM, McLellan JS, Baric RS, Gralinski LE, Walker LM. *Broad and potent activity against SARS-like viruses by an engineered human monoclonal antibody*. *Science*. 2021 February 19; 371(6531):823-829. doi: 10.1126/science.abf4830. Epub 2021 January 25.  
<https://science.sciencemag.org/content/371/6531/823/tab-pdf>

<sup>427</sup> Khailany RA, Safdar M, Ozaslan M. *Genomic characterization of a novel SARS-CoV-2*. *Gene Rep*. 2020 June; 19:100682. doi: 10.1016/j.genrep.2020.100682. Epub 2020 April 16.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7161481/pdf/main.pdf>



- <sup>428</sup> Bar-On YM, Flamholz A, Phillips R, Milo R. SARS-CoV-2 (COVID-19) by the numbers. *Elife*. April 2, 2020. 9:e57309. doi: 10.7554/eLife.57309  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7224694/pdf/elife-57309.pdf>
- <sup>429</sup> Margaux M. Pinney, Daniel A. Mokhtari, Eyal Akiva, Filip Yabukarski, David M. Sanchez, Ruibin Liang, Tzanko Doukov, Todd J. Martinez, Patricia C. Babbitt and Daniel Herschlag (2021) Parallel molecular mechanisms for enzyme temperature adaptation. *Science*. 5 March 2021. Vol. 371, Issue 6533. DOI: 10.1126/science.aay2784 <https://science.sciencemag.org/content/371/6533/eaay2784>
- <sup>430</sup> Oliphant, A. R. and Struhl, K. *An efficient method for generating proteins with altered enzymatic properties: application to beta-lactamase*. Proc Natl Acad Sci U S A. 1989 December; 86(23): pp. 9094-9098. doi: 10.1073/pnas.86.23.9094. Erratum in: Proc Natl Acad Sci 1992 May 15; 89(10): pp. 4779. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC298440/pdf/pnas00290-0052.pdf>
- <sup>431</sup> **B. N. Fields** and **Karen Byers** (1983) “The Genetic Basis of Viral Virulence.” *Philosophical Transactions of the Royal Society of London. B, Biological Sciences*, vol. 303, no. 1114, 15 September 1983, pp. 209–218. doi:10.1098/rstb.1983.0094  
<https://royalsocietypublishing.org/doi/pdf/10.1098/rstb.1983.0094>
- <sup>432</sup> Yanofsky, Charles. “Establishing the Triplet Nature of the Genetic Code.” *Cell*, vol. 128, no. 5, March 2007, pp. 815-818. doi:10.1016/j.cell.2007.02.029.  
<https://www.cell.com/action/showPdf?pii=S0092-8674%2807%2900253-X>
- <sup>433</sup> Matthaei JH, Jones OW, Martin RgG and Nirenberg MW. *Characteristics and composition of RNA coding units*. Proc Natl Acad Sci U S A. 1962 April 15; 48(4):666-77. doi: 10.1073/pnas.48.4.666.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC220831/pdf/pnas00227-0188.pdf>
- <sup>434</sup> <https://www.genome.gov/genetics-glossary/Genetic-Code>
- <sup>435</sup> Ramachandran GN, Ramakrishnan C and Sasisekharan V. *Stereochemistry of polypeptide chain configurations*. J Mol Biol. 1963 July; 7:95-99. doi: 10.1016/s0022-2836(63)80023-6.
- <sup>436</sup> Tapscott SJ. The circuitry of a master switch: MyoD and the regulation of skeletal muscle gene transcription. *Development*. 2005 June; 132(12):268526-95. doi: 10.1242/dev.01874.  
<https://dev.biologists.org/content/develop/132/12/2685.full.pdf>
- <sup>437</sup> Kang, Hyuckjoon, et al. “Bivalent Complexes of PRC1 with Orthologs of BRD4 and MOZ/MORF Target Developmental Genes in *Drosophila*.” *Genes & Dev*, vol. 31, no. 19, Oct. 2017, pp. 1988–2002 doi:10.1101/gad.305987.117
- <sup>438</sup> Andrew Wilber, Arthur W. Nienhuis, Derek A. Persons; Transcriptional regulation of fetal to adult hemoglobin switching: new therapeutic opportunities. *Blood* 2011; 117 (15): 3945–3953.  
<https://doi.org/10.1182/blood-2010-11-316893>  
<https://ashpublications.org/blood/article-pdf/117/15/3945/1461649/zh801511003945.pdf>
- <sup>439</sup> Cho, JH., Tsai, MJ. The role of *BETA2/NeuroD1* in the development of the nervous system. *Mol Neurobiol* 30, 35–47 (2004). <https://doi.org/10.1385/MN:30:1:035>

- <sup>440</sup> Chen YC, Ma NX, Pei ZF, Wu Z, Do-Monte FH, Keefe S, Yellin E, Chen MS, Yin JC, Lee G, Minier-Toribio A, Hu Y, Bai YT, Lee K, Quirk GJ, Chen G. *A NeuroD1 AAV-Based Gene Therapy for Functional Brain Repair after Ischemic Injury through In Vivo Astrocyte-to-Neuron Conversion*. *Molecular Therapy* 2020 January 8; 28(1):217-234. doi: 10.1016/j.ymthe.2019.09.003. Epub 2019 September 6. [www.cell.com/molecular-therapy-family/molecular-therapy/pdfExtended/S1525-0016\(19\)30404-6](http://www.cell.com/molecular-therapy-family/molecular-therapy/pdfExtended/S1525-0016(19)30404-6)
- <sup>441</sup> Deshmukh R, Lakhe D, Kunte K. (2020) *Tissue-specific developmental regulation and isoform usage underlie the role of doublesex in sex differentiation and mimicry in Papilio swallowtails*. *Royal Society Open Science* 7: 200792. <http://dx.doi.org/10.1098/rsos.200792><https://royalsocietypublishing.org/doi/pdf/10.1098/rsos.200792>
- <sup>442</sup> Zhang Z, Zhan X. *GNSS Spoofing Network Monitoring Based on Differential Pseudorange*. *Sensors* (Basel). 2016 October 23; 16(10):1771. doi: 10.3390/s16101771. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5087555/pdf/sensors-16-01771.pdf>
- <sup>443</sup> Kalsotra A, Cooper TA. *Functional consequences of developmentally regulated alternative splicing*. *Nature Reviews Genetics* 2011 September 16; 12(10):715-29. doi: 10.1038/nrg3052. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3321218/pdf/nihms-366480.pdf>
- <sup>444</sup> Weyn-Vanhenhenryck, S.M., Feng, H., Ustianenko, D. *et al.* Precise temporal regulation of alternative splicing during neural development. *Nature Communications* 9 2189 (2018). <https://doi.org/10.1038/s41467-018-04559-0> <https://www.nature.com/articles/s41467-018-04559-0.pdf>
- <sup>445</sup> <https://www.fda.gov/media/137564/download>
- <sup>446</sup> Carey, Benedict. "W.H.O. Rejects Antiviral Drug Remdesivir as a Covid Treatment." *The New York Times*, 20 November 2020. <https://www.nytimes.com/2020/11/19/health/remdesivir-covid-19.html>
- <sup>447</sup> Linsky TW, Vergara R, Codina N, Nelson JW, Walker MJ, Su W, Hsiang TY, Esser-Nobis K, Yu K, Hou YJ, Priya T, Mitsumoto M, Pong A, Lau UY, Mason ML, Chen J, Chen A, Berrocal T, Peng H, Clairmont NS, Castellanos J, Lin YR, Josephson-Day A, Baric R, Walkey CD, Swanson R, Gale M, Blancas-Mejia LM, Yen HL, Silva DA. *De novo design of ACE2 protein decoys to neutralize SARS-CoV-2*. *bioRxiv* [Preprint]. 2020 August 3; 2020.08.03.231340. doi: 10.1101/2020.08.03.231340. Update in: *Science*. 2020 November 5. <https://www.biorxiv.org/content/10.1101/2020.08.03.231340v1.full.pdf>
- <sup>448</sup> Cha L, Berry CM, Nolan D, Castley A, Fernandez S, French MA. *Interferon-alpha, immune activation and immune dysfunction in treated HIV infection*. *Clin Translational Immunology*. 2014 Feb 28;3(2):e10. doi: 10.1038/cti.2014.1. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4232062/pdf/cti20141a.pdf>
- <sup>449</sup> Dinarello CA, Novick D, Kim S, Kaplanski G. *Interleukin-18 and IL-18 binding protein*. *Frontiers in Immunology* 2013 October 8; 4:289. doi: 10.3389/fimmu.2013.00289. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3792554/pdf/fimmu-04-00289.pdf>
- <sup>450</sup> Iyer SS, Cheng G. *Role of interleukin 10 transcriptional regulation in inflammation and autoimmune disease*. *Critical Reviews in Immunology* 2012; 32(1):23-63. doi: 10.1615/critrevimmunol.v32.i1.30. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3410706/pdf/nihms377104.pdf>
- <sup>451</sup> Park A and Iwasaki A. *Type I and Type III Interferons - Induction, Signaling, Evasion, and Application to Combat COVID-19*. *Cell Host Microbe*. 2020 June 10; 27(6):870-878. doi: 10.1016/j.chom.2020.05.008 Epub 2020 May 27. [https://www.cell.com/cell-host-microbe/pdf/S1931-3128\(20\)30290-0.pdf](https://www.cell.com/cell-host-microbe/pdf/S1931-3128(20)30290-0.pdf)

- <sup>452</sup> Kiyoshi Takeda and Shizuo Akira. Toll-like receptors in innate immunity. *International Immunology*, Volume 17, Issue 1, January 2005, Pages 1–14, <https://doi.org/10.1093/intimm/dxh186>
- <sup>453</sup> Kawai T and Akira S. *The roles of TLRs, RLRs and NLRs in pathogen recognition*. *International Immunology* 2009 April; 21(4):317-37. doi: 10.1093/intimm/dxp017. Epub 2009 February 26. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2721684/pdf/dxp017.pdf>
- <sup>454</sup> Baltimore D. *Discovering NF-kappaB*. Cold Spring Harbor Perspectives in Biol. 2009 July; 1(1):a000026. doi: 10.1101/cshperspect.a000026. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2742082/pdf/cshperspect-NFK-a000026.pdf>
- <sup>455</sup> Sen, Ganes, C. *Viruses and interferons*. *Annual Review of Microbiology* 2001; 55: 255-281. doi: 10.1146/annurev.micro.55.1.255 <https://www.annualreviews.org/doi/pdf/10.1146/annurev.micro.55.1.255>
- <sup>456</sup> Lee, J.S., Shin, EC. The type I interferon response in COVID-19: implications for treatment. *Nature Reviews Immunology* 20, 585–586 (2020). <https://doi.org/10.1038/s41577-020-00429-3> <https://www.nature.com/articles/s41577-020-00429-3.pdf>
- <sup>457</sup> Patrick, Deval. *A Reason to Believe: Lessons from an Improbable Life*. 1st ed, Broadway Books, 2011.
- <sup>458</sup> Corbett, Kizzmekia. (2020) *Vaccines*. MIT Course 7.00 “CoVID-19, SARS-CoV-2 and the Pandemic”. <https://biology.mit.edu/undergraduate/current-students/subject-offerings/covid-19-sars-cov-2-and-the-pandemic/>
- <sup>459</sup> Wajnberg A, Amanat F, Firpo A, Altman DR, Bailey MJ, Mansour M, McMahon M, Meade P, Mendu DR, Muellers K, Stadlbauer D, Stone K, Strohmeier S, Simon V, Aberg J, Reich DL, Krammer F, Cordon-Cardo C. *Robust neutralizing antibodies to SARS-CoV-2 infection persist for months*. *Science*. 2020 Oct 28 doi: 10.1126/science.abd7728. <https://science.sciencemag.org/content/370/6521/1227.full.pdf>
- <sup>460</sup> Atkeson, Andrew, et al. *Economic Benefits of COVID-19 Screening Tests*. w28031, NBER. Oct. 2020. doi:10.3386/w28031. [https://www.nber.org/system/files/working\\_papers/w28031/w28031.pdf](https://www.nber.org/system/files/working_papers/w28031/w28031.pdf)
- <sup>461</sup> Biden, Joe and Harris, Kamala (2020) *Build Back Better: Joe Biden’s Jobs Aand Economic Recovery Plan for Working Families*. <https://joebiden.com/build-back-better/#>
- <sup>462</sup> Deaton, Angus. *Health in an Age of Globalization*. w10669, National Bureau of Economic Research, August 2004. doi:10.3386/w10669. [www.nber.org/system/files/working\\_papers/w10669/w10669.pdf](http://www.nber.org/system/files/working_papers/w10669/w10669.pdf)
- <sup>463</sup> One Health. Center for Disease Control and Prevention. <https://www.cdc.gov/onehealth/index.html>
- <sup>464</sup> Gordon DE, Hiatt J, Bouhaddou M, Rezelj VV, Ulferts S, Braberg H, Jureka AS, Obernier K, Guo JZ, Batra J, Kaake RM, Weckstein AR, Owens TW, Gupta M, Pourmal S, Titus EW, Cakir M, Soucheray M, McGregor M, Cakir Z, Jang G, O'Meara MJ, Tummino TA, Zhang Z, Foussard H, Rojc A, Zhou Y, Kuchenov D, Hüttenhain R, Xu J, Eckhardt M, Swaney DL, Fabius JM, Ummadi M, Tutuncuoglu B, Rathore U, Modak M, Haas P, Haas KM, Naing ZZC, Pulido EH, Shi Y, Barrio-Hernandez I, Memon D, Petsalaki E, Dunham A, Marrero MC, Burke D, Koh C, Vallet T, Silvas JA, Azumaya CM, Billesbølle C, Brilot AF, Campbell MG, Diallo A, Dickinson MS, Diwanji D, Herrera N, Hoppe N, Kratochvil HT, Liu Y, Merz GE, Moritz M, Nguyen HC, Nowotny C, Puchades C, Rizo AN, Schulze-Gahmen U, Smith AM, Sun M, Young ID, Zhao J, Asarnow D, Biel J, Bowen A, Braxton JR, Chen J, Chio CM, Chio US, Deshpande I, Doan L, Faust B, Flores S, Jin M, Kim K, Lam VL, Li F, Li J, Li YL, Li Y, Liu X, Lo M, Lopez

---

KE, Melo AA, Moss FR 3rd, Nguyen P, Paulino J, Pawar KI, Peters JK, Pospiech TH Jr, Safari M, Sangwan S, Schaefer K, Thomas PV, Thwin AC, Trenker R, Tse E, Tsui TKM, Wang F, Whitis N, Yu Z, Zhang K, Zhang Y, Zhou F, Saltzberg D; QCRG Structural Biology Consortium, Hodder AJ, Shun-Shion AS, Williams DM, White KM, Rosales R, Kehrer T, Miorin L, Moreno E, Patel AH, Rihn S, Khalid MM, Vallejo-Gracia A, Fozouni P, Simoneau CR, Roth TL, Wu D, Karim MA, Ghousaini M, Dunham I, Berardi F, Weigang S, Chazal M, Park J, Logue J, McGrath M, Weston S, Haupt R, Hastie CJ, Elliott M, Brown F, Burness KA, Reid E, Dorward M, Johnson C, Wilkinson SG, Geyer A, Giesel DM, Baillie C, Raggett S, Leech H, Toth R, Goodman N, Keough KC, Lind AL; Zoonomia Consortium, Klesh RJ, Hemphill KR, Carlson-Stevermer J, Oki J, Holden K, Maures T, Pollard KS, Sali A, Agard DA, Cheng Y, Fraser JS, Frost A, Jura N, Kortemme T, Manglik A, Southworth DR, Stroud RM, Alessi DR, Davies P, Frieman MB, Ideker T, Abate C, Jouvenet N, Kochs G, Shoichet B, Ott M, Palmarini M, Shokat KM, García-Sastre A, Rassen JA, Grosse R, Rosenberg OS, Verba KA, Basler CF, Vignuzzi M, Peden AA, Beltrao P, Krogan NJ. *Comparative host-coronavirus protein interaction networks reveal pan-viral disease mechanisms*. *Science*. 2020 October 15:eabe9403. doi: 10.1126/science.abe9403. Epub ahead of print. Update in: *Science* 04 Dec 2020: Vol. 370, Issue 6521, eabe9403 DOI: 10.1126/science.abe9403 <https://science.sciencemag.org/content/sci/370/6521/eabe9403.full.pdf>

<sup>465</sup> Lalmuanawma S, Hussain J, Chhakchhuak L. *Applications of machine learning and artificial intelligence for Covid-19 (SARS-CoV-2) pandemic: A review*. *Chaos Solitons Fractals*. 2020 Oct; 139:110059. doi: 10.1016/j.chaos.2020.110059. Epub 2020 June 25. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7315944/pdf/main.pdf>

<sup>466</sup> Whitelaw, Sera, et al. “Applications of Digital Technology in COVID-19 Pandemic Planning and Response.” *The Lancet Digital Health*, vol. 2, no. 8, August 2020, pp. e435–40. doi:10.1016/S2589-7500(20)30142-4 <https://www.thelancet.com/action/showPdf?pii=S2589-7500%2820%2930142-4>

<sup>467</sup> J. Laguarda, F. Hueto and B. Subirana, "COVID-19 Artificial Intelligence Diagnosis using only Cough Recordings," in *IEEE Open J of Engineering in Medicine and Biology*, doi: 10.1109/OJEMB.2020.3026928. <https://ieeexplore.ieee.org/stamp/stamp.jsp?tp=&arnumber=9208795>

<sup>468</sup> Yglesias, Matthew. “‘No Malarkey,’ Joe Biden’s Unabashedly Lame New Slogan, Explained.” *Vox*, 3 December 2019. <https://www.vox.com/policy-and-politics/2019/12/3/20991841/joe-biden-no-malarkey>

<sup>469</sup> <https://coronavirus.jhu.edu/map.html>

<sup>470</sup> Pfeifer, Alexander and Verma, Inder M. *Gene therapy: promises and problems*. *Annual Review of Genomics and Human Genetics* 2001 2:1, 177-211 doi: 10.1146/annurev.genom.2.1.177 <https://www.annualreviews.org/doi/pdf/10.1146/annurev.genom.2.1.177>

<sup>471</sup> Karikó K, Buckstein M, Ni H, and Weissman D. (2005) *Suppression of RNA recognition by Toll-like receptors: the impact of nucleoside modification and the evolutionary origin of RNA*. *Immunity*. 2005 August; 23(2):165-75. doi: 10.1016/j.immuni.2005.06.008 <https://www.cell.com/action/showPdf?pii=S1074-7613%2805%2900211-6>

- <sup>472</sup> Haque, SK Manirul, *et al.* “A Comprehensive Review about SARS-CoV-2.” *Future Virology*, vol. 15, no. 9, September 2020, pp. 625–48. doi:10.2217/fvl-2020-0124  
<https://www.futuremedicine.com/doi/pdfplus/10.2217/fvl-2020-0124>
- <sup>473</sup> Sahin U, Türeci Ö. *Personalized vaccines for cancer immunotherapy*. *Science*. 2018 Marcg 23; 359(6382):1355-1360. doi: 10.1126/science.aar7112  
<https://science.sciencemag.org/content/359/6382/1355/tab-pdf>
- <sup>474</sup> “3 Questions: Phillip Sharp on the Discoveries That Enabled RNA Vaccines for Covid-19.” *MIT News Massachusetts Institute of Technology*. <https://news.mit.edu/2020/phillip-sharp-rna-vaccines-1211>
- <sup>475</sup> Sahin, U., Karikó, K. & Türeci, Ö. (2014) mRNA-based therapeutics - developing a new class of drugs. *Nature Reviews Drug Discovery* 13, 759–780 (2014). <https://doi.org/10.1038/nrd4278>  
<https://www.nature.com/articles/nrd4278.pdf>
- <sup>476</sup> Wise, Jacqui. “Covid-19: New Coronavirus Variant Is Identified in UK.” *BMJ*, vol. 371, 16 December 2020, p. m4857. doi:10.1136/bmj.m4857.
- <sup>477</sup> Hodcroft EB, Zuber M, Nadeau S, Crawford KHD, Bloom JD, Veessler D, Vaughan TG, Comas I, Candelas FG, Stadler T, and Neher RA. (2020) *Emergence and spread of a SARS-CoV-2 variant through Europe in the summer of 2020*. medRxiv [Preprint]. November 27, 2020  
doi: 10.1101/2020.10.25.20219063  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7709189/pdf/nihpp-2020.10.25.20219063.pdf>
- <sup>478</sup> Yongping Ma and Changyin Fang (2020) Deciphering the mystery of influenza virus A antigenic drift and immune escape. bioRxiv 2020.12.23.424101; doi: <https://doi.org/10.1101/2020.12.23.424101>  
<https://www.biorxiv.org/content/10.1101/2020.12.23.424101v1.full.pdf>
- <sup>479</sup> Kim H, Webster RG, Webby RJ. *Influenza Virus: Dealing with a Drifting and Shifting Pathogen*. *Viral Immunol*. 2018 March; 31(2):174-183. doi: 10.1089/vim.2017.0141. Epub 2018 January 26.
- <sup>480</sup> Kolata, Gina Bari. *Flu: The Story of the Great Influenza Pandemic of 1918 and the Search for the Virus That Caused It*. Atria Books, 2001. ISBN-13 : 978-0743203982
- <sup>481</sup> Matthew J. Memoli, Brett W. Jagger, Vivien G. Dugan, Li Qi, Jadon P. Jackson, Jeffery K. Taubenberger. Recent Human Influenza A/H3N2 Virus Evolution Driven by Novel Selection Factors in Addition to Antigenic Drift. *The Journal of Infectious Diseases*, Volume 200, Issue 8, 1 October 2009, pages 1232–1241. <https://doi.org/10.1086/605893>
- <sup>482</sup> Carrat F and Flahault A. *Influenza vaccine: the challenge of antigenic drift*. *Vaccine*. 2007 Sep 28; 25(39-40):6852-62. doi: 10.1016/j.vaccine.2007.07.027. Epub 2007 August 3.  
<https://doi.org/10.1016/j.vaccine.2007.07.027>  
<http://www.sciencedirect.com/science/article/pii/S0264410X07008328>
- <sup>483</sup> Kalita P, Padhi AK, Zhang KYJ, Tripathi T. *Design of a peptide-based subunit vaccine against novel coronavirus SARS-CoV-2*. *Microb Pathog*. 2020 Aug; 145:104236. doi: 10.1016/j.micpath.2020.104236.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7196559/pdf/main.pdf>
- <sup>484</sup> Dai L, Gao GF. *Viral targets for vaccines against COVID-19*. *Nat Rev Immunol*. 2020 Dec 18:1–10. doi: 10.1038/s41577-020-00480-0.  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7747004/pdf/41577\\_2020\\_Article\\_480.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7747004/pdf/41577_2020_Article_480.pdf)



- <sup>485</sup> Gold, Larry *et al* (2019) *Method for Generating Aptamers with Improved Off-Rates* (2019) <https://patents.justia.com/patent/20190330634>
- <sup>486</sup> Ng EW, Shima DT, Calias P, Cunningham ET Jr, Guyer DR, and Adamis AP. *Pegaptanib, a targeted anti-VEGF aptamer for ocular vascular disease*. *Nature Reviews Drug Discovery* 2006 Feb; 5(2):123-32. doi: 10.1038/nrd1955
- <sup>487</sup> Collected papers related to progress in aptamers with respect to stringency of binding assays (kinetics) and potential for use as therapeutics. <https://bit.ly/APTAMER-PROGRESS>
- <sup>488</sup> Day T, Gandon S, Lion S, Otto SP. *On the evolutionary epidemiology of SARS-CoV-2*. *Current Biology* 2020 August 3; 30(15):R849-R857. doi: 10.1016/j.cub.2020.06.031. Epub 2020 June 11. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7287426/pdf/main.pdf>
- <sup>489</sup> Minskaia E, Hertzog T, Gorbalenya AE, Campanacci V, Cambillau C, Canard B, Ziebuhr J. *Discovery of an RNA virus 3'->5' exoribonuclease that is critically involved in coronavirus RNA synthesis*. *Proc Natl Acad Sci U S A*. 2006 March 28; 103(13):5108-13. doi: 10.1073/pnas.0508200103. Epub 2006 March 20. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1458802/pdf/zpq5108.pdf>
- <sup>490</sup> Pachetti M, Marini B, Benedetti F, Giudici F, Mauro E, Storici P, Masciovecchio C, Angeletti S, Ciccozzi M, Gallo RC, Zella D, Ippodrino R. *Emerging SARS-CoV-2 mutation hot spots include a novel RNA-dependent-RNA polymerase variant*. *Journal of Translational Medicine*. 2020 April 22; 18(1):179. doi: 10.1186/s12967-020-02344-6. [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7174922/pdf/12967\\_2020\\_Article\\_2344.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7174922/pdf/12967_2020_Article_2344.pdf)
- <sup>491</sup> Eckerle LD, Becker MM, Halpin RA, Li K, Venter E, Lu X, Scherbakova S, Graham RL, Baric RS, Stockwell TB, Spiro DJ, Denison MR. *Infidelity of SARS-CoV Nsp14-exonuclease mutant virus replication is revealed by complete genome sequencing*. *PLoS Pathog*. 2010 May 6; 6(5):e1000896. doi: 10.1371/journal.ppat.1000896. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2865531/pdf/ppat.1000896.pdf>
- <sup>492</sup> Ogando NS, Zevenhoven-Dobbe JC, van der Meer Y, Bredenbeek PJ, Posthuma CC, Snijder EJ. *The Enzymatic Activity of nsp14 Exoribonuclease Is Critical for Replication of MERS-CoV and SARS-CoV-2*. *Journal of Virology*. 2020 November 9; 94(23):e01246-20. doi: 10.1128/JVI.01246-20. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7654266/pdf/JVI.01246-20.pdf>
- <sup>493</sup> Shannon A, Le NT, Selisko B, Eydoux C, Alvarez K, Guillemot JC, Decroly E, Peersen O, Ferron F, Canard B. *Remdesivir and SARS-CoV-2: Structural requirements at both nsp12 RdRp and nsp14 Exonuclease active-sites*. *Antiviral Res*. 2020 June; 178:104793. doi: 10.1016/j.antiviral.2020.104793. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7151495/pdf/main.pdf>
- <sup>494</sup> Ferron F, Subissi L, Silveira De Moraes AT, Le NTT, Sevajol M, Gluais L, Decroly E, Vonnrhein C, Bricogne G, Canard B, Imbert I. *Structural and molecular basis of mismatch correction and ribavirin excision from coronavirus RNA*. *Proc Natl Acad Sci U S A*. 2018 January 9; 115(2):E162-E171. doi: 10.1073/pnas.1718806115. Epub 2017 December 26. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5777078/pdf/pnas.201718806.pdf>

- 
- <sup>495</sup> Koyama T, Platt D, Parida L. *Variant analysis of SARS-CoV-2 genomes*. Bulletin of the World Health Organization (WHO). 2020 July 1; 98(7):495-504. doi: 10.2471/BLT.20.253591. Epub 2020 June 2. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7375210/pdf/BLT.20.253591.pdf>
- <sup>496</sup> Abou-Hamdan M, Hamze K, Abdel Sater A, Akl H, El-Zein N, Dandache I, Abdel-Sater F. *Variant analysis of the first Lebanese SARS-CoV-2 isolates*. Genomics. 2020 October 20:S0888-7543(20)31980-7. doi: 10.1016/j.ygeno.2020.10.021. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7572353/pdf/main.pdf>
- <sup>497</sup> Smith EC, Case JB, Blanc H, Isakov O, Shomron N, Vignuzzi M, Denison MR. *Mutations in coronavirus nonstructural protein 10 decrease virus replication fidelity*. Journal of Virology. 2015 June; 89(12):6418-26. doi: 10.1128/JVI.00110-15. Epub 2015 April 8. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4474304/pdf/zjv6418.pdf>
- <sup>498</sup> Baddock HT, Brolih S, Yosaatmadja Y, Ratnaweera M, Bielinski M, Swift LP, Cruz-Migoni A, Morris GM, Schofield CJ, Gileadi O, McHugh PJ. *Characterisation of the SARS-CoV-2 ExoN (nsp14ExoN-nsp10) complex: implications for its role in viral genome stability and inhibitor identification*. bioRxiv; 2020 August 13. DOI: 10.1101/2020.08.13.248211. <https://www.biorxiv.org/content/10.1101/2020.08.13.248211v1.full.pdf>
- <sup>499</sup> Kannan SR, Spratt AN, Quinn TP, Heng X, Lorson CL, Sönnnerborg A, Byrareddy SN, Singh K. *Infectivity of SARS-CoV-2: there Is Something More than D614G?* Journal Neuroimmune Pharmacology 2020 December; 15(4):574-577. doi: 10.1007/s11481-020-09954-3. Epub 2020 September 15. [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7490321/pdf/11481\\_2020\\_Article\\_9954.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7490321/pdf/11481_2020_Article_9954.pdf)
- <sup>500</sup> Groves DC, Rowland-Jones SL, Angyal A. *The D614G mutations in the SARS-CoV-2 spike protein: Implications for viral infectivity, disease severity and vaccine design*. Biochem Biophys Res Commun. 2020 November 5: S0006-291X(20)32038-6. doi: 10.1016/j.bbrc.2020.10.109. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7643658/pdf/main.pdf>
- <sup>501</sup> Andrew Rambaut, Nick Loman, Oliver Pybus, Wendy Barclay, Jeff Barrett, Alesandro Carabelli, Tom Connor, Tom Peacock, David L Robertson, Erik Volz, on behalf of COVID-19 Genomics Consortium UK (CoG-UK). *Preliminary genomic characterisation of an emergent SARS-CoV-2 lineage in the UK defined by a novel set of spike mutations*. <https://www.cogconsortium.uk/> <https://virological.org/t/preliminary-genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-the-uk-defined-by-a-novel-set-of-spike-mutations/563>
- <sup>502</sup> Bahrami A, Ferns GA. *Genetic and pathogenic characterization of SARS-CoV-2: a review*. Future Virology. 2020 Aug; 10.2217/fvl-2020-0129. doi: 10.2217/fvl-2020-0129. Epub 2020 August 26. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7451412/pdf/fvl-2020-0129.pdf>
- <sup>503</sup> Boni MF, Lemey P, Jiang X, Lam TT, Perry BW, Castoe TA, Rambaut A, Robertson DL. *Evolutionary origins of the SARS-CoV-2 sarbecovirus lineage responsible for the COVID-19 pandemic*. Nature Microbiology. 2020 Nov; 5(11):1408-1417. doi: 10.1038/s41564-020-0771-4. Epub 2020 July 28. <https://www.nature.com/articles/s41564-020-0771-4.pdf>

- <sup>504</sup> Gould, Stephen Jay, and Niles Eldredge. "Punctuated Equilibria: The Tempo and Mode of Evolution Reconsidered." *Paleobiology* 3, no. 2 (1977): 115-51. <http://www.jstor.org/stable/2400177>  
<http://www.somosbacteriasyvirus.com/phyletic.pdf>
- <sup>505</sup> Kosuge M, Furusawa-Nishii E, Ito K, Saito Y, Ogasawara K. *Point mutation bias in SARS-CoV-2 variants results in increased ability to stimulate inflammatory responses*. *Nature Science Reports* 2020 October 20; 10(1):17766. doi: 10.1038/s41598-020-74843-x.  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7575582/pdf/41598\\_2020\\_Article\\_74843.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7575582/pdf/41598_2020_Article_74843.pdf)
- <sup>506</sup> Folegatti PM, Ewer KJ, Aley PK, Angus B, Becker S, Belij-Rammerstorfer S, Bellamy D, Bibi S, Bittaye M, Clutterbuck EA, Dold C, Faust SN, Finn A, Flaxman AL, Hallis B, Heath P, Jenkin D, Lazarus R, Makinson R, Minassian AM, Pollock KM, Ramasamy M, Robinson H, Snape M, Tarrant R, Voysey M, Green C, Douglas AD, Hill AVS, Lambe T, Gilbert SC, Pollard AJ; Oxford COVID Vaccine Trial Group. *Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, single-blind, randomised controlled trial*. *Lancet*. 2020 August 15; 396(10249):467-478. doi: 10.1016/S0140-6736(20)31604-4. Epub 2020 July 20. Erratum in: *Lancet*. 2020 August 15; 396(10249):466. Erratum in: *Lancet*. 2020 December 12; 396(10266):1884.  
<https://www.thelancet.com/action/showPdf?pii=S0140-6736%2820%2931604-4>
- <sup>507</sup> CDC. Storage and Handling Summary.  
<https://www.cdc.gov/vaccines/covid-19/info-by-product/moderna/downloads/storage-summary.pdf>
- <sup>508</sup> <https://somalogic.com/>
- <sup>509</sup> Tilley, Aaron. "Cisco Systems Pulls Back From Smart City Push." *Wall Street Journal*, 28 Dec 2020.  
<https://www.wsj.com/articles/cisco-turns-off-lights-on-smart-city-push-11609178895>
- <sup>510</sup> "The Story of mRNA: From a loose idea to a tool that may help curb CoVID." *STAT*, 10 Nov 2020.  
<https://www.statnews.com/2020/11/10/the-story-of-mrna-how-a-once-dismissed-idea-became-a-leading-technology-in-the-covid-vaccine-race/>
- <sup>511</sup> "Explained: Why RNA Vaccines for Covid-19 Raced to the Front of the Pack." *MIT News*, 2020. *Massachusetts Institute of Technology* <https://news.mit.edu/2020/rna-vaccines-explained-covid-19-1211>
- <sup>512</sup> Langer, Robert and Folkman, Judah. Polymers for the sustained release of proteins and other macromolecules. *Nature* 263, 797–800 (1976). <https://doi.org/10.1038/263797a0>
- <sup>513</sup> Biondi E, Benner SA. *Artificially Expanded Genetic Information Systems for New Aptamer Technologies*. *Biomedicines*. 2018 May 9; 6(2):53. doi: 10.3390/biomedicines6020053.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6027400/pdf/biomedicines-06-00053.pdf>
- <sup>514</sup> Zhang L, Yang Z, Sefah K, Bradley KM, Hoshika S, Kim MJ, Kim HJ, Zhu G, Jiménez E, Cansiz S, Teng IT, Champanhac C, McLendon C, Liu C, Zhang W, Gerloff DL, Huang Z, Tan W, Benner SA. *Evolution of functional six-nucleotide DNA*. *J Am Chem Soc*. 2015 June 3; 137(21):6734-6737. doi: 10.1021/jacs.5b02251. Epub 2015 May 20.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4500535/pdf/nihms700342.pdf>
- <sup>515</sup> Mina, Michael J., and Kristian G. Andersen. "COVID-19 Testing: One Size Does Not Fit All." *Science*, vol. 371, no. 6525, January 2021, pp. 126–127. doi:10.1126/science.abe9187.

- <sup>516</sup> Kahn R, Kennedy-Shaffer L, Grad YH, Robins JM, Lipsitch M. *Potential Biases Arising from Epidemic Dynamics in Observational Seroprotection Studies*. American Journal of Epidemiology. 2020 September 1 doi: 10.1093/aje/kwaa188. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7499481/pdf/kwaa188.pdf>
- <sup>517</sup> Koczera P, Martin L, Marx G, Schuerholz T. *The Ribonuclease A Superfamily in Humans: Canonical RNases as the Buttress of Innate Immunity*. Int Journal of Molecular Sciences 2016 August 5; 17(8):1278. doi: 10.3390/ijms17081278  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5000675/pdf/ijms-17-01278.pdf>
- <sup>518</sup> Altman S. *Ribonuclease P*. Philos Trans R Soc Lond B Biol Sci. 2011 October 27; 366(1580):2936-41. doi: 10.1098/rstb.2011.0142  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3158923/pdf/rstb20110142.pdf>
- <sup>519</sup> Gitlin L, Karelsky S, Andino R. *Short interfering RNA confers intracellular antiviral immunity in human cells*. Nature. 2002 July 25; 418(6896):430-4. doi: 10.1038/nature00873. Epub 2002 June 26.  
<https://www.nature.com/articles/nature00873>
- <sup>520</sup> Manley JL, Fire A, Cano A, Sharp PA, Gefter ML. *DNA-dependent transcription of adenovirus genes in a soluble whole-cell extract*. Proc Natl Acad Sci. 1980 July; 77(7):3855-9. doi: 10.1073/pnas.77.7.3855  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC349725/pdf/pnas00494-0146.pdf>
- <sup>521</sup> Mandel, P. and Metais, P. *Les acides nucléiques du plasma sanguin chez l'homme* [Nuclear Acids In Human Blood Plasma]. C R Seances Soc Biol Fil. 1948 February; 142(3-4):241-243.  
<https://pubmed.ncbi.nlm.nih.gov/18875018/>
- <sup>522</sup> Agrawal S, Mayrand SH, Zamecnik PC, Pederson T. *Site-specific excision from RNA by RNase H and mixed-phosphate-backbone oligodeoxynucleotides*. Proc Natl Acad Sci. 1990 February; 87(4):1401-1405. doi: 10.1073/pnas.87.4.1401.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC53483/pdf/pnas01029-0151.pdf>
- <sup>523</sup> Anderson BR, Muramatsu H, Jha BK, Silverman RH, Weissman D, Karikó K. *Nucleoside modifications in RNA limit activation of 2'-5'-oligoadenylate synthetase and increase resistance to cleavage by RNase L*. Nucleic Acids Research. 2011 November; 39(21):9329-38. doi: 10.1093/nar/gkr586.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3241635/pdf/gkr586.pdf>
- <sup>524</sup> Lee JF, Stovall GM, Ellington AD. *Aptamer therapeutics advance*. Curr Opin Chem Biol. 2006 June; 10(3):282-9. doi: 10.1016/j.cbpa.2006.03.015. Epub 2006 April 18.
- <sup>525</sup> Keefe AD, Pai S, Ellington A. *Aptamers as therapeutics*. Nat Rev Drug Discov. 2010 July; 9(7):537-550. doi: 10.1038/nrd3141. Erratum in: Nat Rev Drug Discov. 2010 August; 9(8):660.  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7097324/pdf/41573\\_2010\\_Article\\_BFnrd3141.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7097324/pdf/41573_2010_Article_BFnrd3141.pdf)
- <sup>526</sup> Murakami E, Niu C, Bao H, Micolochick Steuer HM, Whitaker T, Nachman T, Sofia MA, Wang P, Otto MJ, Furman PA. *The mechanism of action of beta-D-2'-deoxy-2'-fluoro-2'-C-methylcytidine involves a second metabolic pathway leading to beta-D-2'-deoxy-2'-fluoro-2'-C-methyluridine 5'-triphosphate, a potent inhibitor of the hepatitis C virus RNA-dependent RNA polymerase*. Antimicrobial Agents and Chemotherapy. 2008 February; 52(2):458-464. doi: 10.1128/AAC.01184-07. Epub 2007 November 12.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2224766/pdf/1184-07.pdf>

<sup>527</sup> Karuna R, Yokokawa F, Wang K, Zhang J, Xu H, Wang G, Ding M, Chan WL, Abdul Ghafar N, Leonardi A, Seh CC, Seah PG, Liu W, Srinivasa RPS, Lim SP, Lakshminarayana SB, Growcott E, Babu S, Fenaux M, Zhong W, Gu F, Shi PY, Blasco F, Chen YL. *A Cyclic Phosphoramidate Prodrug of 2'-Deoxy-2'-Fluoro-2'-C-Methylguanosine for the Treatment of Dengue Virus Infection*. *Antimicrobial Agents and Chemotherapy*. 2020 November 17; 64(12):e00654-20. doi: 10.1128/AAC.00654-20.

<https://www.biorxiv.org/content/10.1101/2020.04.13.040287v1.full.pdf>

<https://aac.asm.org/content/aac/64/12/e00654-20.full-text.pdf>

<sup>528</sup> Liu J, Guo S, Cinier M, Shlyakhtenko LS, Shu Y, Chen C, Shen G, Guo P. *Fabrication of stable and RNase-resistant RNA nanoparticles active in gearing the nanomotors for viral DNA packaging*. *ACS Nano*. 2011 January 25; 5(1):237-246. doi: 10.1021/nn1024658. Epub 2010 December 14.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3026857/pdf/nihms258686.pdf>

<sup>529</sup> Joshi A, Rienks M, Theofilatos K, Mayr M. *Systems biology in cardiovascular disease: a multiomics approach*. *Nature Reviews Cardiology*. 2020 December 18. doi: 10.1038/s41569-020-00477-1.

<sup>530</sup> Kan CW, Tobos CI, Rissin DM, Wiener AD, Meyer RE, Svancara DM, Comperchio A, Warwick C, Millington R, Collier N, Duffy DC. *Digital enzyme-linked immunosorbent assays with sub-attomolar detection limits based on low numbers of capture beads combined with high efficiency bead analysis*. *Lab Chip*. 2020 June 21; 20(12):2122-2135. doi: 10.1039/d0lc00267d. Epub 2020 May 11.

<sup>531</sup> Shi J, Wen Z, Zhong G, Yang H, Wang C, Huang B, Liu R, He X, Shuai L, Sun Z, Zhao Y, Liu P, Liang L, Cui P, Wang J, Zhang X, Guan Y, Tan W, Wu G, Chen H, Bu Z. *Susceptibility of ferrets, cats, dogs, and other domesticated animals to SARS-coronavirus 2*. *Science*. 2020 May 29; 368(6494):1016-1020. doi: 10.1126/science.abb7015. Epub 2020 April 8.

[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7164390/pdf/368\\_1016.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7164390/pdf/368_1016.pdf)

<sup>532</sup> Datta, Shoumen Palit Austin, Tausifa Jan Saleem, Molood Barati, María Victoria López López, Marie-Laure Furgala, Diana C. Vanegas, Gérald Santucci, Pramod P. Khargonekar and Eric S. McLamore (2020) *Data, Analytics and Interoperability between Systems (IoT) is Incongruous with the Economics of Technology: Evolution of Porous Pareto Partition (P3)*. Chapter 2 in “*Big Data Analytics for Internet of Things*” 1st ed. Editors Tausifa Jan Saleem and Mohammad Ahsan Chishti © 2021 John Wiley & Sons, Inc. Draft copy of “P3” available from the MIT Library <https://dspace.mit.edu/handle/1721.1/123984>

<sup>533</sup> Michael Minna. <https://biology.mit.edu/undergraduate/current-students/subject-offerings/covid-19-sars-cov-2-and-the-pandemic/>

<sup>534</sup> Larremore, Daniel B., et al. *Test Sensitivity Is Secondary to Frequency and Turnaround Time for COVID-19 Surveillance*. preprint, *Infectious Diseases (except HIV/AIDS)*, 25 June 2020.

doi:10.1101/2020.06.22.20136309. [www.medrxiv.org/content/10.1101/2020.06.22.20136309v2.full.pdf](http://www.medrxiv.org/content/10.1101/2020.06.22.20136309v2.full.pdf)

<sup>535</sup> Larremore, Daniel B., et al. *Estimating SARS-CoV-2 Seroprevalence and Epidemiological Parameters with Uncertainty from Serological Surveys*. preprint, *Infectious Diseases (except HIV/AIDS)*, 20 April 2020. <https://www.medrxiv.org/content/10.1101/2020.04.15.20067066v2.full.pdf>



- <sup>536</sup> Lopez L, Nguyen T, Weber G, Kleimola K, Bereda M, Liu Y, Accorsi EK, Skates SJ, Santa Maria JP, Smith KR, Kalinich M. *Seroprevalence of anti-SARS-CoV-2 IgG Antibodies in the Staff of a Public School System in the Midwestern United States*. medRxiv [Preprint]. 2020 October 27:2020.10.23.20218651. doi: 10.1101/2020.10.23.20218651  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7605578/pdf/nihpp-2020.10.23.20218651.pdf>
- <sup>537</sup> Dialogues of Alfred North Whitehead as recorded by Lucien Price. Little, Brown & Co. Jan 1, 1954
- <sup>538</sup> Timeline of the MIT-Harvard Rivalry. <http://mta.scripts.mit.edu/CES/mit-harvard-rivalry-timeline/>
- <sup>539</sup> Majumder S, Deen MJ. *Smartphone Sensors for Health Monitoring and Diagnosis*. Sensors (Basel). 2019 May 9; 19(9):2164. doi: 10.3390/s19092164.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6539461/pdf/sensors-19-02164.pdf>
- <sup>540</sup> Sensors and Cellphones. <https://web.stanford.edu/class/cs75n/Sensors.pdf>
- <sup>541</sup> Byoung-Yong Chang and Su-Moon Park (2010) Electrochemical Impedance Spectroscopy. Annual Review of Analytical Chemistry, Vol. 3:207-229 <https://doi.org/10.1146/annurev.anchem.012809.102211>  
<https://www.annualreviews.org/doi/pdf/10.1146/annurev.anchem.012809.102211>
- <sup>542</sup> Schasfoort, Richard B. M., and Anna J. Tudos, editors. *Handbook of Surface Plasmon Resonance*: Royal Society of Chemistry, 2008. doi:10.1039/9781847558220.  
[www.biodyn.ro/course/literatura/Handbook%20of%20Surface%20Plasmon%20Resonance\\_2017.pdf](http://www.biodyn.ro/course/literatura/Handbook%20of%20Surface%20Plasmon%20Resonance_2017.pdf)
- <sup>543</sup> Steve Bradt “Biden Taps Eric Lander and Maria Zuber for Senior Science Posts.” *MIT News*  
<https://news.mit.edu/2021/biden-taps-lander-zuber-science-posts-0115>
- <sup>544</sup> Eric Lander. <https://biology.mit.edu/undergraduate/current-students/subject-offerings/covid-19-sars-cov-2-and-the-pandemic/> and <https://www.nature.com/articles/d41586-021-00118-8>
- <sup>545</sup> Martin, Emily T., et al. “Multiple versus Single Virus Respiratory Infections: Viral Load and Clinical Disease Severity in Hospitalized Children: Viral Coinfection in Children.” *Influenza and Other Respiratory Viruses*, vol. 6, no. 1, January 2012, pp. 71–77. doi:10.1111/j.1750-2659.2011.00265.x.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3175338/pdf/IRV-6-71.pdf>
- <sup>546</sup> Boldogh I, Albrecht T, Porter DD. *Persistent Viral Infections*. In: Baron S, editor. Medical Microbiology. 4th edition. Galveston (TX): University of Texas Medical Branch at Galveston; 1996. Chapter 46. <https://www.ncbi.nlm.nih.gov/books/NBK8538/>
- <sup>547</sup> Forsberg, J. (2020) Origins of Six Sigma [www.ss-mi.com/post/2020/06/17/the-origins-of-six-sigma](http://www.ss-mi.com/post/2020/06/17/the-origins-of-six-sigma)
- <sup>548</sup> John F. Kennedy, Rice University Stadium (Sept 12, 1962) <https://er.jsc.nasa.gov/seh/ricetalk.htm>
- <sup>549</sup> Koshland D.E. (1958) *Application of a Theory of Enzyme Specificity to Protein Synthesis*. Proc Natl Acad Sci 1958 Feb; 44(2):98-104. doi: 10.1073/pnas.44.2.98.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC335371/pdf/pnas00681-0050.pdf>  
<http://www.tulane.edu/~biochem/lecture/gbch701/koshland2.pdf>
- <sup>550</sup> Dereka, Bogdan *et al* (2021) Crossover from hydrogen to chemical bonding. *Science* 08 Jan 2021: Vol. 371, Issue 6525, pp. 160-164 DOI: 10.1126/science.abe1951  
<https://science.sciencemag.org/content/371/6525/160/tab-pdf>
- <sup>551</sup> Green DF, Tidor B. *Evaluation of electrostatic interactions*. Current Protocols in Bioinformatics. 2003 August; Chapter 8:Unit 8.3. doi: 10.1002/0471250953.bi0803s02

- <sup>552</sup> Rumelhart, D., Hinton, G. & Williams, R. Learning representations by back-propagating errors. *Nature* 323, 533–536 (1986). <https://doi.org/10.1038/323533a0>
- <sup>553</sup> Motulsky, Harvey, and Arthur Christopoulos. *Fitting Models to Biological Data Using Linear and Nonlinear Regression: A Practical Guide to Curve Fitting*. Oxford Univ Press, 2004. ISBN 9780195171808 <http://www.mcb5068.wustl.edu/MCB/Lecturers/Baranski/Articles/RegressionBook.pdf>
- <sup>554</sup> Machine Learning for Pharmaceutical Discovery and Synthesis. MIT <https://mlpds.mit.edu/papers/>
- <sup>555</sup> Mo, Yiming, *et al.* “Evaluating and Clustering Retrosynthesis Pathways with Learned Strategy.” *Chem Science*, 2021. doi:10.1039/D0SC05078D. <https://pubs.rsc.org/en/content/articlepdf/2021/sc/d0sc05078d>
- <sup>556</sup> Schreck JS, Coley CW, Bishop KJM. Learning Retrosynthetic Planning through Simulated Experience. *ACS Cent Sci*. 2019 June 26; 5(6):970-981. doi: 10.1021/acscentsci.9b00055. Epub 2019 May 31. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6598174/pdf/oc9b00055.pdf>
- <sup>557</sup> Project Oxygen <http://oxygen.csail.mit.edu/videosketching.html> & <http://bit.ly/META-TOOL-SUITE>
- <sup>558</sup> Brooks BR, Brooks CL 3rd, Mackerell AD Jr, Nilsson L, Petrella RJ, Roux B, Won Y, Archontis G, Bartels C, Boresch S, Caflisch A, Caves L, Cui Q, Dinner AR, Feig M, Fischer S, Gao J, Hodoscek M, Im W, Kuczera K, Lazaridis T, Ma J, Ovchinnikov V, Paci E, Pastor RW, Post CB, Pu JZ, Schaefer M, Tidor B, Venable RM, Woodcock HL, Wu X, Yang W, York DM, Karplus M. *CHARMM: the biomolecular simulation program*. *J Computational Chemistry* 2009 July 30; 30(10):1545-614. doi: 10.1002/jcc.21287. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2810661/pdf/nihms101618.pdf>
- <sup>559</sup> Simulation Tools [www.ansys.com/products/3d-design](http://www.ansys.com/products/3d-design)
- <sup>560</sup> Huggins DJ, Sherman W, Tidor B. *Rational approaches to improving selectivity in drug design*. *J Med Chem*. 2012 February 23; 55(4):1424-44. doi: 10.1021/jm2010332. Epub 2012 January 12. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3285144/pdf/jm2010332.pdf>
- <sup>561</sup> Lillicrap, T., Cownden, D., Tweed, D. *et al.* Random synaptic feedback weights support error backpropagation for deep learning. *Nature Communications* 7, 13276 (2016). <https://doi.org/10.1038/ncomms13276> and <https://www.nature.com/articles/ncomms13276.pdf>
- <sup>562</sup> Jonathan M. Stokes, Kevin Yang, Kyle Swanson, Wengong Jin, Andres Cubillos-Ruiz, Nina M. Donghia, Craig R. MacNair, Shawn French, Lindsey A. Carfrae, Zohar Bloom-Ackerman, Victoria M. Tran, Anush Chiappino-Pepe, Ahmed H. Badran, Ian W. Andrews, Emma J. Chory, George M. Church, Eric D. Brown, Tommi S. Jaakkola, Regina Barzilay and James J. Collins (2020) “A Deep Learning Approach to Antibiotic Discovery.” *Cell*, vol. 180, no. 4, February 2020, pp. 688-702.e13 doi:10.1016/j.cell.2020.01.021 [https://www.cell.com/cell/pdf/S0092-8674\(20\)30102-1.pdf](https://www.cell.com/cell/pdf/S0092-8674(20)30102-1.pdf)
- <sup>563</sup> St John PC, Phillips C, Kemper TW, Wilson AN, Guan Y, Crowley MF, Nimlos MR, Larsen RE. *Message-passing neural networks for high-throughput polymer screening*. *J Chem Phys*. 2019 June 21; 150(23):234111. doi: 10.1063/1.5099132. <https://arxiv.org/pdf/1704.01212.pdf>
- <sup>564</sup> Zhang, Xiang, *et al.* (2019) “Deep Neural Network Hyperparameter Optimization with Orthogonal Array Tuning.” July 2019. <http://arxiv.org/abs/1907.13359> <https://arxiv.org/pdf/1907.13359.pdf>
- <sup>565</sup> “Artificial Intelligence Yields New Antibiotic.” *MIT News. Massachusetts Institute of Technology*. <http://news.mit.edu/2020/artificial-intelligence-identifies-new-antibiotic-0220>

- <sup>566</sup> Datta, Shoumen Palit Austin, Tausifa Jan Saleem, Molood Barati, María Victoria López López, Marie-Laure Furgala, Diana C. Vanegas, Gérald Santucci, Pramod P. Khargonekar and Eric S. McLamore (2020) *Data, Analytics and Interoperability between Systems (IoT) is Incongruous with the Economics of Technology: Evolution of Porous Pareto Partition (P3)*. Chapter 2. Draft copy of “P3” from MIT Library <https://dspace.mit.edu/handle/1721.1/123984> in “*Big Data Analytics for Internet of Things*” 1st ed. Editors Tausifa Jan Saleem and Mohammad Ahsan Chishti, 2021. John Wiley & Sons, Inc.
- <sup>567</sup> Lars Kai Hansen and Peter Salamon (1990) *Neural Network Ensembles*. IEEE Transactions on Pattern Analysis and Machine Intelligence, Vol. 12, No. 10, October 1990. <https://pdfs.semanticscholar.org/257d/c8ae2a8353bb2e86c1b7186e7d989fb433d3.pdf>
- <sup>568</sup> Hannah K. Wayment-Steele, Wipapat Kladwang, Eterna Participants, Rhiju Das. *RNA secondary structure packages ranked and improved by high-throughput experiments*. bioRxiv 2020.05.29.124511; doi: <https://doi.org/10.1101/2020.05.29.124511> <https://www.biorxiv.org/content/10.1101/2020.05.29.124511v1.full.pdf>
- <sup>569</sup> Huang, Yuan, et al. “Structural and Functional Properties of SARS-CoV-2 Spike Protein: Potential Antivirus Drug Development for COVID-19.” *Acta Pharmacologica Sinica*, vol. 41, no. 9, Sep 2020, pp. 1141–1149. doi:10.1038/s41401-020-0485-4. [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7396720/pdf/41401\\_2020\\_Article\\_485.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7396720/pdf/41401_2020_Article_485.pdf)
- <sup>570</sup> Zeng W, Liu G, Ma H, Zhao D, Yang Y, Liu M, Mohammed A, Zhao C, Yang Y, Xie J, Ding C, Ma X, Weng J, Gao Y, He H, Jin T. *Biochemical characterization of SARS-CoV-2 nucleocapsid protein*. *Biochem Biophysics Research Communications*. 2020 June 30; 527(3):618-623. doi: 10.1016/j.bbrc.2020.04.136. Epub 2020 April 30. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7190499/pdf/main.pdf>
- <sup>571</sup> Thomas S. *The Structure of the Membrane Protein of SARS-CoV-2 Resembles the Sugar Transporter SemiSWEET*. *Pathogens and Immunity*. 2020 October 19; 5(1):342-363. doi: 10.20411/pai.v5i1.377 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7608487/pdf/pai-5-342.pdf>
- <sup>572</sup> Schoeman D, Fielding BC. *Coronavirus envelope protein: current knowledge*. *Virol J*. 2019 May 27; 16(1):69. doi: 10.1186/s12985-019-1182-0. [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6537279/pdf/12985\\_2019\\_Article\\_1182.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6537279/pdf/12985_2019_Article_1182.pdf)
- <sup>573</sup> Deeks, Jonathan J., and Angela E. Raffle. “Lateral Flow Tests Cannot Rule out SARS-CoV-2 Infection.” *British Medical Journal* <https://doi.org/10.1136/bmj.m4787> (Published 11 December 2020) <https://www.bmj.com/content/bmj/371/bmj.m4787.full.pdf>
- <sup>574</sup> Peeling RW, Wedderburn CJ, Garcia PJ, Boeras D, Fongwen N, Nkengasong J, Sall A, Tanuri A, Heymann DL. *Serology testing in the COVID-19 pandemic response*. *Lancet Infect Dis*. 2020 September; 20(9):e245-e249. doi: 10.1016/S1473-3099(20)30517-X. Epub 2020 July 17. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7367660/pdf/main.pdf>
- <sup>575</sup> Nishiga M, Wang DW, Han Y, Lewis DB, Wu JC. *COVID-19 and cardiovascular disease: from basic mechanisms to clinical perspectives*. *Nature Reviews in Cardiology*. 2020 September ; 17(9):543-558. doi: 10.1038/s41569-020-0413-9. Epub 2020 July 20. [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7370876/pdf/41569\\_2020\\_Article\\_413.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7370876/pdf/41569_2020_Article_413.pdf)

- <sup>576</sup> Taguchi M, Ptitsyn A, McLamore ES, Claussen JC. *Nanomaterial-mediated Biosensors for Monitoring Glucose*. J Diabetes Sci Technol. 2014 March; 8(2):403-411. doi: 10.1177/1932296814522799. Epub 2014 March 2 [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4455391/pdf/10.1177\\_1932296814522799.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4455391/pdf/10.1177_1932296814522799.pdf)
- <sup>577</sup> Deo PN, Deshmukh R. *Oral microbiome: Unveiling the fundamentals*. J Oral Maxillofac Pathol. 2019 January-April; 23(1):122-128. doi: 10.4103/jomfp.JOMFP\_304\_18.
- <sup>578</sup> Blume, C., Jackson, C.L., Spalluto, C.M. *et al.* A novel ACE2 isoform is expressed in human respiratory epithelia and is upregulated in response to interferons and RNA respiratory virus infection. *Nature Genetics* (2021). <https://doi.org/10.1038/s41588-020-00759-x>  
<https://www.nature.com/articles/s41588-020-00759-x.pdf>  
<https://www.biorxiv.org/content/10.1101/2020.07.31.230870v1.full.pdf> (July 31, 2020)
- <sup>579</sup> Watanabe Y, Berndsen ZT, Raghvani J, Seabright GE, Allen JD, Pybus OG, McLellan JS, Wilson IA, Bowden TA, Ward AB, Crispin M. *Vulnerabilities in coronavirus glycan shields despite extensive glycosylation*. Nat Commun. 2020 May 27; 11(1):2688. doi: 10.1038/s41467-020-16567-0  
<https://www.nature.com/articles/s41467-020-16567-0>  
<https://www.nature.com/articles/s41467-020-16567-0.pdf>  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7253482/pdf/41467\\_2020\\_Article\\_16567.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7253482/pdf/41467_2020_Article_16567.pdf)
- <sup>580</sup> Grant OC, Montgomery D, Ito K, Woods RJ. *Analysis of the SARS-CoV-2 spike protein glycan shield: implications for immune recognition*. bioRxiv [Preprint]. 2020 May 1: 2020.04.07.030445. doi: 10.1101/2020.04.07.030445. Update in: Nature Science Reports. 2020 September 14; 10(1):14991.  
<https://www.nature.com/articles/s41598-020-71748-7>  
<https://www.nature.com/articles/s41598-020-71748-7.pdf>  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7217288/pdf/nihpp-2020.04.07.030445.pdf>
- <sup>581</sup> Zhou Y, Lu K, Pfefferle S, Bertram S, Glowacka I, Drosten C, Pöhlmann S, Simmons G. *A single asparagine-linked glycosylation site of the severe acute respiratory syndrome coronavirus spike glycoprotein facilitates inhibition by mannose-binding lectin through multiple mechanisms*. J of Virology 2010 September; 84(17):8753-64. doi: 10.1128/JVI.00554-10. <https://jvi.asm.org/content/84/17/8753>  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2919028/pdf/0554-10.pdf>
- <sup>582</sup> Cummings RD, McEver RP. *C-type Lectins*. In: Varki A, Cummings RD, Esko JD, et al., editors. *Essentials of Glycobiology*. 2nd edition. Cold Spring Harbor Laboratory Press; 2009. Chapter 31.  
<https://www.ncbi.nlm.nih.gov/books/NBK1943/>
- <sup>583</sup> Chiodo, Fabrizio, et al. *Novel ACE2-Independent Carbohydrate-Binding of SARS-CoV-2 Spike Protein to Host Lectins and Lung Microbiota*. preprint, Immunology, 14 May 2020. doi:10.1101/2020.05.13.092478. bioRxiv; 2020. DOI: 10.1101/2020.05.13.092478.  
<https://www.biorxiv.org/content/10.1101/2020.05.13.092478v1.full.pdf>
- <sup>584</sup> Daniela A. Oliveira, Loreen R. Stromberg, Cicero C. Pola, Kshama Parate, Nicholas D. Cavallaro, Jonathan C. Claussen, Eric S. McLamore, and Carmen L. Gomes. "Biomimetic nanosensors for measuring pathogenic bacteria in complex food matrices". Proc. SPIE 11020, Smart Biomedical and Physiological Sensor Technology XVI, 110200J (16 August 2019); <https://doi.org/10.1117/12.2519523>

- <sup>585</sup> Gupta B, Sadaria D, Warriar VU, Kirtonia A, Kant R, Awasthi A, Baligar P, Pal JK, Yuba E, Sethi G, Garg M, Gupta RK. *Plant lectins and their usage in preparing targeted nanovaccines for cancer immunotherapy*. *Seminars in Cancer Biology*. 2020 February 14; S1044-579X(20)30038-9. doi: 10.1016/j.semcancer.2020.02.005
- <sup>586</sup> Keeffe, Jennifer Rae (2009) *Engineering Cyanovirin-N for Enhanced Viral Neutralization*. Dissertation (Ph.D.), California Institute of Technology. doi:10.7907/QSJH-5X56 <https://resolver.caltech.edu/CaltechETD:etd-12232008-110031>  
Chapter 4 Lectibody: *Design and characterization of a cyanovirin-N – Fc chimera*  
[https://thesis.library.caltech.edu/5131/5/04\\_Lectibody.pdf](https://thesis.library.caltech.edu/5131/5/04_Lectibody.pdf)
- <sup>587</sup> Hamorsky KT, Kouokam JC, Dent MW, Grooms TN, Husk AS, Hume SD, Rogers KA, Villinger F, Morris MK, Hanson CV, Matoba N. *Engineering of a Lectibody Targeting High-Mannose-Type Glycans of the HIV Envelope*. *Mol Ther*. 2019 November 6; 27(11):2038-2052. doi: 10.1016/j.ymthe.2019.07.021. [www.cell.com/molecular-therapy-family/molecular-therapy/pdfExtended/S1525-0016\(19\)30360-0](http://www.cell.com/molecular-therapy-family/molecular-therapy/pdfExtended/S1525-0016(19)30360-0)
- <sup>588</sup> Hong SA, Kwon J, Kim D, Yang S. *A rapid, sensitive and selective electrochemical biosensor with concanavalin A for the preemptive detection of norovirus*. *Biosens Bioelectron*. 2015 Feb 15; 64:338-44. doi: 10.1016/j.bios.2014.09.025
- <sup>589</sup> Simão EP, Silva DBS, Cordeiro MT, Gil LHV, Andrade CAS, Oliveira MDL. *Nanostructured impedimetric lectin-based biosensor for arboviruses detection*. *Talanta*. 2020 February 1; 208:120338. doi: 10.1016/j.talanta.2019.120338. Epub 2019 September 6.
- <sup>590</sup> Vanegas DC, Gomes CL, Cavallaro ND, Giraldo-Escobar D, McLamore ES. *Emerging Biorecognition and Transduction Schemes for Rapid Detection of Pathogenic Bacteria in Food*. *Compr Rev Food Sci Food Saf*. 2017 November; 16(6):1188-1205. doi: 10.1111/1541-4337.12294. Epub 2017 September 11. <https://onlinelibrary.wiley.com/doi/epdf/10.1111/1541-4337.12294>
- <sup>591</sup> Dornelles LN, Pereira-Ferrari L, Messias-Reason I. *Mannan-binding lectin plasma levels in leprosy: deficiency confers protection against the lepromatous but not the tuberculoid forms*. *Clin Exp Immunol*. 2006 September; 145(3):463-8. doi: 10.1111/j.1365-2249.2006.03161.x. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1809702/pdf/cei0145-0463.pdf>
- <sup>592</sup> Jin, SH., Ahn, K.J. & An, S. Importance of the immune response to *Mycobacterium leprae* in the skin. *Biomedical Dermatology* 2, 1 (2018). <https://doi.org/10.1186/s41702-017-0012-5>  
<https://biomeddermatol.biomedcentral.com/track/pdf/10.1186/s41702-017-0012-5.pdf>
- <sup>593</sup> Luís Cláudio Nascimento da Silva, Juliana Silva Pereira Mendonça, Wesley Felix de Oliveira, Karla Lílian Rodrigues Batista, Adrielle Zagnignan, Isabelle Freire Tabosa Viana, Maria Tereza dos Santos Correia. Exploring lectin–glycan interactions to combat COVID-19: Lessons acquired from other enveloped viruses. *Glycobiology*, 2020; cwaa099, <https://doi.org/10.1093/glycob/cwaa099>  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7665446/pdf/cwaa099.pdf>
- <sup>594</sup> Olenik, S., Lee, H.S. & Güder, F. The future of near-field communication-based wireless sensing. *Nat Rev Mater* (2021). 02 March 2021 <https://doi.org/10.1038/s41578-021-00299-8>  
<https://www.nature.com/articles/s41578-021-00299-8.pdf>



- <sup>595</sup> Coronavirus Tracker. *Financial Times*. February 16, 2021.  
<https://www.ft.com/content/a2901ce8-5eb7-4633-b89c-cbdf5b386938>
- <sup>596</sup> Hunt, Caroline (1996) *Child waste pickers in India: the occupation and its health risks*. *Environment and Urbanization*, Vol. 8, No. 2, October 1996, pages 111-118.  
[https://www.ucl.ac.uk/dpu-projects/drivers\\_urb\\_change/urb\\_environment/pdf\\_hazards\\_pollution/IIED\\_hunt\\_indiachildren.pdf](https://www.ucl.ac.uk/dpu-projects/drivers_urb_change/urb_environment/pdf_hazards_pollution/IIED_hunt_indiachildren.pdf)
- <sup>597</sup> Changeux JP, Amoura Z, Rey FA, Miyara M. *A nicotinic hypothesis for Covid-19 with preventive and therapeutic implications*. *C R Biol*. 2020 June 5; 343(1):33-39. doi: 10.5802/crbio.8.  
<https://www.qeios.com/read/FXGQSB/pdf>
- <sup>598</sup> Cheng MH, Porritt RA, Rivas MN, Krieger JM, Ozdemir AB, Garcia G, Arumugaswami V, Fries BC, Arditi M, Bahar I. *A monoclonal antibody against staphylococcal enterotoxin B superantigen inhibits SARS-CoV-2 entry in vitro*. *bioRxiv* [Preprint]. 2020 November 24. doi: 10.1101/2020.11.24.395079.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7709177/pdf/nihpp-2020.11.24.395079.pdf>
- <sup>599</sup> The Logical Indian. *35.5 Crore Women Don't Have Access To Toilet In India: Report*. 20 Nov. 2017.  
<https://thelogicalindian.com/story-feed/awareness/india-worst-in-the-world-for-the-highest-number-of-people-without-basic-sanitation/>
- <sup>600</sup> Caruso BA, Clasen TF, Hadley C, Yount KM, Haardörfer R, Rout M, Dasmohapatra M, Cooper HL. *Understanding and defining sanitation insecurity: women's gendered experiences of urination, defecation and menstruation in rural Odisha, India*. *BMJ Glob Health*. 2017 October 9; 2(4):e000414.  
doi: 10.1136/bmjgh-2017-000414.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5640070/pdf/bmjgh-2017-000414.pdf>
- <sup>601</sup> Global Atlas of Helminth Infections. [www.thiswormyworld.org/maps/open-defecation-in-india](http://www.thiswormyworld.org/maps/open-defecation-in-india)
- <sup>602</sup> *Shocking Picture of India's Tribal Kids Eating from Garbage - Planet Custodian*.  
<https://www.planetcustodian.com/shocking-picture-of-indias-tribal-kids-eating-from-garbage/8639/>
- <sup>603</sup> "Kolkata Dump That's Permanently on Fire: Most People Die by 50." *The Guardian*, 24 Oct. 2016.  
<http://www.theguardian.com/cities/2016/oct/24/difficult-breathe-inside-kolkata-india-rubbish-dump-permanently-fire>
- <sup>604</sup> <https://www.pinterest.com/pin/506866133033374604/>
- <sup>605</sup> "Interview: Photojournalist Captures the Heartbreaking Reality of Child Labor in Bangladesh." *My Modern Met*, 15 November 2018. <https://mymodernmet.com/gmb-akash-photojournalist/>.
- <sup>606</sup> Ainla A, Mousavi MPS, Tsaloglou MN, Redston J, Bell JG, Fernández-Abedul MT, Whitesides GM. *Open-Source Potentiostat for Wireless Electrochemical Detection with Smartphones*. *Anal Chem*. 2018 May 15; 90(10):6240-6246. doi: 10.1021/acs.analchem.8b00850. Epub 2018 April 24.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5997382/pdf/ac8b00850.pdf>
- <sup>607</sup> Cucinotta D, Vanelli M. *WHO Declares COVID-19 a Pandemic*. *Acta Biomed*. 2020 March 19; 91(1):157-160. doi: 10.23750/abm.v91i1.9397  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7569573/pdf/ACTA-91-157.pdf>
- <sup>608</sup> Coronavirus Pandemic (CoVID-19) <https://ourworldindata.org/coronavirus>

- <sup>609</sup> O'Donnell, Jeff Mason, Carl. "‘Things Are Tenuous,’ U.S. CDC Says as Downward Trend in COVID-19 Cases Stalls." *Reuters*, 26 February 2021. [www.reuters.com](http://www.reuters.com)  
<https://www.reuters.com/article/us-health-coronavirus-whitehouse-idUSKBN2AQ2GG>
- <sup>610</sup> Mustafa Kemal Sezgintürk (ed) *COMMERCIAL BIOSENSORS AND THEIR APPLICATIONS: Clinical, Food, And Beyond*. ELSEVIER, 2020. ISBN-13 978-0-12-818592-6  
<https://doi.org/10.1016/C2018-0-03960-2>
- <sup>611</sup> Yoo EH, Lee SY. Glucose biosensors: an overview of use in clinical practice. *Sensors (Basel)*. 2010; 10(5):4558-76. doi: 10.3390/s100504558. Epub 2010 May 4.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3292132/pdf/sensors-10-04558.pdf>
- <sup>612</sup> Datta, S. (2020) *Sensible Sensor Systems are Essential Tools for Humans, Animals, Plants and the Environment: Understanding the Context of What to Sense in the Climate of Infectious Diseases, SARS-CoV-2, CoVID-19 and How to Prepare to Predict Future Pandemics, Epidemics and Endemics by Implementing Connected Networks of CITCOM*. Unpublished manuscript "CITCOM" available from the MIT Library <https://dspace.mit.edu/handle/1721.1/128017>
- <sup>613</sup> Santos RAS, Sampaio WO, Alzamora AC, Motta-Santos D, Alenina N, Bader M, Campagnole-Santos MJ. (2018) *The ACE2/Angiotensin-(1-7)/MAS Axis of the Renin-Angiotensin System: Focus on Angiotensin-(1-7)*. *Physiol Rev*. 2018 Jan 1; 98(1): 505-553. doi: 10.1152/physrev.00023.2016  
<https://journals.physiology.org/doi/pdf/10.1152/physrev.00023.2016>  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7203574/?report=printable>
- <sup>614</sup> Liu, P., Wysocki, J., Serfozo, P. *et al*. A Fluorometric Method of Measuring Carboxypeptidase Activities for Angiotensin II and Apelin-13. *Nature Scientific Reports* 7, 45473 (2017).  
<https://doi.org/10.1038/srep45473> <https://www.nature.com/articles/srep45473.pdf>
- <sup>615</sup> Phenylalanine Ammonia Lyase (PAL) [https://www.genome.jp/dbget-bin/www\\_bget?ec:4.3.1.24](https://www.genome.jp/dbget-bin/www_bget?ec:4.3.1.24)
- <sup>616</sup> National Library of Medicine (US), National Center for Biotechnology Information; PubChem Compound Summary for CID 5957728, Cinnamate.  
<https://pubchem.ncbi.nlm.nih.gov/compound/Cinnamate>
- <sup>617</sup> Smith HQ, Li C, Stanley CA, Smith TJ. *Glutamate Dehydrogenase, a Complex Enzyme at a Crucial Metabolic Branch Point*. *Neurochem Res*. 2019 January; 44(1):117-132. doi: 10.1007/s11064-017-2428-0. Epub 2017 October 27. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5924581/pdf/nihms943481.pdf>
- <sup>618</sup> Muus, Jytte. "The Amino Acid Composition of Human Salivary Amylase <sup>1</sup>." *Journal of the American Chemical Society*, vol. 76, no. 20, October 1954, pp. 5163-5165. doi:10.1021/ja01649a059.
- <sup>619</sup> Human Alpha-Amylase 1B. <https://www.uniprot.org/uniprot/P0DTE7>
- <sup>620</sup> Brain Natriuretic Peptide (BNP) <https://www.uniprot.org/uniprot/P16860>
- <sup>621</sup> Trevor, Anthony J., et al. *Katzung & Trevor's Pharmacology: Examination & Board Review*. 11th edition, McGraw-Hill Medical, Lange, 2021. ISBN-13 978-0071826358. In Ch. 17: Vasoactive Peptides.  
<https://doctorlib.info/pharmacology/pharmacology-examination-board-review/17.html>

- <sup>622</sup> Yang P, Kuc RE, Brame AL, Dyson A, Singer M, Glen RC, Cheriyan J, Wilkinson IB, Davenport AP, Maguire JJ. [Pyr<sup>1</sup>]Apelin-13<sub>(1-12)</sub> Is a Biologically Active ACE2 Metabolite of the Endogenous Cardiovascular Peptide [Pyr<sup>1</sup>]Apelin-13. *Frontiers in Neuroscience*. 2017 February 28; 11:92. doi: 10.3389/fnins.2017.00092 [www.ncbi.nlm.nih.gov/pmc/articles/PMC5329011/pdf/fnins-11-00092.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5329011/pdf/fnins-11-00092.pdf)
- <sup>623</sup> Karikó K, Buckstein M, Ni H, Weissman D. (2005) *Suppression of RNA recognition by Toll-like receptors: the impact of nucleoside modification and the evolutionary origin of RNA*. *Immunity*. 2005 August; 23(2):165-175. DOI: 10.1016/j.immuni.2005.06.008 <https://www.cell.com/action/showPdf?pii=S1074-7613%2805%2900211-6>
- <sup>624</sup> Graham, Barney S. and Sullivan, Nancy J. (2017) Emerging viral diseases from a vaccinology perspective: preparing for the next pandemic. *Nature Immunology* 19, 20–28 (2018) <https://doi.org/10.1038/s41590-017-0007-9> & <https://www.nature.com/articles/s41590-017-0007-9.pdf>
- <sup>625</sup> US CDC. “Polio Elimination in the United States.” *US Centers for Disease Control and Prevention*, 23 July 2021. <https://www.cdc.gov/polio/what-is-polio/polio-us.html>
- <sup>626</sup> Alleman MM, Jorba J, Greene SA, et al. Update on Vaccine-Derived Poliovirus Outbreaks: Worldwide July 2019–February 2020. *MMWR Morbidity and Mortality Weekly Report* 2020; 69: 489–495. DOI: <http://dx.doi.org/10.15585/mmwr.mm6916a1> <https://www.cdc.gov/mmwr/volumes/69/wr/pdfs/mm6916a1-H.pdf>
- <sup>627</sup> Willrich, Michael. *Pox: An American History*. Penguin Press, 2011. ISBN-13 978-0143120780
- <sup>628</sup> Babkin IV, Babkina IN. *The origin of the variola virus*. *Viruses*. 2015 March 10; 7(3):1100-1112. DOI: 10.3390/v7031100. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4379562/pdf/viruses-07-01100.pdf>
- <sup>629</sup> Pergam SA, Limaye AP; AST Infectious Diseases Community of Practice. Varicella zoster virus (VZV) in solid organ transplant recipients. *Am J Transplant*. 2009 December 9 (Suppl 4): S108-115. DOI: 10.1111/j.1600-6143.2009.02901.x <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2919834/pdf/nihms-208518.pdf>
- <sup>630</sup> Mandavilli, Apoorva. “A ‘Historic Event’: First Malaria Vaccine Approved by W.H.O.” *The New York Times*, 6 October 2021. <https://www.nytimes.com/2021/10/06/health/malaria-vaccine-who.html>
- <sup>631</sup> Centers for Disease Control and Prevention. *CDC - Malaria - About Malaria - History - Ross and the Discovery That Mosquitoes Transmit Malaria Parasites*. [www.cdc.gov/malaria/about/history/ross.html](http://www.cdc.gov/malaria/about/history/ross.html)
- <sup>632</sup> Nobel Prize in Physiology or Medicine. 1902. [www.nobelprize.org/prizes/medicine/1902/summary/](http://www.nobelprize.org/prizes/medicine/1902/summary/)
- <sup>633</sup> UN Vote on Waiving Patent Law for Life-Saving CoVID-19 Vaccines [www.reddit.com/r/MapPorn/comments/m50ae8/un\\_vote\\_on\\_waiving\\_patent\\_law\\_for\\_lifesaving/](http://www.reddit.com/r/MapPorn/comments/m50ae8/un_vote_on_waiving_patent_law_for_lifesaving/)
- <sup>634</sup> “Oxfam Response to WTO TRIPS Waiver for COVID-19 Vaccines Being Blocked Again by Rich Countries - a Massive Missed Opportunity to Speed up and Scale up the Production of Lifesaving Vaccines Worldwide - World.” *ReliefWeb*. <https://reliefweb.int/report/world/oxfam-response-wto-trips-waiver-covid-19-vaccines-being-blocked-again-rich-countries>

- <sup>635</sup> Johns Hopkins University. Corona Virus Resource Center. <https://coronavirus.jhu.edu/map.html>
- <sup>636</sup> “The Pandemic’s True Death Toll.” *The Economist*, October 2021.  
[www.economist.com/graphic-detail/coronavirus-excess-deaths-tracker](http://www.economist.com/graphic-detail/coronavirus-excess-deaths-tracker)
- <sup>637</sup> Gilman MSA, Furmanova-Hollenstein P, Pascual G, B van 't Wout A, Langedijk JPM, McLellan JS. *Transient opening of trimeric prefusion RSV F proteins*. Nat Communications. 2019 May 8; 10(1):2105. DOI: 10.1038/s41467-019-09807-5  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6506550/pdf/41467\\_2019\\_Article\\_9807.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6506550/pdf/41467_2019_Article_9807.pdf)
- <sup>638</sup> Zhang J, Cai Y, Xiao T, Lu J, Peng H, Sterling SM, Walsh RM Jr, Rits-Volloch S, Zhu H, Woosley AN, Yang W, Sliz P, Chen B. *Structural impact on SARS-CoV-2 spike protein by D614G substitution*. Science. 2021 April 30; 372(6541):525-530. doi: 10.1126/science.abf2303. Epub 2021 March 16.  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8139424/pdf/372\\_525.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8139424/pdf/372_525.pdf)
- <sup>639</sup> Gobeil SM, Janowska K, McDowell S, Mansouri K, Parks R, Manne K, Stalls V, Kopp MF, Henderson R, Edwards RJ, Haynes BF, Acharya P. *D614G Mutation Alters SARS-CoV-2 Spike Conformation and Enhances Protease Cleavage at the S1/S2 Junction*. Cell Reports 2021 January 12; 34(2):108630. DOI: 10.1016/j.celrep.2020.108630. Epub 2020 December 26  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7762703/pdf/main.pdf>
- <sup>640</sup> McLellan JS, Chen M, Leung S, Graepel KW, Du X, Yang Y, Zhou T, Baxa U, Yasuda E, Beaumont T, Kumar A, Modjarrad K, Zheng Z, Zhao M, Xia N, Kwong PD, Graham BS. *Structure of RSV fusion glycoprotein trimer bound to a prefusion-specific neutralizing antibody*. Science. 2013 May 31; 340(6136):1113-1117. DOI: 10.1126/science.1234914. Epub 2013 April 25.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4459498/pdf/nihms695082.pdf>
- <sup>641</sup> Swanson KA, Settembre EC, Shaw CA, Dey AK, Rappuoli R, Mandl CW, Dormitzer PR, Carfi A. *Structural basis for immunization with postfusion respiratory syncytial virus fusion F glycoprotein (RSV F) to elicit high neutralizing antibody titers*. Proceedings of the National Academy of Sciences (USA). 2011 June 7; 108(23):9619-24. DOI: 10.1073/pnas.1106536108. Epub 2011 May 17  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3111287/pdf/pnas.201106536.pdf>
- <sup>642</sup> McLellan JS, Yang Y, Graham BS, Kwong PD. *Structure of respiratory syncytial virus fusion glycoprotein in the postfusion conformation reveals preservation of neutralizing epitopes*. J of Virology 2011 August; 85(15):7788-96. DOI: 10.1128/JVI.00555-11. Epub 2011 May 25.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3147929/pdf/zjv7788.pdf>
- <sup>643</sup> Judith Falloon, Jing Yu, Mark T Esser, Tonya Villafana, Li Yu, Filip Dubovsky, Therese Takas, Myron J Levin, Ann R Falsey. An Adjuvanted, Postfusion F Protein–Based Vaccine Did Not Prevent Respiratory Syncytial Virus Illness in Older Adults, *The Journal of Infectious Diseases*, 216, Issue 11, 1 Dece 2017, pp. 1362–1370. <https://doi.org/10.1093/infdis/jix503>
- <sup>644</sup> Sanders RW, Moore JP. *Virus vaccines: proteins prefer prolines*. Cell Host Microbe. 2021 March 10; 29(3):327-333. DOI: 10.1016/j.chom.2021.02.002  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7945883/pdf/main.pdf>
- <sup>645</sup> Cross, Ryan (2020) *The tiny tweak behind COVID-19 vaccines*. 2020 September 29. C&EN 98 issue 38  
<https://cen.acs.org/pharmaceuticals/vaccines/tiny-tweak-behind-COVID-19/98/i38>

- <sup>646</sup> Dai L, Gao GF. *Viral targets for vaccines against COVID-19*. Nature Reviews in Immunology. 2021 February; 21(2):73-82. DOI: 10.1038/s41577-020-00480-0. Epub 2020 December 18.  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7747004/pdf/41577\\_2020\\_Article\\_480.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7747004/pdf/41577_2020_Article_480.pdf)
- <sup>647</sup> Hsieh CL, Goldsmith JA, Schaub JM, DiVenere AM, Kuo HC, Javanmardi K, Le KC, Wrapp D, Lee AG, Liu Y, Chou CW, Byrne PO, Hjorth CK, Johnson NV, Ludes-Meyers J, Nguyen AW, Park J, Wang N, Amengor D, Lavinder JJ, Ippolito GC, Maynard JA, Finkelstein IJ, McLellan JS. *Structure-based design of prefusion-stabilized SARS-CoV-2 spikes*. Science. 2020 September 18; 369(6510):1501-1505. DOI: 10.1126/science.abd0826. Epub 2020 July 23.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7402631/pdf/abd0826.pdf>
- <sup>648</sup> Battles MB, Langedijk JP, Furmanova-Hollenstein P, Chaiwatpongsakorn S, Costello HM, Kwanten L, Vranckx L, Vink P, Jaensch S, Jonckers TH, Koul A, Arnoult E, Peeples ME, Roymans D, McLellan JS. *Molecular mechanism of respiratory syncytial virus fusion inhibitors*. Nat Chem Biol. 2016 February; 12(2):87-93. DOI: 10.1038/nchembio.1982. Epub 2015 December 7.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4731865/pdf/nihms745245.pdf>
- <sup>649</sup> Robbins, Rebecca. “Moderna, Racing for Profits, Keeps Covid Vaccine Out of Reach of Poor.” *The New York Times*, 9 October 2021.  
<https://www.nytimes.com/2021/10/09/business/moderna-covid-vaccine.html>
- <sup>650</sup> Gertrude B. Elion. Nobel Prize in Physiology or Medicine. 1988.  
<https://www.nobelprize.org/womenwhochangedscience/stories/gertrude-elion>
- <sup>651</sup> Elion GB, Furman PA, Fyfe JA, de Miranda P, Beauchamp L, Schaeffer HJ. *Selectivity of action of an antihypertensive agent, 9-(2-hydroxyethoxymethyl) guanine*. Proceedings of the National Academy of Sciences (USA) 1977 December; 74(12):5716-20. DOI: 10.1073/pnas.74.12.5716  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC431864/pdf/pnas00043-0524.pdf>
- <sup>652</sup> Institute of Medicine 1995. *Review of the Fialuridine (FIAU) Clinical Trials*. Washington, DC. NAP. The National Academies Press. <https://doi.org/10.17226/4887>  
[https://www.nap.edu/cart/download.cgi?record\\_id=4887](https://www.nap.edu/cart/download.cgi?record_id=4887)
- <sup>653</sup> Homi Kharas and Meagan Dooley. “Long-Run Impacts of COVID-19 on Extreme Poverty.” *Brookings*, 2 June 2021. <https://www.brookings.edu/blog/future-development/2021/06/02/long-run-impacts-of-covid-19-on-extreme-poverty/>
- <sup>654</sup> Sheahan TP, Sims AC, Zhou S, Graham RL, Pruijssers AJ, Agostini ML, Leist SR, Schäfer A, Dinnon KH 3rd, Stevens LJ, Chappell JD, Lu X, Hughes TM, George AS, Hill CS, Montgomery SA, Brown AJ, Bluemling GR, Natchus MG, Saindane M, Kolykhalov AA, Painter G, Harcourt J, Tamin A, Thornburg NJ, Swanstrom R, Denison MR, Baric RS. *An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 in human airway epithelial cell cultures and multiple coronaviruses in mice*. Science Transl Med. 2020 April 29; 12(541):eabb5883. doi: 10.1126/scitranslmed.abb5883. Epub 2020 April 6.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7164393/pdf/abb5883.pdf>
- <sup>655</sup> Willyard C. *How antiviral pill molnupiravir shot ahead in the COVID drug hunt*. Nature. 2021 Oct 8. DOI: 10.1038/d41586-021-02783-1



- 
- <sup>656</sup> Bhagwati, Jagdish N. *In Defense of Globalization*. Oxford University Press, 2007. ISBN-13: 978-0195330939 [http://www.columbia.edu/~jb38/books/pdf/idg\\_reviews.pdf](http://www.columbia.edu/~jb38/books/pdf/idg_reviews.pdf)
- <sup>657</sup> Stiglitz, Joseph E. *Globalization and Its Discontents*. W.W. Norton, 2003. ISBN-13: 978-0393324396 [https://wcfia.harvard.edu/files/wcfia/files/570\\_stiglitzrev.pdf](https://wcfia.harvard.edu/files/wcfia/files/570_stiglitzrev.pdf)
- <sup>658</sup> Shanmugaraj B, I Bulaon CJ, Phoolcharoen W. *Plant Molecular Farming: A Viable Platform for Recombinant Biopharmaceutical Production*. *Plants* (Basel). 2020 July 4; 9(7):842. DOI:10.3390/plants9070842 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7411908/pdf/plants-09-00842.pdf>
- <sup>659</sup> Tulip and Economic History – see “NOTES” on the page preceding “ACKNOWLEDGEMENTS”
- <sup>660</sup> Hiatt A, Cafferkey R, Bowdish K. *Production of antibodies in transgenic plants*. *Nature*. 1989 Nov 2; 342(6245):76-78. DOI: 10.1038/342076a0
- <sup>661</sup> James E, Lee JM. *The production of foreign proteins from genetically modified plant cells*. *Advances in Biochemical Engineering and Biotechnology*. 2001. 72:127-156. DOI: 10.1007/3-540-45302-4\_5
- <sup>662</sup> Taverne, Dick. *The March of Unreason: Science, Democracy, and the New Fundamentalism*. Oxford University Press, 2006. ISBN-13: 978-0199205622 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC558032/pdf/bmj33001214.pdf>
- <sup>663</sup> Kupferschmidt, Kai (9 August 2013) *Activists Destroy “Golden Rice” Field Trial*. <https://www.science.org/content/article/activists-destroy-golden-rice-field-trial>
- <sup>664</sup> Lynas, Mark. “Anti-GMO Activists Lie About Attack on Rice Crop (and About So Many Other Things).” *Slate Magazine*, 26 August 2013. <https://slate.com/technology/2013/08/golden-rice-attack-in-philippines-anti-gmo-activists-lie-about-protest-and-safety.html>
- <sup>665</sup> Republic of the Philippines SUPREME COURT Manila G.R. No. 193459 February 15, 2011. <https://www.scribd.com/document/437684421/8-Gutierrez-v-HoR-on-Justice-pdf>
- <sup>666</sup> Feroze KB, Kaufman EJ. *Xerophthalmia*. [Updated 2021 April 25] StatPearls Publishing; 2021 January. <https://www.ncbi.nlm.nih.gov/books/NBK431094/>
- <sup>667</sup> Burkhardt PK, Beyer P, Wünn J, Klöti A, Armstrong GA, Schledz M, von Lintig J, Potrykus I. *Transgenic rice (Oryza sativa) endosperm expressing daffodil (Narcissus pseudonarcissus) phytoene synthase accumulates phytoene, a key intermediate of provitamin A biosynthesis*. *Plant Journal* 1997 May; 11(5):1071-1078. DOI: 10.1046/j.1365-313x.1997.11051071.x <https://onlinelibrary.wiley.com/doi/epdf/10.1046/j.1365-313X.1997.11051071.x>
- <sup>668</sup> *Golden Rice - The Embryo Project Encyclopedia*. <https://embryo.asu.edu/pages/golden-rice>
- <sup>669</sup> Beyer P, Al-Babili S, Ye X, Lucca P, Schaub P, Welsch R, Potrykus I. *Golden Rice: introducing the beta-carotene biosynthesis pathway into rice endosperm by genetic engineering to defeat vitamin A deficiency*. *Journal of Nutrition*, 2002 March; 132(3):506S-510S. DOI: 10.1093/jn/132.3.506S <https://academic.oup.com/jn/article/132/3/506S/4687202>
- <sup>670</sup> Taverne, D. Suppressing science. *Nature* 453, 857–858 (2008). <https://doi.org/10.1038/453857b> <https://www.nature.com/articles/453857b.pdf>

- <sup>671</sup> Philippines Approves Commercial Use of Genetically Engineered Rice. *Reuters*, 25 August 2021. <https://www.reuters.com/article/uk-philippines-rice-gmo-idUSKBN2FQ1D9>
- <sup>672</sup> Siddiqui, M. S. Approval of Golden Rice for Production and Consumption. *The Asian Age*. <http://dailyasianage.com/news/220741/?regenerate>
- <sup>673</sup> Pezzotti G, Zhu W, Chikaguchi H, Marin E, Boschetto F, Masumura T, Sato YI, Nakazaki T. Raman *Molecular Fingerprints of Rice Nutritional Quality and the Concept of Raman Barcode*. *Frontiers of Nutrition*, 2021 June 23; 8:663569. DOI: 10.3389/fnut.2021.663569 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8260989/pdf/fnut-08-663569.pdf>
- <sup>674</sup> Mason HS, Lam DM, Arntzen CJ. *Expression of hepatitis B surface antigen in transgenic plants*. *Proc Natl Acad Sci U S A*. 1992 December 15; 89(24):11745-11749. DOI: 10.1073/pnas.89.24.11745 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC50633/pdf/pnas01098-0106.pdf>
- <sup>675</sup> Thanavala Y, Yang YF, Lyons P, Mason HS, Arntzen C. *Immunogenicity of transgenic plant-derived hepatitis B surface antigen*. *Proc Natl Acad Sci U S A*. 1995 April 11; 92(8):3358-3361. DOI: 10.1073/pnas.92.8.3358 [www.ncbi.nlm.nih.gov/pmc/articles/PMC42165/pdf/pnas01492-0291.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC42165/pdf/pnas01492-0291.pdf)
- <sup>676</sup> Richter LJ, Thanavala Y, Arntzen CJ, Mason HS. *Production of hepatitis B surface antigen in transgenic plants for oral immunization*. *Nature Biotechnology*. 2000 November; 18(11):1167-1171. DOI: 10.1038/81153
- <sup>677</sup> Kapusta J, Modelska A, Figlerowicz M, Pniewski T, Letellier M, Lisowa O, Yusibov V, Koprowski H, Plucienniczak A, Legocki AB. *A plant-derived edible vaccine against hepatitis B virus*. *FASEB Journal*. 1999 October; 13(13):1796-9. DOI: 10.1096/fasebj.13.13.1796. Erratum: *FASEB J* 1999 Dec; 13(15):2339. <https://faseb.onlinelibrary.wiley.com/doi/epdf/10.1096/fasebj.13.13.1796>
- <sup>678</sup> Kong Q, Richter L, Yang YF, Arntzen CJ, Mason HS, Thanavala Y. *Oral immunization with hepatitis B surface antigen expressed in transgenic plants*. *Proc Natl Acad Sci U S A*. 2001 September 25; 98(20):11539-11544. DOI: 10.1073/pnas.191617598. Epub 2001 September 11. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC58765/pdf/pq011539.pdf>
- <sup>679</sup> Liang TJ. *Hepatitis B: the virus and disease*. *Hepatology*. 2009 May; 49(5 Suppl):S13-21. DOI: 10.1002/hep.22881 [www.ncbi.nlm.nih.gov/pmc/articles/PMC2809016/pdf/nihms164711.pdf](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2809016/pdf/nihms164711.pdf)
- <sup>680</sup> Mason HS, Ball JM, Shi JJ, Jiang X, Estes MK, Arntzen CJ. *Expression of Norwalk virus capsid protein in transgenic tobacco and potato and its oral immunogenicity in mice*. *Proc Natl Acad Sci U S A*. 1996 May 28; 93(11):5335-5340. DOI: 10.1073/pnas.93.11.5335 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC39246/pdf/pnas01512-0172.pdf>
- <sup>681</sup> Biemelt S, Sonnewald U, Galmbacher P, Willmitzer L, Müller M. *Production of human papillomavirus type 16 virus-like particles in transgenic plants*. *Journal of Virology*. 2003 September; 77(17):9211-9220. DOI: 10.1128/jvi.77.17.9211-9220.2003 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC187377/pdf/0131.pdf>
- <sup>682</sup> Warzecha H, Mason HS, Lane C, Tryggvesson A, Rybicki E, Williamson AL, Clements JD, Rose RC. *Oral immunogenicity of human papillomavirus-like particles expressed in potato*. *J Virol*. 2003 August; 77(16):8702-8711. DOI: 10.1128/jvi.77.16.8702-8711.2003 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC167207/pdf/0052.pdf>

- <sup>683</sup> Greco R, Michel M, Guetard D, Cervantes-Gonzalez M, Pelucchi N, Wain-Hobson S, Sala F, Sala M. *Production of recombinant HIV-1/HBV virus-like particles in Nicotiana tabacum and Arabidopsis thaliana plants for a bivalent plant-based vaccine*. *Vaccine*. 2007 November 28; 25(49):8228-40. DOI: 10.1016/j.vaccine.2007.09.061. Epub 2007 October 16.
- <sup>684</sup> Bright RA, Carter DM, Daniluk S, Toapanta FR, Ahmad A, Gavrilov V, Massare M, Pushko P, Mytle N, Rowe T, Smith G, Ross TM. *Influenza virus-like particles elicit broader immune responses than whole virion inactivated influenza virus or recombinant hemagglutinin*. *Vaccine*. 2007 May 10; 25(19):3871-78. DOI: 10.1016/j.vaccine.2007.01.106. Epub 2007 February 15.  
<https://onlinelibrary.wiley.com/doi/epdf/10.1111/j.1467-7652.2008.00384.x>
- <sup>685</sup> D'Aoust MA, Couture MM, Charland N, Trépanier S, Landry N, Ors F, Vézina LP. *The production of hemagglutinin-based virus-like particles in plants: a rapid, efficient and safe response to pandemic influenza*. *Plant Biotechnol J*. 2010 Jun; 8(5):607-19. DOI: 10.1111/j.1467-7652.2009.00496.x Epub 2010 February 18. <https://onlinelibrary.wiley.com/doi/epdf/10.1111/j.1467-7652.2009.00496.x>
- <sup>686</sup> Makarkov AI, Golizeh M, Ruiz-Lancheros E, Gopal AA, Costas-Cancelas IN, Chierzi S, Pillet S, Charland N, Landry N, Rouiller I, Wiseman PW, Ndao M, Ward BJ. *Plant-derived virus-like particle vaccines drive cross-presentation of influenza A hemagglutinin peptides by human monocyte-derived macrophages*. *NPJ Vaccines*. 2019 May 15; 4:17. DOI: 10.1038/s41541-019-0111-y  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6520342/pdf/41541\\_2019\\_Article\\_111.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6520342/pdf/41541_2019_Article_111.pdf)
- <sup>687</sup> Ward BJ, Gobeil P, Séguin A, Atkins J, Boulay I, Charbonneau PY, Couture M, D'Aoust MA, Dhaliwall J, Finkle C, Hager K, Mahmood A, Makarkov A, Cheng MP, Pillet S, Schimke P, St-Martin S, Trépanier S, Landry N. *Phase 1 randomized trial of a plant-derived virus-like particle vaccine for COVID-19*. *Nature Medicine* 2021 June; 27(6):1071-1078. DOI: 10.1038/s41591-021-01370-1 Epub 2021 May 18  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8205852/pdf/41591\\_2021\\_Article\\_1370.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8205852/pdf/41591_2021_Article_1370.pdf)
- <sup>688</sup> Mahmood N, Nasir SB, Hefferon K. *Plant-Based Drugs and Vaccines for COVID-19*. *Vaccines (Basel)*. 2020 December 30; 9(1):15. doi: 10.3390/vaccines9010015  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7823519/pdf/vaccines-09-00015.pdf>
- <sup>689</sup> Datta, Shoumen (2022) *Vaccines: Health of Nations*. <https://dspace.mit.edu/handle/1721.1/145774>  
PLANT BIOLOGICS (anti-virals; select papers 1989-2021) – <https://bit.ly/PLANT-BIOLOGICS>
- <sup>690</sup> Elbeaino, Toufic, *et al* “ICTV Virus Taxonomy Profile: Fimoviridae.” *Journal of General Virology*, vol. 99, no. 11, November 2018, pp. 1478–1479. <https://doi.org/10.1099/jgv.0.001143>
- <sup>691</sup> Wylie SJ, Adams M, Chalam C, Kreuze J, López-Moya JJ, Ohshima K, Praveen S, Rabenstein F, Stenger D, Wang A, Zerbini FM, ICTV Report Consortium. *ICTV Virus Taxonomy Profile: Potyviridae*. *Journal of General Virology* 2017 March; 98(3):352-354. DOI: 10.1099/jgv.0.000740  
Erratum: *J Gen Virol*. 2017 November; 98(11):2893  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5797945/pdf/jgv-98-352.pdf>
- <sup>692</sup> Lesnaw JA, Ghabrial SA. *Tulip Breaking: Past, Present, and Future*. *Plant Diseases* 2000 October; 84(10):1052-1060. DOI: 10.1094/PDIS.2000.84.10.1052  
<https://apsjournals.apsnet.org/doi/pdf/10.1094/PDIS.2000.84.10.1052>

- 
- <sup>693</sup> Verchot J, Herath V, Urrutia CD, Gayral M, Lyle K, Shires MK, Ong K, Byrne D. *Development of a Reverse Genetic System for Studying Rose Rosette Virus in Whole Plants*. *Mol Plant Microbe Interact*. 2020 October; 33(10):1209-1221. DOI: 10.1094/MPMI-04-20-0094-R. Epub 2020 August 20.  
<https://apsjournals.apsnet.org/doi/pdf/10.1094/MPMI-04-20-0094-R>
- <sup>694</sup> Mollov D, Lockhart B, Zlesak D. *Complete nucleotide sequence of rose yellow mosaic virus, a novel member of the family Potyviridae*. *Archives of Virology*. 2013 September; 158(9):1917-23.  
DOI: 10.1007/s00705-013-1686-7. Epub 2013 April 4
- <sup>695</sup> Diseases You Almost Forgot About (Thanks to Vaccines)  
<https://www.cdc.gov/vaccines/parents/diseases/forgot-14-diseases.html>
- <sup>696</sup> Tan J, Zhou Z, Niu Y, Sun X, Deng Z. *Identification and Functional Characterization of Tomato CircRNAs Derived from Genes Involved in Fruit Pigment Accumulation*. *Science Reports* 2017 August 17; 7(1):8594. DOI: 10.1038/s41598-017-08806-0  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5561264/pdf/41598\\_2017\\_Article\\_8806.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5561264/pdf/41598_2017_Article_8806.pdf)
- <sup>697</sup> Alzohairy MA. *Therapeutics Role of Azadirachta indica (Neem) and Their Active Constituents in Diseases Prevention and Treatment*. *Evid Based Complement Alternat Med*. 2016; 2016:7382506.  
DOI: 10.1155/2016/7382506. Epub 2016 March 1.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4791507/pdf/ECAM2016-7382506.pdf>
- <sup>698</sup> Abbasi J. *Moderna's mRNA Vaccine for Seasonal Flu Enters Clinical Trials*. *JAMA*. 2021 October 12; 326(14):1365. DOI: 10.1001/jama.2021.17499  
<https://jamanetwork.com/journals/jama/fullarticle/2784977>
- <sup>699</sup> Usha R, Rohll JB, Spall VE, Shanks M, Maule AJ, Johnson JE, Lomonosoff GP. *Expression of an animal virus antigenic site on the surface of a plant virus particle*. *Virology*. 1993 Nov; 197(1):366-74.  
DOI: 10.1006/viro.1993.1598
- <sup>700</sup> Cai Q, Qiao L, Wang M, He B, Lin FM, Palmquist J, Huang SD, Jin H. *Plants send small RNAs in extracellular vesicles to fungal pathogen to silence virulence genes*. *Science* 2018 June 8; 360(6393):1126-9.  
DOI: 10.1126/science.aar4142. Epub 2018 May 17.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6442475/pdf/nihms-1019813.pdf>
- <sup>701</sup> Cai Q, He B, Weiberg A, Buck AH, Jin H. *Small RNAs and extracellular vesicles: New mechanisms of cross-species communication and innovative tools for disease control*. *PLoS Pathog*. 2019 December 30; 15(12):e1008090. DOI: 10.1371/journal.ppat.1008090  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6936782/pdf/ppat.1008090.pdf>
- <sup>702</sup> Oliphant AR, Struhl K. *An efficient method for generating proteins with altered enzymatic properties: application to beta-lactamase*. *Proc Natl Acad Sci U S A*. 1989 December; 86(23):9094-9098.  
DOI: 10.1073/pnas.86.23.9094. Erratum: *Proc Natl Acad Sci U S A* 1992 May 15; 89(10):4779.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC298440/pdf/pnas00290-0052.pdf>
- <sup>703</sup> National Research Council. 2014. *Convergence: Facilitating Transdisciplinary Integration of Life Sciences, Physical Sciences, Engineering, and Beyond*. Washington, DC: The National Academies Press.  
<https://doi.org/10.17226/18722> & <https://www.nap.edu/download/18722>

- 
- <sup>704</sup> Tan J, Zhou Z, Niu Y, Sun X, Deng Z. *Identification and Functional Characterization of Tomato CircRNAs Derived from Genes Involved in Fruit Pigment Accumulation*. *Sci Rep*. 2017 Aug 17; 7(1):8594. DOI: 10.1038/s41598-017-08806-0  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5561264/pdf/41598\\_2017\\_Article\\_8806.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5561264/pdf/41598_2017_Article_8806.pdf)
- <sup>705</sup> Fan J, Quan W, Li GB, Hu XH, Wang Q, Wang H, Li XP, Luo X, Feng Q, Hu ZJ, Feng H, Pu M, Zhao JQ, Huang YY, Li Y, Zhang Y, Wang WM. *circRNAs Are Involved in the Rice-Magnaporthe oryzae Interaction*. *Plant Physiol*. 2020 January; 182(1):272-286. doi: 10.1104/pp.19.00716. Epub 2019 Oct 18.  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6945833/pdf/PP\\_201900716R1.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6945833/pdf/PP_201900716R1.pdf)
- <sup>706</sup> Beier JC. *Malaria parasite development in mosquitoes*. *Annual Review of Entomology* 1998; 43:519-43. DOI: 10.1146/annurev.ento.43.1.519
- <sup>707</sup> *Ronald Ross Explains Malaria*. <https://cosmosmagazine.com/science/ronald-ross-explains-malaria/>
- <sup>708</sup> Sullivan, David. (2006) *Malariology Overview*. Johns Hopkins University, Johns Hopkins Bloomberg School of Public Health. <https://ocw.jhsph.edu/courses/Malariology/PDFs/lecture1.pdf>
- <sup>709</sup> Ross R. *On some Peculiar Pigmented Cells Found in Two Mosquitos Fed on Malarial Blood*. *British Medical Journal*. 1897 December 18; 2(1929):1786-1788. DOI: 10.1136/bmj.2.1929.1786  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2408186/pdf/brmedj08748-0014.pdf>
- <sup>710</sup> CDC-Centers for Disease Control and Prevention. *CDC - Malaria - About Malaria - History - Ross and the Discovery That Mosquitoes Transmit Malaria Parasites*. 28 March 2017.  
<https://www.cdc.gov/malaria/about/history/ross.html>
- <sup>711</sup> The Nobel Prize in Physiology or Medicine 1902.  
<https://www.nobelprize.org/prizes/medicine/1902/summary/>
- <sup>712</sup> CDC-Centers for Disease Control and Prevention. *CDC - Malaria - About Malaria - History - Laveran and the Discovery of the Malaria Parasite*. 28 March 2017.  
<https://www.cdc.gov/malaria/about/history/laveran.html>
- <sup>713</sup> The Nobel Prize in Physiology or Medicine 1907.”  
<https://www.nobelprize.org/prizes/medicine/1907/summary/>
- <sup>714</sup> White, Nicholas J. “A Vaccine for Malaria.” *New England Journal of Medicine*, vol. 365, no. 20, November 2011, pp. 1926–1927. <https://doi.org/10.1056/NEJMe1111777>  
<https://www.nejm.org/doi/pdf/10.1056/NEJMe1111777?articleTools=true>
- <sup>715</sup> Duffy PE, Patrick Gorres J. *Malaria vaccines since 2000: progress, priorities, products*. *NPJ Vaccines*. 2020 June 9; 5(1):48. DOI: 10.1038/s41541-020-0196-3  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7283239/pdf/41541\\_2020\\_Article\\_196.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7283239/pdf/41541_2020_Article_196.pdf)



<sup>716</sup> RTS,S Clinical Trials Partnership, Agnandji ST, Lell B, Fernandes JF, Abossolo BP, Methogo BG, Kabwende AL, Adegnika AA, Mordmüller B, Issifou S, Kremsner PG, Sacarlal J, Aide P, Lanaspa M, Aponte JJ, Machevo S, Acacio S, Bulo H, Sigauque B, Macete E, Alonso P, Abdulla S, Salim N, Minja R, Mpina M, Ahmed S, Ali AM, Mtoro AT, Hamad AS, Mutani P, Tanner M, Tinto H, D'Alessandro U, Sorgho H, Valea I, Bihoun B, Guiraud I, Kaboré B, Sombié O, Guiguemdé RT, Ouédraogo JB, Hamel MJ, Kariuki S, Oneko M, Odero C, Otieno K, Awino N, McMorro M, Muturi-Kioi V, Laserson KF, Slutsker L, Otieno W, Otieno L, Otsyula N, Gondi S, Otieno A, Owira V, Oguk E, Odongo G, Woods JB, Ogutu B, Njuguna P, Chilengi R, Akoo P, Kerubo C, Maingi C, Lang T, Olotu A, Bejon P, Marsh K, Mwambingu G, Owusu-Agyei S, Asante KP, Osei-Kwakye K, Boahen O, Dosoo D, Asante I, Adjei G, Kwara E, Chandramohan D, Greenwood B, Lusingu J, Gesase S, Malabeja A, Abdul O, Mahende C, Liheluka E, Malle L, Lemnge M, Theander TG, Drakeley C, Ansong D, Agbenyega T, Adjei S, Boateng HO, Rettig T, Bawa J, Sylverken J, Sambian D, Sarfo A, Agyekum A, Martinson F, Hoffman I, Mvalo T, Kamthunzi P, Nkomo R, Tembo T, Tegha G, Tsidya M, Kilembe J, Chawinga C, Ballou WR, Cohen J, Guerra Y, Jongert E, Lapierre D, Leach A, Lievens M, Ofori-Anyinam O, Olivier A, Vekemans J, Carter T, Kaslow D, Leboulleux D, Loucq C, Radford A, Savarese B, Schellenberg D, Sillman M, Vansadia P. *A phase 3 trial of RTS,S/AS01 malaria vaccine in African infants*. *New England Journal of Medicine*. 2012 December 13; 367(24):2284-95. DOI: 10.1056/NEJMoa1208394. Epub 2012 November 9.

<https://www.nejm.org/doi/pdf/10.1056/NEJMoa1208394?articleTools=true>

<sup>717</sup> Laurens MB. *RTS,S/AS01 vaccine (Mosquirix™): overview*. *Hum Vaccin Immunother*. 2020 March 3; 16(3):480-489. DOI: 10.1080/21645515.2019.1669415. Epub 2019 October 22.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7227679/pdf/khvi-16-03-1669415.pdf>

<sup>718</sup> Mandavilli, Apoorva. “A ‘Historic Event’: First Malaria Vaccine Approved by W.H.O.” *The New York Times*, 6 October 2021. <https://www.nytimes.com/2021/10/06/health/malaria-vaccine-who.html>

<sup>719</sup> Gouglas D, Thanh Le T, Henderson K, Kaloudis A, Danielsen T, Hammersland NC, Robinson JM, Heaton PM, Røttingen JA. *Estimating the cost of vaccine development against epidemic infectious diseases: a cost minimisation study*. *Lancet Global Health*. 2018 December; 6(12):e1386-e1396.

DOI: 10.1016/S2214-109X(18)30346-2. Epub 2018 October 18.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7164811/pdf/main.pdf>

<sup>720</sup> *Vaccine Infrastructure And Education Is The Best Medical Investment Our Country Can Make*. Health Affairs Blog. <https://www.healthaffairs.org/doi/10.1377/hblog20200117.291021/full/>

<sup>721</sup> WHO International. *Some Vaccine Costs Are Hidden Below The Surface*.

[www.who.int/immunization/programmes\\_systems/supply\\_chain/optimize/immunization\\_costs.pdf](http://www.who.int/immunization/programmes_systems/supply_chain/optimize/immunization_costs.pdf)

<sup>722</sup> U.S. Department of Health and Human Services. 2021. *Vaccines National Strategic Plan 2021–2025*. Washington, DC. <https://www.hhs.gov/sites/default/files/HHS-Vaccines-Report.pdf>

<sup>723</sup> The Calcutta School of Tropical Medicine and Hygiene. *Nature* 111, 550–551 (1923).

<https://doi.org/10.1038/111550b0> & <https://www.nature.com/articles/111550b0.pdf>

<sup>724</sup> Bhattacharya, A. K. (1989) *Calcutta School of Tropical Medicine*. *National Medical Journal Of India* Vol.2, No.1. <https://archive.nmji.in/approval/archive/Volume-2/issue-1/indian-medical-colleges.pdf>

- 
- <sup>725</sup> Power H. *The Calcutta School of Tropical Medicine: institutionalizing medical research in the periphery*. *Med Hist*. 1996 April; 40(2):197-214. DOI: 10.1017/s0025727300061007  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1037095/pdf/medhist00030-0063.pdf>
- <sup>726</sup> *Malaria Milestones*. <https://kids.wng.org/node/3583>
- <sup>727</sup> Waltz, Emily. “First Genetically Modified Mosquitoes Released in the United States.” *Nature*, vol. 593, no. 7858, May 2021, pp. 175–176. <https://doi.org/10.1038/d41586-021-01186-6>  
<https://media.nature.com/original/magazine-assets/d41586-021-01186-6/d41586-021-01186-6.pdf>
- <sup>728</sup> Coffey, Donavyn. “First Genetically Modified Mosquitoes Released in U.S. Are Hatching Now.” *Scientific American*, <https://www.scientificamerican.com/article/first-genetically-modified-mosquitoes-released-in-u-s-are-hatching-now/>
- <sup>729</sup> Powell JR. *An Evolutionary Perspective on Vector-Borne Diseases*. *Front Genet*. 2019 December 17; 10:1266. DOI: 10.3389/fgene.2019.01266  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6929172/pdf/fgene-10-01266.pdf>
- <sup>730</sup> Knols BGJ, Bossin HC, Mukabana WR, et al. *Transgenic Mosquitoes and the Fight against Malaria: Managing Technology Push in a Turbulent GMO World*. In: Breman JG, Alilio MS, White NJ, editors. *Defining and Defeating the Intolerable Burden of Malaria III: Progress and Perspectives: Supplement to Volume 77(6) of American Journal of Tropical Medicine and Hygiene*. Northbrook (IL): American Society of Tropical Medicine and Hygiene; 2007 December.  
<https://www.ncbi.nlm.nih.gov/books/NBK1696/>  
[https://www.ncbi.nlm.nih.gov/books/NBK1696/pdf/Bookshelf\\_NBK1696.pdf](https://www.ncbi.nlm.nih.gov/books/NBK1696/pdf/Bookshelf_NBK1696.pdf)
- <sup>731</sup> Li M, Yang T, Bui M, Gamez S, Wise T, Kandul NP, Liu J, Alcantara L, Lee H, Edula JR, Raban R, Zhan Y, Wang Y, DeBeaubien N, Chen J, Sánchez C HM, Bennett JB, Antoshechkin I, Montell C, Marshall JM, Akbari OS. *Suppressing mosquito populations with precision guided sterile males*. *Nature Communications*. 2021 September 10; 12(1):5374. DOI: 10.1038/s41467-021-25421-w  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8433431/pdf/41467\\_2021\\_Article\\_25421.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8433431/pdf/41467_2021_Article_25421.pdf)
- <sup>732</sup> Nanfack Minkeu F, Vernick KD. *A Systematic Review of the Natural Virome of Anopheles Mosquitoes*. *Viruses*. 2018 April 25; 10(5):222. DOI: 10.3390/v10050222  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5977215/pdf/viruses-10-00222.pdf>
- <sup>733</sup> Powell JR. Mosquito-Borne Human Viral Diseases: Why *Aedes aegypti*? *American Journal of Tropical Medicine and Hygiene*. 2018 June; 98(6):1563-1565. DOI: 10.4269/ajtmh.17-0866. Epub 2018 March 15.  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6086192/pdf/tpmd170866.pdf>
- <sup>734</sup> Olmo RP, Ferreira AGA, Izidoro-Toledo TC, Aguiar ERGR, de Faria IJS, de Souza KPR, Osório KP, Kuhn L, Hammann P, de Andrade EG, Todjro YM, Rocha MN, Leite THJF, Amadou SCG, Armache JN, Paro S, de Oliveira CD, Carvalho FD, Moreira LA, Marois E, Imler JL, Marques JT. *Control of dengue virus in the midgut of Aedes aegypti by ectopic expression of the dsRNA-binding protein Loqs2*. *Nature Microbiology* 2018 December; 3(12):1385-1393. DOI: 10.1038/s41564-018-0268-6. Epub 2018 Oct 29.  
<https://www.nature.com/articles/s41564-018-0268-6>

<sup>735</sup> Wickenhagen A, Sugrue E, Lytras S, Kuchi S, Noerenberg M, Turnbull ML, Loney C, Herder V, Allan J, Jarmson I, Cameron-Ruiz N, Varjak M, Pinto RM, Lee JY, Iselin L, Palmalux N, Stewart DG, Swingler S, Greenwood EJD, Crozier TWM, Gu Q, Davies EL, Clohisey S, Wang B, Trindade Maranhão Costa F, Freire Santana M, de Lima Ferreira LC, Murphy L, Fawkes A, Meynert A, Grimes G; ISARIC4C Investigators, Da Silva Filho JL, Marti M, Hughes J, Stanton RJ, Wang ECY, Ho A, Davis I, Jarrett RF, Castello A, Robertson DL, Semple MG, Openshaw PJM, Palmarini M, Lehner PJ, Baillie JK, Rihn SJ, Wilson SJ. *A prenylated dsRNA sensor protects against severe COVID-19*. *Science*. 2021 October 29; 374(6567):eabj3624. DOI: 10.1126/science.abj3624. Epub 2021 October 29.

<https://eprints.whiterose.ac.uk/179044/1/A%20prenylated%20dsRNA%20sensor%20protects%20against%20severe%20COVID-19.pdf>

<sup>736</sup> Weiberg A, Wang M, Bellingier M, Jin H. *Small RNAs: a new paradigm in plant-microbe interactions*. *Annual Review of Phytopathology*. 2014; 52:495-516. DOI: 10.1146/annurev-phyto-102313-045933

<sup>737</sup> Huang CY, Wang H, Hu P, Hamby R, Jin H. *Small RNAs - Big Players in Plant-Microbe Interactions*. *Cell Host Microbe*. 2019 August 14; 26(2):173-182. DOI: 10.1016/j.chom.2019.07.021

<https://www.cell.com/action/showPdf?pii=S1931-3128%2819%2930365-8>

<sup>738</sup> Altae-Tran H, Kannan S, Demircioglu FE, Oshiro R, Nety SP, McKay LJ, Dlakić M, Inskeep WP, Makarova KS, Macrae RK, Koonin EV, Zhang F. *The widespread IS200/IS605 transposon family encodes diverse programmable RNA-guided endonucleases*. *Science*. 2021 October; 374(6563):57-65. DOI: 10.1126/science.abj6856. Epub 2021 September 9.

<sup>739</sup> Thélot FA, Zhang W, Song K, Xu C, Huang J, Liao M. *Distinct allosteric mechanisms of first-generation MsbA inhibitors*. *Science*. 2021 October 29; 374(6567):580-85. DOI: 10.1126/science.abi9009. Epub 2021 September 23. <https://www.science.org/doi/10.1126/science.abi9009>

<sup>740</sup> Fedorov NB, Benson LC, Graef J, Lippiello PM, Bencherif M. *Differential pharmacologies of mecamylamine enantiomers: positive allosteric modulation and noncompetitive inhibition*. *J Pharmacol Exp Ther*. 2009 February; 328(2):525-32. DOI: 10.1124/jpet.108.146910. Epub 2008 October 28. <https://jpet.aspetjournals.org/content/jpet/328/2/525.full.pdf>

<sup>741</sup> Laprairie RB, Kulkarni PM, Deschamps JR, Kelly MEM, Janero DR, Cascio MG, Stevenson LA, Pertwee RG, Kenakin TP, Denovan-Wright EM, Thakur GA. *Enantiospecific Allosteric Modulation of Cannabinoid 1 Receptor*. *ACS Chem Neuroscience*. 2017 June 21; 8(6):1188-1203. DOI: 10.1021/acscemneuro.6b00310. Epub 2017 February 7.

<https://pubs.acs.org/doi/pdf/10.1021/acscemneuro.6b00310>

<sup>742</sup> <https://www.bitesquad.com/>

<sup>743</sup> Bose, R., and P. K. Sarkar. *The Role Of Cycle Rickshaws - Low Cost Means Of Transport In Old Part Of A City*. 2002. <https://trid.trb.org/view/735172>

<sup>744</sup> Auerbach, Stuart. "From India: A Bullock Cart That Meets the Needs of Modern Times." *Washington Post*, 11 December 1981. <https://www.washingtonpost.com/archive/politics/1981/12/11/from-india-a-bullock-cart-that-meets-the-needs-of-modern-times/00d6a2a0-5902-4a31-8680-7644bcbab3cc/>

- 
- <sup>745</sup> Raj M. Desai, Homi Kharas, and Selen Özdoğan. “Over 1 Billion People Live in Poverty Hotspots.” *Brookings*, 2 December 2020.  
<https://www.brookings.edu/blog/future-development/2020/12/02/over-1-billion-people-live-in-poverty-hotspots/>
- <sup>746</sup> <https://xicanaphd.com/2017/08/24/if-its-inaccessible-to-the-poor-then-what-is-it/>
- <sup>747</sup> Tulipomania. [http://penelope.uchicago.edu/~grout/encyclopaedia\\_romana/aconite/tulipomania.html](http://penelope.uchicago.edu/~grout/encyclopaedia_romana/aconite/tulipomania.html)
- <sup>748</sup> Goldgar, Anne. *Tulipmania: Money, Honor, and Knowledge in the Dutch Golden Age*. University of Chicago Press, 2007. ISBN-13: 978-0226301266.  
<https://www.press.uchicago.edu/ucp/books/book/chicago/T/bo5414939.html>
- <sup>749</sup> Garber, Peter M. “Famous First Bubbles.” *Journal of Economic Perspectives*, vol. 4, no. 2, May 1990, pp. 35–54. <https://doi.org/10.1257/jep.4.2.35> & <https://pubs.aeaweb.org/doi/pdfplus/10.1257/jep.4.2.35>
- <sup>750</sup> Gebhardt, Andrew. *Holland Flowering: How the Dutch Flower Industry Conquered the World*. Amsterdam University Press, 2014. ISBN: 978-90-485-2259-0  
<https://doi.org/10.2307/j.ctt128783w> & <http://www.jstor.org/stable/j.ctt128783w>
- <sup>751</sup> Racaniello, Vincent (2012) *Tulips Broken by Viruses*.  
<http://www.virology.ws/2012/03/14/tulips-broken-by-viruses/>
- <sup>752</sup> Economic Theory of Networks. The Origin of Dutch Auction.  
<https://tuecontheoryofnetworks.wordpress.com/2013/02/25/the-origin-of-the-dutch-auction>
- <sup>753</sup> Inglis-Arkell, Esther (2021) “The Virus That Destroyed the Dutch Economy.” *Gizmodo*.  
<https://gizmodo.com/the-virus-that-destroyed-the-dutch-economy-5905247>
- <sup>754</sup> O’Sullivan, Feargus (2021) How Amsterdam Recovered From a Deadly Outbreak – in 1665.  
[www.bloomberg.com/news/features/2021-04-14/how-amsterdam-recovered-from-bubonic-plague](http://www.bloomberg.com/news/features/2021-04-14/how-amsterdam-recovered-from-bubonic-plague)
- <sup>755</sup> Brandes, J. and Wetter, C. *Classification of elongated plant viruses on the basis of particle morphology*. *Virology*. 1959 May; 8(1):99-115. DOI: 10.1016/0042-6822(59)90022-4  
[https://doi.org/10.1016/0042-6822\(59\)90022-4](https://doi.org/10.1016/0042-6822(59)90022-4)

**Cartoon for Reference 372** (additional discussion is here DT-2025 <https://dspace.mit.edu/handle/1721.1/155442>)

Machine Learning Algorithms – ML makes better sense without the misnomer of AI since there is no canonical ‘intelligence’ in ‘artificial intelligence’ or its “new” cronies (generative AI, ChatGPT, etc.). The term AI is an outcome of historical quagmire where human ego overtook science and facts. At best AI may be referred to as artificial reasoning tools (ART) but there is nothing about reasoning that is “artificial” because the logic and rules in *any analytics* approach or analytical technique is programmed by humans in the loop. Causality is not a part of AI. Determining cause/effect are not properties of AI or ML which makes it impossible to claim any degree of any form of cognition or cognitive ability. Hence, AI will remain a marketing miracle created as an egregious error due to human hubris. Mathematically, *analysis* is a term applied to calculus and all higher mathematics that uses calculus. Logic, rules and reasoning tools (LRRT) devoid of calculus is likely to be hand-waving subjectivity of limited value, if at all. Similarly, machine “learning” also takes “artistic” license by creating the illusion that machines are *learning*, when in reality machines apply stored logic and rules, programmed by humans, to data and information that is supplied by “schema” created by humans in the loop. Neither ML nor AI creates anything new or novel but uses programmed logic and rules in all possible and “allowed” permutations and combinations to data. There aren’t any “magic inside the black box” but “machinery” which *supplies correlations using correspondence rules (provided by equations) that govern function*. There is no “intelligence” or anything “artificial” because the *machinery* is the relation between variables determined by functions. Function is a relation between two variables which maps to values given by domain, range, Cartesian coordinates (x,y) or polar coordinates (r,θ). Values or sets of values and limits are deduced, derived, formulated and programmed by humans (algorithms) at the heart of the engine in any *learning machine*. Much to the chagrin of buzz-word peddlers (consulting firms) and marketing teams (sound bite manufacturers), the purpose of percolating the term “AI” is to deliberately distract us from facts and truth in order to catalyze collusive strategies for snake-oil sales. ML is tolerable but presents illusions of grandeur when *learning* refers to a mathematically informed ensemble of logic, rules and reasoning tools (LRRT). Pedantically speaking “LRRT” are *machinery applications* of logic (LO), rules and reasoning (RE) tools (TO). LRRT or LORETO are not glib and smug acronyms but conveys the unvarnished concept. In our world polishing the chrome is valued higher than tuning the engine. Efforts to reduce bias and increase credibility (what is truly deliverable), is an exercise in futility by scripturient fools (first author).

